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# Early growth phase and caffeine content response to recent and projected increases in atmospheric carbon dioxide in coffee (*Coffea arabica* and *C. canephora*)

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While [CO<sub>2</sub>] effects on growth and secondary chemistry are well characterized for annual plant species, little is known about perennials. Among perennials, production of *Coffea arabica* and *C. canephora* (robusta) have enormous economic importance worldwide. Three Arabica cultivars (Bourbon, Catimor, Typica) and robusta coffee were grown from germination to ca. 12 months at four CO<sub>2</sub> concentrations: 300, 400, 500 or 600 ppm. There were significant increases in all leaf area and biomass markers in response to [CO<sub>2</sub>] with significant [CO<sub>2</sub>] by taxa differences beginning at 122–124 days after sowing (DAS). At 366–368 DAS, CO<sub>2</sub> by cultivar variation in growth and biomass response among Arabica cultivars was not significant; however, significant trends in leaf area, branch number and total above-ground biomass were observed between Arabica and robusta. For caffeine concentration, there were significant differences in [CO<sub>2</sub>] response between Arabica and robusta. A reduction in caffeine in coffee leaves and seeds might result in decreased ability against deterrence, and consequently, an increase in pest pressure. We suggest that the interspecific differences observed (robusta vs. Arabica) may be due to differences in ploidy level (2n = 22 vs. 2n = 4x = 44). Differential quantitative and qualitative responses during early growth and development of Arabica and robusta may have already occurred with recent [CO<sub>2</sub>] increases, and such differences may be exacerbated, with production and quality consequences, as [CO<sub>2</sub>] continues to increase.

Because CO<sub>2</sub> represents the sole source of carbon for photosynthesis, and because CO<sub>2</sub> levels have been low for the recent geological past (<800,000 years before present), recent (317–412 ppm since 1960) and projected increases<sup>1</sup> (450–600 ppm by 2050) represent a major shift in an essential resource needed for plant growth. The biological role of rising atmospheric carbon dioxide concentration [CO<sub>2</sub>] is well recognized as altering physical (e.g., growth rates, stomatal aperture), biochemical (e.g., carbon to nitrogen (C:N) ratios, photorespiration), phenological (e.g., time to flowering), and reproductive (e.g., seed yield) characteristics for a wide variety of plant taxa, including agricultural crops<sup>2–6</sup>.

Because interspecific and intraspecific variation exists in response to resource changes, there has been a merited focus on quantifying intraspecific variation that could be used as a means of selection for adaptation to rising [CO<sub>2</sub>] levels. For example, studies have confirmed that there is significant intraspecific variation in the yield response to future CO<sub>2</sub> levels for cowpea (*Vigna unguiculata* (L.) Walp.)<sup>7</sup>; common bean (*Phaseolus vulgaris* L.)<sup>8</sup>,

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Parameter	[CO <sub>2</sub> ]	4T	A/R	[CO <sub>2</sub> ] × CV	[CO <sub>2</sub> ] × A/R	[CO <sub>2</sub> ] × 4T
<b>122–124 DAS</b>						
Leaf Area	***	***	**	0.15	0.68	0.47
Abv. Ground Wt.	*	***	***	(*)	0.12	**
<b>211–213 DAS</b>						
Leaf Area	***	***	***	0.44	0.35	0.27
Abv. Ground Wt.	***	***	***	0.73	*	0.21
<b>366–368 DAS</b>						
% Nitrogen	***	0.12	*	0.30	0.66	0.38
C:N	***	**	***	0.30	0.37	0.27
Caffeine (mg g <sup>-1</sup> )	*	*	*	0.27	*	0.26
Height (cm)	***	***	***	0.82	0.43	0.53
True Leaf No.	0.07	***	***	0.99	0.63	0.91
Branch No.	***	***	***	0.27	*	*
Leaf Area	**	***	***	0.89	0.20	0.60
Leaf Wt.	**	***	***	0.93	(*)	0.42
Branch Wt.	**	***	***	0.98	0.54	0.93
Stem Wt.	**	***	***	0.73	0.16	0.34
Total Wt.	***	***	***	0.97	(*)	0.57

**Table 1.** Statistical values for the three Arabica cultivars and robusta coffee response to recent and projected increases in atmospheric CO<sub>2</sub> at three sampling periods (DAS, days after sowing). A/R is Arabica vs. robusta; [CO<sub>2</sub>] × CV is CO<sub>2</sub> × Arabica cultivars only; [CO<sub>2</sub>] × 4T is [CO<sub>2</sub>] × all four taxa. Total above ground weight and vegetative characteristics are in g per plant. Leaf area is in cm<sup>2</sup>. (\*) Indicates a P value between 0.05 and 0.10; \* Indicates a P value between 0.05 and 0.01; \*\* Indicates a P value between 0.01 and 0.001; \*\*\* Indicates a P value < 0.001.

rice (*Oryza sativa* L.)<sup>9–11</sup>; wheat (*Triticum aestivum* L.)<sup>12,13</sup> and soybean (*Glycine max* (L.) Merr.)<sup>14</sup>, such that breeders could begin to select for CO<sub>2</sub> responsiveness among currently available germplasm.

However, such efforts have been focused, in general, to annual crops, particularly those of global importance (e.g., wheat, rice). Less attention, overall, has been given for CO<sub>2</sub> selection among perennial crops. In that regard, coffee (*Coffea arabica* L. (Arabica coffee) and *C. canephora* Pierre ex A. Froehner (robusta coffee)) is one of the world's most important perennial crops, and represents not only a widely traded agricultural commodity, but also a social and economic foundation for numerous tropical developing countries, with approximately 125 million people involved in coffee growing<sup>15</sup>. Although there are 124 *Coffea* species<sup>16</sup>, only two, Arabica and robusta are associated with the bulk of global coffee production<sup>17</sup>.

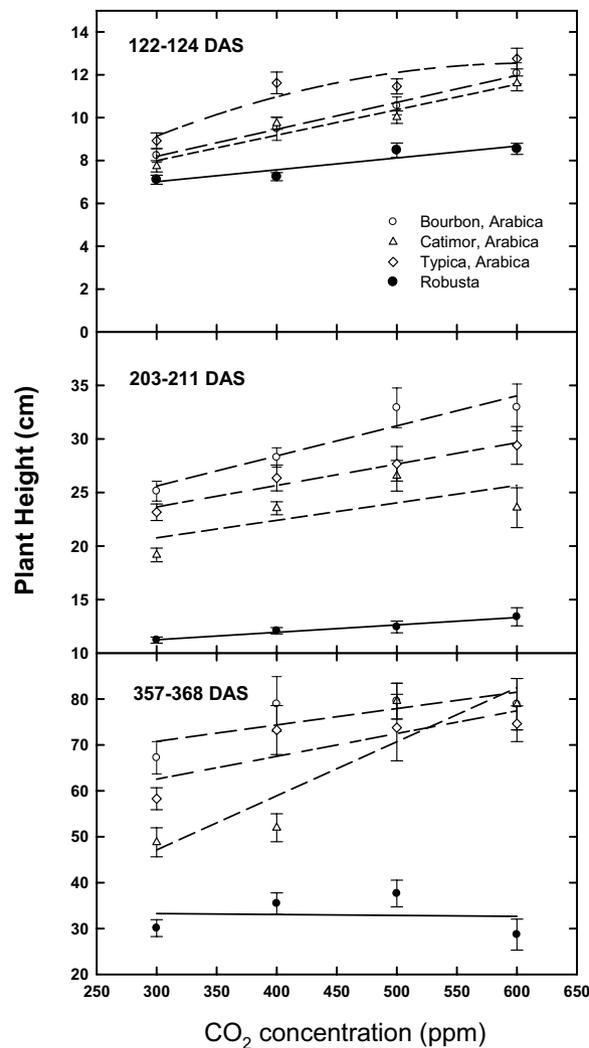
Arabica and robusta field responses to ca. 550 ppm CO<sub>2</sub>, with an emphasis on photosynthetic metabolism, is available<sup>18</sup>. Additional growth chamber studies evaluating temperature and [CO<sub>2</sub>] in the context of growth and photosynthetic acclimation response (including transformations in stomatal characteristics) are also available for coffee<sup>19–22</sup>. However, these data represent the growth and metabolic response of coffee following transfer of 12 to 18-month-old coffee plantlets into Free-Air CO<sub>2</sub> enrichment (FACE) or [CO<sub>2</sub>] growth chambers. At present, any differential growth response within, or between Arabica and robusta to recent and projected increases in CO<sub>2</sub> from germination through early growth (ca. 1 year) is not available. Yet, early exposure may be critical, as initial vegetative growth may represent the temporal period of greatest physiological sensitivity to additional CO<sub>2</sub>, for annuals<sup>23</sup>.

In addition to differential growth, there is substantial evidence that supplementary CO<sub>2</sub> may reduce protein content and increase carbon to nitrogen (C:N) ratios for numerous plant taxa<sup>4,24,25</sup> with potential effects on secondary compounds that have a high N content<sup>26</sup>. Caffeine (C<sub>8</sub>H<sub>10</sub>N<sub>4</sub>O<sub>2</sub>; 1,3,7-trimethylxanthine; ca. 29% N by molecular weight) may act as a defense against herbivores<sup>27–29</sup> and consequently, CO<sub>2</sub>-induced changes in leaf and seed caffeine concentration may be of ecological interest<sup>30</sup> including unforeseen consequences for climate change impact as a result of changes in plant-herbivore relationships<sup>31</sup>.

To determine the physiological impact of recent and projected increases in CO<sub>2</sub> levels four *Coffea* taxa, i.e., three Arabica cultivars (Bourbon, Catimor, Typica) and robusta coffee, were grown from germination for approximately one year at CO<sub>2</sub> concentrations of 300, 400, 500 or 600 ppm, and measured growth (plant height, leaf area, biomass, leaf weight, number of branches, dry weight), C: N ratio, and caffeine concentration (mg g<sup>-1</sup>).

## Results

Comparisons of plant height indicate significant increases at all sampling periods as a function of [CO<sub>2</sub>] above the 300 ppm baseline (Table 1; Fig. 1). However, by the 12-month period (357–368 days after sowing; DAS), there was no significant effect of [CO<sub>2</sub>] on plant height for robusta (Fig. 1). Similarly, [CO<sub>2</sub>] stimulation of leaf area was observed for all taxa at the 4 and 7-month period (122–124 and 203–211 DAS, respectively) in response to rising [CO<sub>2</sub>]; however, by the 12-month period, robusta plants had stopped responding (Fig. 2). Differences in leaf area as a function of [CO<sub>2</sub>] by Arabica/robusta were not significant (P = 0.20; Table 1).



**Figure 1.** Change in plant height (Average  $\pm$  SE) as a function of days after sowing (DAS) and  $[\text{CO}_2]$  for three Arabica cultivars and robusta coffee.

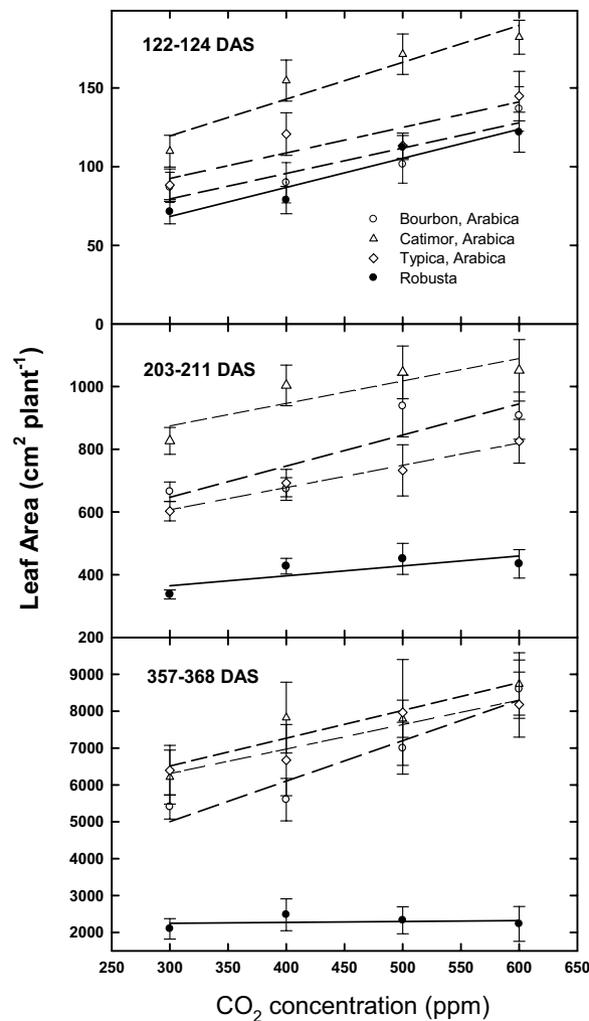
For above-ground plant biomass, increasing  $[\text{CO}_2]$  resulted in  $[\text{CO}_2]$  by Arabica cultivar responses at the 4-month period (122–124 DAS), but not at 7 (203–211 DAS) or 12 months (357–368) (Table 1; Fig. 3). By the end of the study (~12 months), no effect of  $[\text{CO}_2]$  was evident for robusta (Fig. 3); however, marginally significant differences ( $P < 0.1$ ) between Arabica and robusta for above ground dry weight were observed (Table 1; Fig. 4). Overall, by 12 months, Arabica, on average, showed a significant response to increasing  $[\text{CO}_2]$  for several vegetative parameters; whereas robusta was insensitive to  $[\text{CO}_2]$  for several vegetative parameters (Table 1, Fig. 4).

In addition to growth and vegetative response,  $[\text{CO}_2]$  induced changes in qualitative parameters, e.g., % N, carbon to nitrogen (C:N) ratio and caffeine concentration are of interest.

For the final harvest, when averaged for all taxa, significant effects were noted for C:N ratio for  $[\text{CO}_2]$  (Table 1, Fig. 5A), and for Arabica vs. robusta (Table 1, Fig. 5B). Differences for the Arabica cultivars were also noted for C:N and caffeine, but not for % N ( $P = 0.12$ ) (Table 1; Fig. 5C). Interactions,  $[\text{CO}_2] \times$  Arabica cultivars only, Arabica vs. robusta or cultivar (all four taxa) were not significant for % N or C:N ratio (Table 1). When averaged for all taxa, there were no significant differences in caffeine (Table 1, Fig. 6A), in contrast to a significant difference in reductions of caffeine with increasing  $[\text{CO}_2]$  for robusta but not Arabica (Table 1, Fig. 6B). No caffeine concentration ( $\text{mg g}^{-1}$ ) differences were observed among the three Arabica cultivars (Table 1, Fig. 6C),

## Discussion

Plant growth and development, assuming physiologically relevant temperatures, relies on four environmental (abiotic) resources: nutrients (macro- and micro-), light, water, and  $\text{CO}_2$ . Any change in one (or more) of these resources could lead to a change in fitness among different genotypes<sup>32</sup>. In managed plant systems, there have been numerous studies indicating intraspecific variation to  $[\text{CO}_2]$  with respect to vegetative and physiological characteristics, including yield, for a given crop species<sup>33</sup>. Sufficient variation has been reported so that screening or selecting for enhanced  $[\text{CO}_2]$  responsive cultivars offers a potential means to increase crop yields and improve nutrition, which are important steps to help adapt production to global climate change<sup>2,13,33,34</sup>.



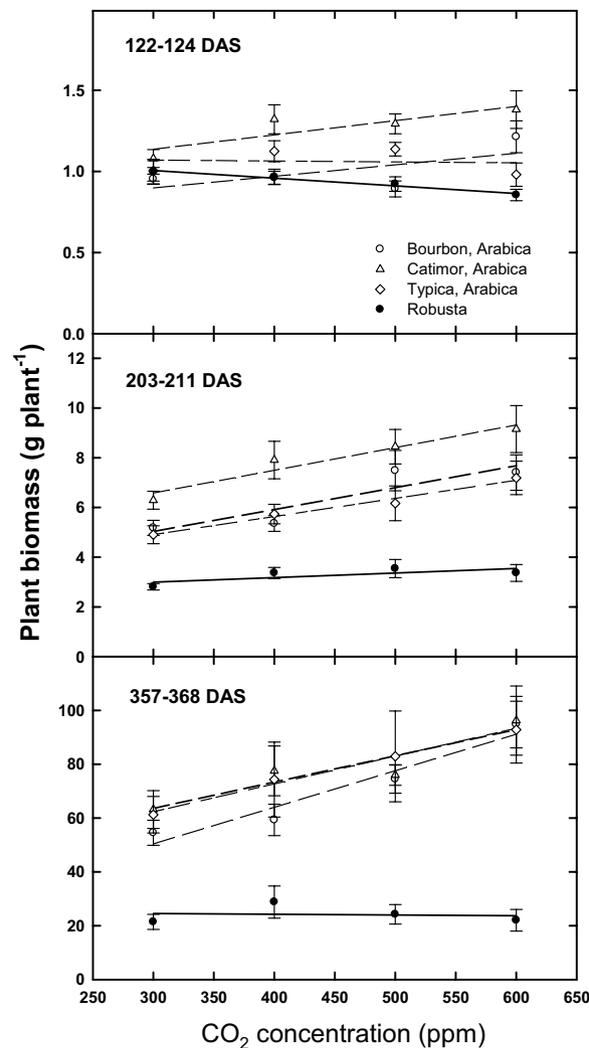
**Figure 2.** Change in leaf area (cm<sup>2</sup> per plant, average  $\pm$  SE) for three Arabica cultivars and robusta coffee at three different sampling times (days after sowing, DAS) in response to [CO<sub>2</sub>].

Efforts have been made to identify variation in productivity responses to elevated [CO<sub>2</sub>] for forest tree species<sup>35</sup>. Such studies have found considerable intraspecific variation in photosynthesis, stem biomass and volume for poplar, pine, birch, eucalyptus, etc., at elevated [CO<sub>2</sub>], suggesting that under non-limiting environmental conditions, (e.g., temperature, nutrients, water), intraspecific variation could be used to select for increased productivity as atmospheric CO<sub>2</sub> increases<sup>36,37</sup>. However, similar efforts for intraspecific or interspecific selection to [CO<sub>2</sub>] among tree crops (e.g., apples, cacao) are, at present, unavailable, despite experiments showing that trees can be more responsive than herbaceous plants to elevated CO<sub>2</sub><sup>34</sup>.

In the current study, while Arabica cultivars showed a significant response to rising [CO<sub>2</sub>] above the mid-20<sup>th</sup> century baseline (i.e., 317 ppm) for leaf area and growth parameters, significant variation among Arabica cultivars was not evident for any DAS harvest. In contrast, robusta coffee was consistently less responsive to rising [CO<sub>2</sub>] for growth biomass traits. Accordingly, there is a clear interspecific (between species) difference between Arabica and robusta to rising CO<sub>2</sub> with respect to the degree of [CO<sub>2</sub>] stimulation. Such divergence is evident in leaf weight, number of branches, and above ground biomass (Fig. 4).

In addition to differential growth response to rising [CO<sub>2</sub>], numerous reports have indicated CO<sub>2</sub> induced changes in secondary plant chemistry<sup>4</sup>. Of ubiquitous note in these observed changes is the CO<sub>2</sub> induced decline in protein and N, with subsequent increases in C:N ratio<sup>26</sup>. In the current study, similar N declines were observed, but no interspecific or intraspecific differences were recorded. However, caffeine concentration (mg g<sup>-1</sup>) when averaged for all Arabica cultivars and for robusta combined, declined with additional [CO<sub>2</sub>], and this decline was significantly more for robusta vs. Arabica. Whether such declines may improve or reduce beverage quality in the future remains to be determined.

If caffeine acts as a deterrent against herbivores<sup>27-29</sup>, a reduction in caffeine in coffee leaves and seeds might result in decreased ability against deterrence, and consequently, increase pest pressure on the plants. Even though the projected effects of climate change on the coffee berry borer (*Hypothenemus hampei*), coffee leaf miner (*Leucoptera coffeella*), coffee white stem borer (*Monochamus leonotus*), root-knot nematode (*Meloidogyne incognita*), and coffee leaf rust have been examined<sup>38</sup>, none of these studies considers possible changes in caffeine levels, and other chemistry, as a result of increasing CO<sub>2</sub> levels.

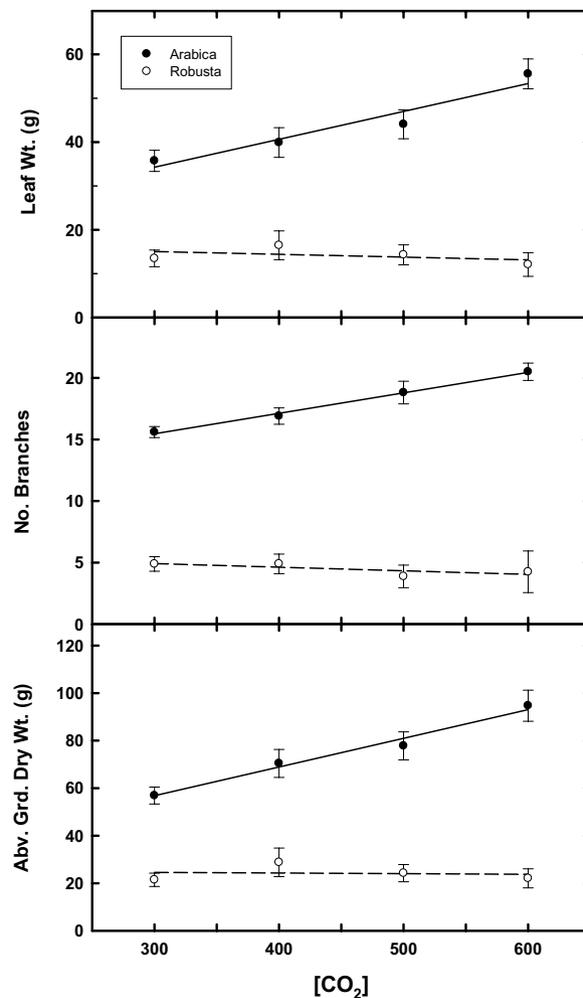


**Figure 3.** Change in total plant biomass (grams per plant, average  $\pm$  SE) for three Arabica cultivars and robusta coffee at three different sampling times (days after sowing, DAS) in response to  $[\text{CO}_2]$ .

The results presented here indicate no significant intraspecific variation in response to  $[\text{CO}_2]$  among Arabica cultivars and hence, no clear indication as to whether recent or projected changes in atmospheric  $\text{CO}_2$  could be used as a selection factor for Arabica coffee adaptation. However, there appear to be clear interspecific differences between Arabica and robusta in relation to both growth and caffeine concentration at  $[\text{CO}_2]$  above a 300 ppm baseline. Such differences suggest potential for differential selection in fitness as  $\text{CO}_2$  continues to increase.

There are some obvious challenges in analyzing these data in the larger context of whether recent or projected  $[\text{CO}_2]$  can be used to select for more  $\text{CO}_2$  responsive coffee cultivars or coffee species. For example, vegetative development is known to be the most sensitive stage of growth in relation to rising  $[\text{CO}_2]$ <sup>23</sup> and has been suggested as a means to select for cultivar responsiveness in annual crops<sup>9,39</sup>. However, for tree crops, with slower relative growth, first year assessments may be useful in assessing initial response, but insufficient to discern longer-term differential effects on seed production (i.e., crop yield and quality). There are additional interspecific and intraspecific issues related to environmental shifts likely to change in parallel to rising  $[\text{CO}_2]$  such as precipitation and/or temperature that, in turn, will also influence selection and adaptation of coffee to climate change. Yet, as indicated by these initial data, it seems unlikely that Arabica and robusta will respond similarly to increasing  $[\text{CO}_2]$  and such potential differences may have long-term qualitative and quantitative consequences for Arabica and robusta production globally. In addition, it will be of interest to compare interspecific differences between Arabica with *C. eugenioides*, the other parent of Arabica coffee in a future study to determine if a similar response pattern is observed for *C. eugenioides*.

The basis for differential responses to rising  $[\text{CO}_2]$  between Arabica and robusta is uncertain. They may be related to: (1) interspecific variation, due to physical (morphological) or physiological differences, or a combination of both; (2) the effect of polyploidy, which amongst other features, influences cell size, genomic stability, gene expression and evolution rates<sup>40</sup>. All species of coffee are diploid ( $2n = 2x = 22$ ), except Arabica coffee, which is an allotetraploid ( $2n = 4x = 44$ )<sup>41,42</sup>. One of the recorded features of polyploidy in coffee is that higher ploidy results in fewer but larger stomata<sup>43,44</sup>, and this may be linked to the different  $\text{CO}_2$  effects we record in coffee. Another



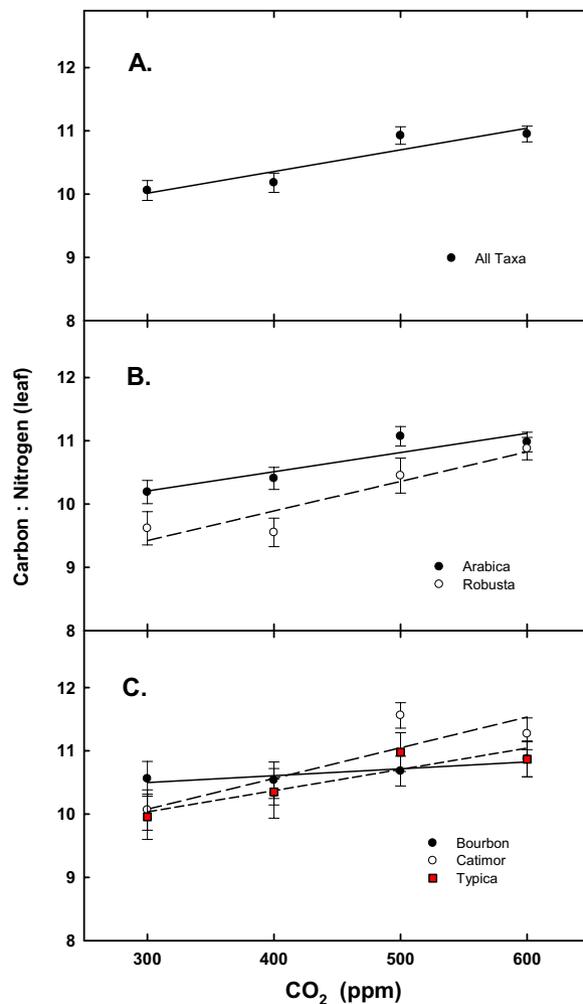
**Figure 4.** Differential changes between Arabica and robusta coffee (average  $\pm$  SE) for above ground dry weight, number of branches, and leaf weight in response to [CO<sub>2</sub>] at 357–368 DAS.

well-known consequence of polyploidy and specifically allopolyploids, is self-compatibility (self-fertilization)<sup>45–47</sup>. There are numerous evolutionary consequences for self-compatibility, including the reduction in genetic diversity<sup>48</sup>; for cultivated (farmed) Arabica coffee, this would be compounded by the severe genetic bottleneck created through the domestication process<sup>49</sup>. This may explain the lack of difference in CO<sub>2</sub> response in the three Arabica cultivars we have examined, although our sample size is not large enough to make any meaningful assessment.

There would appear to be potential for [CO<sub>2</sub>] to be used as a selective factor in adaptation and yield response for tree and perennial crops. Such efforts, however, are still in their infancy. The current study, the first to examine Arabica and robusta responses to recent and projected levels of CO<sub>2</sub>, from germination through the first year of growth, is suggestive of either interspecific differences or polyploidy level, but additional, long-term information will be needed to adequately determine how, and to what extent, recent and ongoing increases in [CO<sub>2</sub>] and/or climate change may act as a selection factor among Arabica cultivars. Moreover, it will be necessary to consider drought stress (reduced water availability), which so far has received scant attention in CO<sub>2</sub> enrichment influences for coffee with regard to climate change<sup>50</sup>. It has been argued that the influence of climate change on coffee production has been overestimated, although work so far has focused on elevated air temperatures<sup>22</sup>. Indeed, mitigation of elevated temperatures due to elevated CO<sub>2</sub> does seem to offer potential where there is adequate soil water availability (e.g., at field capacity)<sup>19,20</sup> but in many circumstances it is soil water availability (including temporal availability), and its relationship with other climatic variables (including temperature), that is the main limiting factor when considering climate change induced morbidity and mortality<sup>50</sup>. The interaction between elevated CO<sub>2</sub> and abscisic acid signaling, stomatal closure and CO<sub>2</sub> influx, as well as other physiological and chemical processes involved with drought<sup>51</sup>, require careful investigation.

## Methods

**Seeds.** Three Arabica cultivars widely grown throughout Latin America were tested: cv. ‘Bourbon’, cv. ‘Catimor’, and cv. ‘Typica’<sup>52,53</sup>. Typica and Bourbon are the progenitors of most Arabica coffee cultivars grown worldwide and are believed to have originated from coffee grown in Yemen of Ethiopian origin<sup>54,55</sup>. Arabica coffee grown in Indonesia originated from Yemen, and seeds taken from Java (Indonesia) to Amsterdam and then to



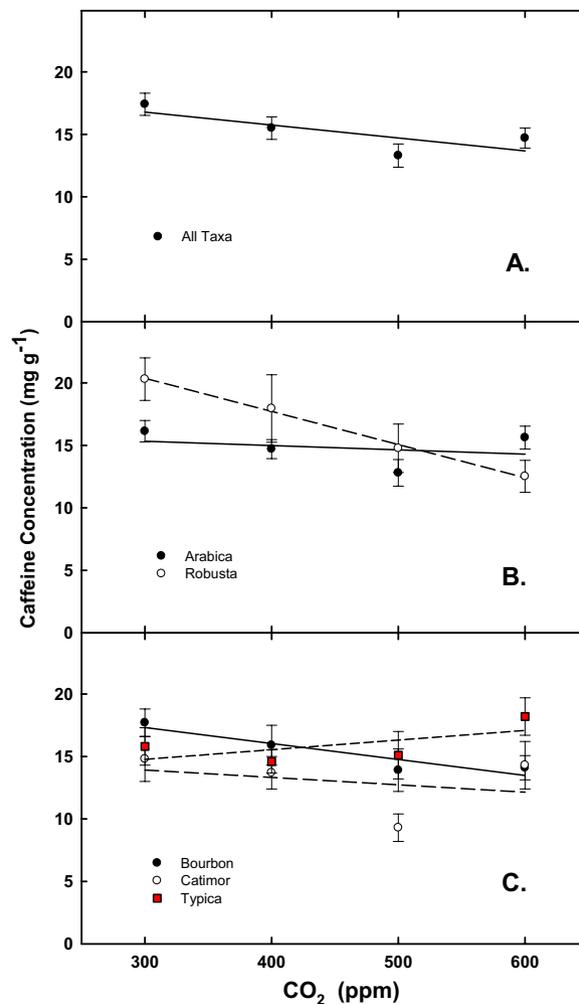
**Figure 5.** Response of carbon to nitrogen ratio ( $\pm$ SE) for: (A) all taxa; (B) Arabica and robusta coffee; and (C) all Arabica cultivars in response to  $[\text{CO}_2]$ .

the American continent led to the denomination Typica<sup>53</sup>. Seeds taken from Yemen and grown in Île de Bourbon (Bourbon Island; present day La Réunion) led to the denomination Bourbon<sup>18</sup>. Catimor is the result of crossing of two coffee cultivars: cv. ‘Caturra’ and cv. ‘Híbrido de Timor’ or ‘Timor Hybrid’ (a natural polyploid hybrid originating in Timor, an island in the Malay Archipelago, and resulting from a crossing between Arabica and robusta). Híbrido de Timor and the derived Catimor are resistant to coffee leaf rust (*Hemileia vastatrix*) and gained their resistance genes from robusta coffee<sup>52,53</sup>.

Mature coffee fruits for the Arabica cultivars were collected in August 2016, and again in September 2017 from plants at Rancho El Porvenir (869 masl; N 15.13229, W 92.20151) in Chiapas, Mexico. Robusta has higher levels of caffeine compared to Arabica (ca. 1.7% vs. 1%, respectively)<sup>56</sup> and is adapted to growth at lower elevations in Guineo-Congolian forests<sup>57</sup> and thus warmer and mostly wetter conditions relative to Arabica, which originates from high altitudes forest in Ethiopia and South Sudan and is adapted to a cooler, more seasonal environment<sup>58</sup>. Robusta fruits were collected in 2016 and again in 2017 from plants at Ejido Salvador Urbina (693 masl; N 15.04415 W 92.18578) in Chiapas, Mexico. Fruits were depulped, fermented, washed, and dried (ca. 12% moisture) and sent to the USDA-ARS Beltsville laboratory for germination.

**Planting.** Twelve plastic bins measuring ca. 60 cm  $\times$  50 cm  $\times$  33 cm deep (ca. 99 L by volume) were used to provide three monocultures of the four (three Arabica and one robusta) taxa for each  $[\text{CO}_2]$  treatment (four chambers). Each bin was perforated with 12 holes (1 cm diam.) to allow for water drainage. A screen mesh was placed at the bottom of each bin prior to adding the growing medium (Pro-Mix BX; Premier Horticulture Inc., Quakertown, CA, USA) to minimize growing medium loss after watering.

Seeds were soaked in water 24 h prior to planting, to promote germination. Each bin was moistened before planting 72 seeds per tub, ca. 2.5 cm deep, and ca. 5 cm apart. For the first run, seeds were planted on August 10, 2016 and the first germination occurred on September 5, 2016. For the second run, seeds were planted on September 12, 2017 and the first germination occurred on October 11, 2017. Rates of germination did not vary as a function of  $[\text{CO}_2]$ .



**Figure 6.** Caffeine concentration as a function of [CO<sub>2</sub>] (average +SE) for: (A) all taxa; (B) Arabica and robusta coffee; and, (C) all Arabica cultivars in response to [CO<sub>2</sub>].

For both trials, nutrients were initially provided at sowing and again at two months post-planting using a complete nutrient solution<sup>59</sup>. MiracleGro 24-8-16 (Marysville, OH) was provided at ca. 3 months following planting and given at 2–3 weeks' intervals until final harvest. An iron chelate micronutrient (Sprint 330, Becker Underwood, Ames, IA, USA) was sprayed as needed. The growth medium/soil was maintained at, or close to, field capacity.

**Environmental chambers.** Providing pre-ambient [CO<sub>2</sub>] concentrations is not possible *in situ*; therefore, controlled environment chambers (Bio-Chambers, Incorporated, Winnipeg, Canada) were used. The temperature for each chamber was kept constant at 25 °C, day/night. Light, quantified as photosynthetically active radiation (PAR), was maintained at 400 μmol mol<sup>-1</sup>. The daily light period was 12 h light was supplied by height-adjustable, dimmable banks of metal halide and high-pressure sodium bulbs (400 μmol m<sup>-2</sup> s<sup>-1</sup>).

CO<sub>2</sub> concentrations were maintained by injection of either CO<sub>2</sub> or CO<sub>2</sub>-free air using a TC-2 controller that monitors [CO<sub>2</sub>] in real time as measured by an infrared gas maintained in absolute mode. To maintain a range of recent and projected atmospheric CO<sub>2</sub>, concentrations were set at 300, 400, 500 and 600 ppm, 24 h day<sup>-1</sup>. These [CO<sub>2</sub>] values represent the measured Mauna Loa values from 1915 to 2015, and those projected by the end of the current century<sup>60</sup>. Actual mean [CO<sub>2</sub>] values (±SD, in [ppm]), from measurements recorded every three minutes throughout the experiments in each of the chambers, were 326 ± 38.6, 430 ± 42.7, 511 ± 26.2, and 607 ± 27.9 in the first run, and 303 ± 23.2, 409 ± 29.6, 499 ± 20.4, and 596 ± 23.0 in the second run.

**Harvests.** Destructive harvests were performed at three different times, ca. 4, 7, and 12 months post-planting. At each harvest, 3–5 plants within a bin (for all taxa and [CO<sub>2</sub>] treatments) were removed from the tubs, height determined (cm), then separated into leaf laminae, branches, stems, and roots. Leaf (cm<sup>2</sup>) area was determined photometrically using a leaf area meter (Li-Cor 3100, Lincoln, NE, USA). All plant material was weighed (g) after drying at 65 °C until dry weight was constant. Root binding did not occur as indicated by visual examination at the conclusion of the experiment when plants were removed from tubs.

**C:N ratios and caffeine analysis.** For each sample, all leaves, per plant were pooled and oven-dried (65 °C) until the sample was completely dry. Each dried sample was ground using a Wiley Mill with a mesh size #20. Total C and N contents were determined using a Vario Max CN (Elementary Americas, Inc., Ronkonkoma, NY, USA). Nitrogen and carbon content were determined as a percentage of the dry weight of the sample.

For extraction and determination of caffeine, leaves within a replicate were flash frozen in liquid N and stored at −80 °C until lyophilized. Leaves were then pulverized using an A11 Basic Analytical Mill (IKA Works Inc., Wilmington, NC, USA). A total of 100 mg of pulverized leaf material was added into 15 ml centrifuge tubes with 5.0 mL of a 70% methanol/water mixture. Tubes were then vortexed for 30 s and sonicated for 60 min. The slurry was centrifuged at 5,000 rpm for 10 min before being diluted (1:20), filtered, and ultimately stored in 1.5 mL HPLC vials. All reagents used for the analysis were of HPLC grade purity and prepared fresh on each day of the analysis. Instrumental analysis was performed using a Shimadzu Prominence High Performance Liquid Chromatograph (Shimadzu Scientific Instruments, Columbia, MD, USA) using a mobile phase of 80% methanol/water and 15 mM phosphate buffer at pH 6.2. Separation was conducted using a Thermo Scientific Aquasil reverse phase C18 column (4.6 × 250 mm, 5 μm particle size; Thermo Fisher Scientific, Waltham, MA, USA) at a flow rate of 0.550 ml/min. Detection and quantification was done using a UV detector at 275 nm and determined using a calibration curve. The caffeine calibration curve was created using an HPLC grade caffeine standard (99.7% purity; ACROS Organics #10816-5000; Thermo Fisher Scientific, Waltham, MA, USA) across five concentrations 2.5, 5, 10, 20, and 25 ppm. The fitted curve showed excellent linear responsivity as demonstrated by an  $r^2$  of 0.998. In addition, there was negligible variation between replicate injections at 10 ppm using the same standard as measured by its percent relative standard deviation of 0.385%.

The caffeine concentration in leaves can also be used as a proxy for concentrations in coffee beans, based on a correlation between caffeine concentration in seedling leaves and seeds<sup>61,62</sup>. Dias Chaves *et al.*<sup>61</sup> focused on the 1<sup>st</sup> and 3<sup>rd</sup> pair of leaves in the seedlings, while de Moraes *et al.*<sup>62</sup> used the 3<sup>rd</sup> and 4<sup>th</sup> pair. We found no significant differences in caffeine content between the last pair of fully expanded leaves and all remaining leaves combined (cotyledons excluded; using March 2017 samples, i.e., first year, second sampling; 7 months and 18 days post-planting). Based on these results, we pooled all leaves at each sampling date for caffeine analysis. Mazzafera and Magalhães<sup>63</sup> found no correlation between leaves and seeds, but these were collected from mature plants, not seedlings.

**Statistical analysis.** Three replicate bins for each Arabica cultivar and for robusta coffee (i.e., 12 bins per chamber) were present for each of four [CO<sub>2</sub>] treatments. Within each chamber [CO<sub>2</sub>], bins were randomized; and randomized again after the first two harvests at 4 and 7 months to avoid edge effects. After the first run of the experiment (i.e., one year), the chambers were randomly reassigned [CO<sub>2</sub>] treatments and the experiment repeated. Humidity, PAR, and temperature were quantified before and at the end of each harvest to determine within chamber and among chamber variability. Values for each parameter were consistent between experimental runs. All measured parameters were based on tub averages (3–4 plants per tub) for both runs. All measured and calculated parameters were analyzed using analysis of variance including [CO<sub>2</sub>], Arabica cultivars, Arabica vs. robusta, and harvest time (Statview Software, Cary, NC, USA).

Received: 6 September 2019; Accepted: 16 March 2020;

Published online: 03 April 2020

## References

- Burton, D. A. 2020. Sea-Level information. [https://www.sealevel.info/co2\\_and\\_ch4.html](https://www.sealevel.info/co2_and_ch4.html).
- Cure, J. D. & Acock, B. Crop responses to carbon dioxide doubling: a literature survey. *Agric. For. Meteorol.* **38**, 127–145 (1986).
- Kimball, B. A., Kobayashi, K. & Bindi, M. Responses of agricultural crops to free-air CO<sub>2</sub> enrichment. *Adv. Agron.* **77**, 293–368 (2002).
- Loladze, I. Hidden shift of the ionome of plants exposed to elevated CO<sub>2</sub> depletes minerals at the base of human nutrition. *elife* **3**, e02245 (2014).
- Gamage, D. *et al.* New insights into the cellular mechanisms of plant growth at elevated atmospheric carbon dioxide concentrations. *Plant, Cell Environ.* **41**, 1233–1246 (2018).
- Bertolino, L. T., Caine, R. S. & Gray, J. E. Impact of stomatal density and morphology on water-use efficiency in a changing world. *Front. Plant Sci.* **10**, 225 (2019).
- Ahmed, F. E., Hall, A. E. & Madore, M. A. Interactive effects of high temperature and elevated carbon dioxide concentration on cowpea (*Vigna unguiculata* (L.) Walp.). *Plant, Cell Environ.* **16**, 835–842 (1993).
- Bunce, J. A. Contrasting responses of seed yield to elevated carbon dioxide under field conditions within *Phaseolus vulgaris*. *Agric., Ecosyst. Environ.* **128**, 219–224 (2008).
- Shimono, H. *et al.* Genotypic variation in rice yield enhancement by elevated CO<sub>2</sub> relates to growth before heading, and not to maturity group. *J. Exp. Bot.* **60**, 523–532 (2008).
- Hasegawa, T. *et al.* Rice cultivar responses to elevated CO<sub>2</sub> at two free-air CO<sub>2</sub> enrichment (FACE) sites in Japan. *Funct. Plant Biol.* **40**, 148–59 (2013).
- Wang, D. R. *et al.* Evidence for divergence of response in *Indica*, *Japonica*, and wild rice to high CO<sub>2</sub> × temperature interaction. *Global Change Biol.* **22**, 2620–2632 (2016).
- Ziska, L. H., Morris, C. F. & Goins, E. W. Quantitative and qualitative evaluation of selected wheat varieties released since 1903 to increasing atmospheric carbon dioxide: can yield sensitivity to carbon dioxide be a factor in wheat performance? *Global Change Biol.* **10**, 1810–1819 (2004).
- Bunce, J. Using FACE systems to screen wheat cultivars for yield increases at elevated CO<sub>2</sub>. *Agronomy* **7**, 20 (2017).
- Li, Y. *et al.* Soybean intraspecific genetic variation in response to elevated CO<sub>2</sub>. *Arch. Agron. Soil Sci.* **65**, 1733–1744 (2019).
- Osorio, N. The global coffee crisis: a threat to sustainable development. *International Coffee Organization, London*, <http://dev.ico.org/documents/globalcrisis.pdf> (2002).
- Davis, A. P., Chadburn, H., Moat, J., O'Sullivan, R. & Hargreaves, S. E. Nic Lughadha, High extinction risk for wild coffee species and implications for coffee sector sustainability. *Sci. Adv.* **5**, eaav3473 (2019).
- USDA-FAS, Coffee: world markets and trade. United States Department of Agriculture, Foreign Agricultural Service <https://downloads.usda.library.cornell.edu/usda-esmis/files/m900nt40f/xk81jw68v/kp78gs60d/coffee.pdf> (June 2019).
- DaMatta, F. M. *et al.* Sustained enhancement of photosynthesis in coffee trees grown under free-air CO<sub>2</sub> enrichment conditions: disentangling the contributions of stomatal, mesophyll, and biochemical limitations. *J. Exp. Bot.* **67**, 341–352 (2016).

19. Ramalho, J. C. *et al.* Sustained photosynthetic performance of *Coffea* spp. under long-term enhanced [CO<sub>2</sub>]. *PLoS ONE* **8**(12), e82712 (2013).
20. Ramalho, J. C. *et al.* Can elevated air [CO<sub>2</sub>] conditions mitigate the predicted warming impact on the quality of coffee bean? *Front. Plant Sci.* **9**, 287 (2018).
21. Martins, L. D., Tomaz, M. A., Lidon, F. C., DaMatta, F. M. & Ramalho, J. C. Combined effects of elevated [CO<sub>2</sub>] and high temperature on leaf mineral balance in *Coffea* spp. plants. *Clim. Change* **126**, 365–379 (2014).
22. Rodrigues, W. P. *et al.* Long-term elevated air [CO<sub>2</sub>] strengthens photosynthetic functioning and mitigates the impact of supra-optimal temperatures in tropical *Coffea arabica* and *C. canephora* species. *Global Change Biol.* **22**, 415–431 (2016).
23. Sakai, H., Hasegawa, T. & Kobayashi, K. Enhancement of rice canopy carbon gain by elevated CO<sub>2</sub> is sensitive to growth stage and leaf nitrogen concentration. *New Phytol.* **170**, 321–332 (2006).
24. Cotrufo, M. F., Ineson, P. & Scott, A. Elevated CO<sub>2</sub> reduces the nitrogen concentration of plant tissues. *Global Change Biol.* **4**, 43–54 (1998).
25. Pleijel, H., Broberg, M. C., Högy, P. & J. Uddling, P. Nitrogen application is required to realize wheat yield stimulation by elevated CO<sub>2</sub> but will not remove the CO<sub>2</sub>-induced reduction in grain protein concentration. *Global Change Biol.* **25**, 1868–1876 (2019).
26. Zhu, C. *et al.* Carbon dioxide (CO<sub>2</sub>) levels this century will alter the protein, micronutrients, and vitamin content of rice grains with potential health consequences for the poorest rice-dependent countries. *Sci. Adv.* **4**(5), eaq1012 (2018).
27. Levinson, H. Z. The defensive role of alkaloids in insects and plants. *Experientia* **32**, 408–411 (1976).
28. Nathanson, J. A. Caffeine and related methylxanthines: possible naturally occurring pesticides. *Science* **226**, 184–187 (1984).
29. Vega, F. E., Blackburn, M. B., Kurtzman, C. P. & Dowd, P. F. Identification of a coffee berry borer-associated yeast: does it break down caffeine? *Entomol. Exp. Appl.* **107**, 19–24 (2003).
30. Araque, P., Casanova, H., Ortiz, C., Henao, B. & Peláez, C. Insecticidal activity of caffeine aqueous solutions and caffeine oleate emulsions against *Drosophila melanogaster* and *Hypothenemus hampei*. *J. Agric. Food Chem.* **55**, 6918–6922 (2007).
31. Davis, A. P., Gole, T. W., Baena, S. & Moat, J. The impact of climate change on natural populations of Arabica coffee: predicting future trends and identifying priorities. *PLoS ONE* **7**, e47981 (2012).
32. Curtis, P. S., Snow, A. A. & Miller, A. S. Genotype-specific effects of elevated CO<sub>2</sub> on fecundity in wild radish (*Raphanus raphanistrum*). *Oecologia* **97**, 100–105 (1994).
33. Ziska, L. H. *et al.* Food security and climate change: on the potential to adapt global crop production by active selection to rising atmospheric carbon dioxide. *Proc. Roy. Soc. London, Ser. B* **279**, 4097–4105 (2012).
34. Ainsworth, E. A. & Long, S. P. What have we learned from 15 years of free-air CO<sub>2</sub> enrichment (FACE)? A meta-analytic review of the responses of photosynthesis, canopy properties and plant production to rising CO<sub>2</sub>. *New Phytol.* **165**, 351–371 (2005).
35. Resco de Dios, V., Mereed, T. E., Ferrio, J. P., Tissue, D. T. & Voltas, J. Intraspecific variation in juvenile tree growth under elevated CO<sub>2</sub> alone and with O<sub>3</sub>: a meta-analysis. *Tree Physiol.* **36**, 682–693 (2016).
36. Ainsworth, E. A. The importance of intraspecific variation in tree responses to elevated [CO<sub>2</sub>]: breeding and management of future forests. *Tree Physiol.* **36**, 679–681 (2016).
37. Aspinwall, M. J. *et al.* Tissue, photosynthesis and carbon allocation are both important predictors of genotype productivity responses to elevated CO<sub>2</sub> in *Eucalyptus camaldulensis*. *Tree Physiol.* **38**, 1286–1301 (2018).
38. Ziska, L. H. *et al.* Climate change, carbon dioxide, and pest biology, managing the future: coffee as a case study. *Agronomy* **8**, 152 (2018).
39. Shimono, H. *et al.* Prescreening in large populations as a tool for identifying elevated CO<sub>2</sub>-responsive genotypes in plants. *Funct. Plant Biol.* **46**, 1–14 (2019).
40. Otto, S. P. The evolutionary consequences of polyploidy. *Cell* **131**, 452–462 (2007).
41. Krug, C. A. & Mendes, A. J. T. Cytological observations in *Coffea* IV. *J. Genet.* **39**, 189–203 (1940).
42. Krug, C. A., Carvalho, A. The genetics of *Coffea*. *Adv. Genet.* **4**, 127–158 (1951).
43. Franco, C. M. Relation between chromosome number and stomata in *Coffea*. *Bot. Gaz.* **100**, 817–827 (1939).
44. Mishra, M. K. Stomatal characteristics at different ploidy levels in *Coffea* L. *Ann. Bot.* **80**, 689–692 (1997).
45. Osabe, K. *et al.* Multiple mechanisms and challenges for the application of allopolyploidy in plants. *Int. J. Mol. Sci.* **13**, 8696–8721 (2012).
46. Comai, L. The advantages and disadvantages of being polyploid. *Nat. Rev. Genet.* **6**, 836–846 (2005).
47. Okamoto, S. *et al.* Self-compatibility in *Brassica napus* is caused by independent mutations in S-locus genes. *Plant J.* **50**, 391–400 (2007).
48. Wright, S. I., Kalisz, S. & Slotte, T. Evolutionary consequences of self-fertilization in plants. *Proc. R. Soc. B* **280**, 20130133 (2013).
49. Anthony, F. *et al.* The origin of cultivated *Coffea arabica* L. varieties revealed by AFLP and SSR markers. *Theor. Appl. Genet.* **104**, 894–900 (2002).
50. Moat, J. *et al.* Resilience potential of the Ethiopian coffee sector under climate change. *Nat. Plants* **3**, 17081 (2017).
51. Farooq, M., Wahid, A., Kobayashi, N., Fujita, D. & Basra, S. M. A. Plant drought stress: effects, mechanisms and management. *Agron. Sustain. Develop.* **29**, 185–212 (2009).
52. van der Vossen, H. A. M. Coffee selection and breeding, in *Coffee. Botany, Biochemistry and Production of Beans and Beverage*, Clifford, M. N., Willson, K. C., Eds., pp. 48–96 (Croom Helm, 1985).
53. Wrigley, G. *Coffee* (Longman Scientific & Technical, 1988).
54. Wellman, F. L. *Coffee: Botany, Cultivation, and Utilization* (Leonard Hill [Books] Ltd., 1961).
55. Vega, F. E. The rise of coffee. *Am. Sci.* **96**, 138–145 (2008).
56. Ashihara, H. & Suzuki, T. Distribution and biosynthesis of caffeine in plants. *Front. Biosci.* **9**, 1864–1876 (2004).
57. Davis, A. P., Govaerts, R., Bridson, D. M. & Stoffelen, P. An annotated taxonomic conspectus of the genus *Coffea* (Rubiaceae). *Bot. J. Linn. Soc.* **152**, 465–512 (2006).
58. Davis, A. P. *et al.* *Coffee Atlas of Ethiopia*. Royal Botanic Gardens, Kew (2018).
59. Robinson, J. M. Photosynthetic carbon metabolism in leaves and isolated chloroplasts from spinach plants grown under short and intermediate photosynthetic periods. *Plant Physiol.* **75**, 397–409 (1984).
60. National Oceanic and Atmospheric Administration (NOAA). Earth System Research Laboratory, Global Monitoring Division. <http://www.esrl.noaa.gov/gmd/ccgg/trends/>.
61. Dias Chaves, J. C., Miyazawa, M., de, M., Mesarina Bloch, F. & Yamakami, J. K. Estimativa do teor de cafeína nas sementes de café baseada na sua concentração nas folhas de mudas e de plantas adultas. *Acta Scientiarum. Agronomy* **26**, 287–292 (2004).
62. De Moraes, B. F. X. *et al.* Correlação entre teor de cafeína em folhas e grãos de café. XIX Congresso de Pós-Graduação da UFPA, 27 de setembro a 01 de outubro de 2010, 5 pp. (2010).
63. Mazzafera, P. & Magalhães, A. C. Cafeína em folhas e sementes de *Coffea* e *Paracoffea*. *Rev. Bras. Bot.* **14**, 157–160 (1991).

## Acknowledgements

We thank H. Vázquez Álvarez and L. Vázquez López in Chiapas, Mexico, for giving us permission to collect coffee fruits from their plantations; two reviewers for comments on a previous version of this paper; D. Baxam (USDA-ARS) for assistance in running the growth chambers; and R. Erdman (USDA-ARS) for help in sampling coffee plants.

### Author contributions

F.E.V., L.H.Z. and A.S. conceived the project and designed the study; F.E.V., A.S. and J.W. ran the experiments; F.E.V., L.H.Z. and A.P.D. did the literature review; L.H.Z. did the statistical analysis; L.H.Z., F.E.V. and A.P.D. wrote the report. F.E.V., L.H.Z., A.S., F.I., A.P.D., J.A.R., J.Y.B. and J.W. interpreted the results, commented on the draft version of the report, and approved the submission draft.

### Competing interests

The authors declare no competing interests.

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