

## Synergistic ammonia losses from animal wastewater



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### HIGHLIGHTS

- ▶ Measured ammonia emissions from pure (NH<sub>4</sub>)<sub>2</sub>SO<sub>4</sub> solution and animal wastewater.
- ▶ Observed enhanced ammonia emissions from both swine and dairy wastewater.
- ▶ Multiple reactions in animal wastewater control NH<sub>3</sub> emissions across thin layers.
- ▶ Modeling application needs to go with better understanding of ammonia emissions.

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### ABSTRACT

Thin-layer models are commonly used to estimate NH<sub>3</sub> emissions from liquid waste. However, such models differ in their ability to accurately reproduce observed emissions, which may be partly due to an incomplete mechanistic understanding of NH<sub>3</sub> volatilization. In this study, NH<sub>3</sub> release from pure solutions of (NH<sub>4</sub>)<sub>2</sub>SO<sub>4</sub> (AS), swine pit liquid (PL), swine lagoon liquid (LL), dairy lagoon liquid (DLL), and dairy manure liquid (DML) were evaluated under controlled conditions (pH 7.6 and temperature 20 °C). Relationships between the NH<sub>3</sub> release and pH, temperature, and total ammoniacal nitrogen (TAN) were established. Under similar TAN conditions, the NH<sub>3</sub> release was higher for PL, LL, DLL, and DML than for AS. Pure solutions of AS that were spiked with NaHCO<sub>3</sub> showed NH<sub>3</sub> emissions rates that were comparable to DML, DLL, LL and PL. The enhanced NH<sub>3</sub> emissions of PL, LL, DLL and DML were therefore explained by linkages between TAN and HCO<sub>3</sub><sup>-</sup>.

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### 1. Introduction

Animal feed operations (AFOs) are estimated to account for 71% of the total human-induced NH<sub>3</sub> emissions in the United States (USEPA, 2004). Atmospheric deposition of NH<sub>3</sub> can contribute to eutrophication of surface waters (Paerl, 1995), over-fertilization of crops (Galloway and Dentener, 2004), and soil acidification (Van Breemen et al., 1982). Furthermore, NH<sub>3</sub> emission from AFOs is regarded as a significant source for atmospheric particulate matter <2.5 μm (Qi and Kleeman, 2006), PM<sub>2.5</sub>. Health studies indicate that exposure of individuals to PM<sub>2.5</sub> is associated with increased risks of respiratory and cardiovascular problems (Franklin and Zeka, 2006). In addition, PM<sub>2.5</sub> levels influence haze formation (Trijonis et al., 1991). An understanding of factors that influence

NH<sub>3</sub> emission from AFOs is therefore essential for ecosystem and human health.

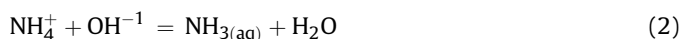
In dilute solutions, the equilibrium partitioning of NH<sub>3</sub> between the liquid and gas phases is described using Henry's Law (Sawyer and McCarty, 1978; Buonicore and Davis, 1992; Stumm and Morgan, 1995):

$$v_p = k_H C \quad (1)$$

where  $v_p$  is the vapor pressure of the gaseous species,  $C$  is the concentration of the dissolved gaseous species in the liquid phase, and  $k_H$  is the Henry's Law constant. Eq. (1) implies that the emission of NH<sub>3</sub> from a liquid phase would be controlled by the concentration of dissolved NH<sub>3</sub> within the liquid phase (Sawyer and McCarty, 1978; Buonicore and Davis, 1992; Stumm and Morgan, 1995), which in turn depends on the concentration of ammonium (NH<sub>4</sub><sup>+</sup>) in solution as demonstrated in the following mass law expression:

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where  $(\text{aq})$  means as a dissolved gas in solution. Eq. (2) illustrates that the concentration of dissolved  $\text{NH}_3$  is a direct function of the concentration of  $\text{NH}_4^+$  in solution and pH. Under conditions of constant pH, increasing the concentration of  $\text{NH}_4^+$  in solution should increase  $\text{NH}_3$  emissions due to the corresponding increase in dissolved  $\text{NH}_3$  concentrations in the liquid phase. In theory, there is no upper limit on the possible  $\text{NH}_3$  emissions with increasing  $\text{NH}_4^+$  concentrations in solution.

Wastewaters from AFOs are complex multicomponent chemical mixtures that contain bacteria. Some researchers have found that  $\text{NH}_3$  emission is dependent on the solution chemistry condition (Berger and Libby, 1969; Ni et al., 2009). Furthermore, microbial activity in the wastewater is expected to influence many of these reactions.  $\text{NH}_3$  emissions from wastewaters are therefore likely to be much more complicated than Eq. (2) suggests. For example, microbially mediated production of  $\text{CO}_2$  and/or production and consumption of other gases (e.g., volatile organic carbons) may influence the emission of  $\text{NH}_3$  (Chai et al., 2005). Ni et al. (2009) developed a model that includes a dependency of  $\text{NH}_3$  emission on  $\text{CO}_2$ . The emission of  $\text{NH}_3$  is also well-known to be dependent on temperature (Loeher, 1984; Jayaweera and Mikkelsen, 1990) which influences the rates of chemical reactions and microbial activity, as well as diffusion coefficients. For example, the value of  $k_H$  is a function of temperature that can be described using the Arrhenius equation (Stumm and Morgan, 1995).

Thin-film mass transfer models are commonly used to estimate  $\text{NH}_3$  emissions from liquid animal wastes (Ni et al., 2000). The underlying assumption of this approach is that the mass transfer rate of a gas compound from a liquid to the overlying air is controlled by the rates of diffusion through the boundary layers on each side of the interface. When the liquid phase is well mixed, the diffusive  $\text{NH}_3$  flux ( $J_{gw}$ ) is commonly described using a quasi-steady approximation of Fick's first law for diffusion in the gas phase as:

$$J_{gw} = K_{gw}(k_H C - C_g) \quad (3)$$

where  $K_{gw}$  is the mass transfer coefficient and  $C_g$  and  $C$ , respectively, are the bulk gas phase and liquid phase concentrations of  $\text{NH}_3$ . The mass transfer coefficient is a function of the diffusion coefficient and the boundary layer thickness. Another central parameter in this modeling framework is the concentration gradient across the boundary layer that depends on the equilibrium concentration ( $k_H C$ ) at the air–water interface (Jury et al., 1991).

Our knowledge about factors that influence  $\text{NH}_3$  emissions from wastewaters is currently limited. Empirical and mechanistic models of  $\text{NH}_3$  emissions often yield different results, underscoring the need to examine the processes driving emissions (Harper and Sharpe, 1998; Aneja et al., 1997; 1996a, 1996b). An incomplete mechanistic understanding of  $\text{NH}_3$  production and emission from wastewater may partly be responsible for this disagreement. The uncertainties surrounding the form and assumptions underlying the thin-film modeling approach are the primary motivation for this study. In this study,  $\text{NH}_3$  emissions from pure solutions of  $(\text{NH}_4)_2\text{SO}_4$  (AS), swine and dairy wastewaters were evaluated under controlled laboratory conditions. Relationships between  $\text{NH}_3$  fluxes and total ammoniacal nitrogen ( $\text{TAN} = \text{NH}_3 + \text{NH}_4^+$ ), pH, and temperature were examined. Comparison of these relationships across treatments established an experimental framework by which the assumptions underlying the typical thin-film  $\text{NH}_3$  emission model were investigated.

## 2. Methods

### 2.1. Test solutions

Pure ammonium sulfate (AS) solutions at varying TAN concentrations were prepared using reagent grade AS. Samples of swine liquid waste, lagoon liquid (LL) and pit liquid (PL) were obtained from an anaerobic lagoon and shallow pits under the housing units from a swine finisher facility in Warsaw, NC. The samples of liquid waste from the shallow pits under the housing units at the finisher farm were composites and included liquid from several houses. Dairy wastewater samples were obtained from a trench containing dairy manure and liquid (DML) wastewater, and from a dairy anaerobic lagoon liquid (DLL) wastewater on a dairy farm in San Jacinto, CA. Wastewater samples were placed in 20 L carboys, transported back to the laboratory, and stored at 4 °C until use.

The pH buffer solutions were prepared using nearly equal volumes of 0.1 M  $\text{KH}_2\text{PO}_4$  solutions and 0.1 M NaOH solution to achieve the desired pH (pH 7.6). The TAN concentration and pH of the DML, DLL, LL and PL samples was controlled by diluting with deionized water, AS solutions, and pH buffer solutions. The final pH of the buffer solutions and the test solutions was determined before and after experiments using a pH meter and combination glass electrode calibrated with NIST-traceable pH standards (Denver UB-10 pH  $\text{mV}^{-1}$  meter equipped with a High performance pH ATC<sup>-1</sup> Glass-body Electrode).

The TAN concentration of the AS, PL and LL samples was determined using a steam distillation method (U.S. EPA Method 350.2) that is well documented in the literature (Bremner, 1965; Bundy and Bremner, 1973; Stevenson, 1982a, b). TAN concentration of the AS, DML and DLL samples was determined using an Alpkem/OI Analytical Flow Solution IV with autosampler (U.S. EPA Method 350.1). The Alpkem Flow Solution IV with autosampler was equipped with ammonia/TKN cartridges allowing for a variety of measurements of different forms of nitrogen in solution. Additional details about these analytical methods and quality assurance and control protocols are available in the Supporting information (si).

### 2.2. Experimental setups

Fig. 1 presents illustrations of two experimental setups that were used to contain test solutions and to measure the emission of  $\text{NH}_3$  in response to changes in solution composition, pH, and temperature. The first setup consisted of a 4 L pyrex reagent bottle chamber (RBC) (Fig. 1). The RBC has an internal volume of 4 L (depth 21 cm × diameter 16 cm) and a surface area ( $S_A$ ) of 201  $\text{cm}^2$  at the air–liquid interface. The inlet (0.8 cm internal diameter) and outlet (0.4 cm internal diameter) of the RBC consisted of valves placed in the interior of a #8 stopper. The RBC was filled with 2 L of a selected test solution and positioned into a water bath. The temperature of the water bath was controlled by an internal heating and refrigeration unit (Forma Scientific CH/P temperature control system). A pump (Cole Parmer Instruments Model Number 7553-00) was used to slowly circulate the test solution in the RBC. The pump rate was set to completely circulate all ~2 L of the test solution once every 20 min. The temperature of the test solution in the RBC was monitored manually at the center location using a digital thermometer with a tolerance of  $\pm 0.1$  °C (Fisher Scientific digi-thermo thermocouples with external stainless steel probes).

To test the validity of results from the RBC to larger scales a second experimental setup was employed. This setup (Fig. 1b) consisted of a simple flow-through teflon-lined chamber (SFTC). The SFTC has a volume of 9 L (length 30 cm × width 20 cm × height 15 cm) and a  $S_A$  of 600  $\text{cm}^2$  at the air–liquid interface. Two inlets (~1 cm internal diameter) were used to allow air flow into the

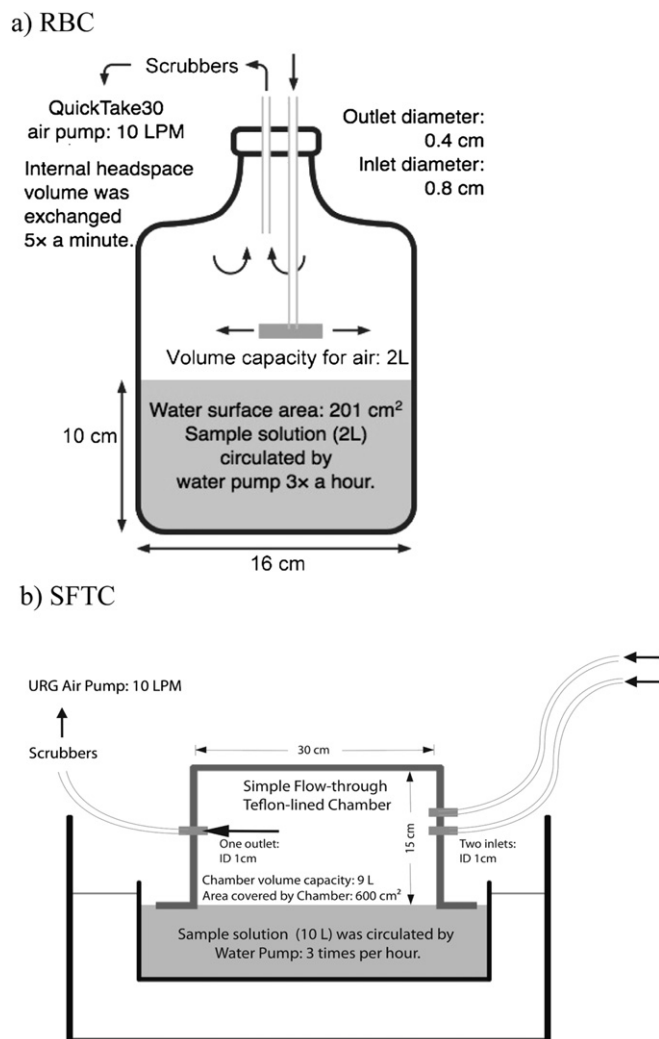


Fig. 1. Schematics of a) reagent bottle chamber (RBC) and b) simple flow-through teflon-lined chamber (SFTC).

chamber. A single outlet ( $\sim 1$  cm internal diameter) was located on the opposite wall of the SFTC. The inlets and outlets were bulkhead fittings which contained internal valves that were only open when the appropriate hose connector was inserted into the fitting. The SFTC was positioned over the test solutions contained in a plastic pan and lowered into the pan until the SFTC was floating on the test solution supported by the flange around the outside of the chamber. Direct contact of the chamber with the test solutions resulted in an air-tight seal of the chamber at the air–liquid interface.

Air flow ( $Q_A$ ) through the RBC and SFTC setups was held constant at 10 LPM using calibrated air pumps located downstream of the chamber outlet. This translates to effective wind speeds ( $Q_A/S_A$ ) of approximately  $0.008$  to  $0.003$  m s<sup>-1</sup> in the RBC and SFTC setups, respectively, and limited the impact of turbulence on emissions. NH<sub>3</sub> emitted from the test solutions was scrubbed from the air exiting the chambers using glass scrubbers containing H<sub>2</sub>SO<sub>4</sub> solution (0.05 or 0.025 N) and fritted glass on the inlets. Ammonia loss measurements from the test solutions were typically conducted over periods of 1 h after the solution achieved the desired temperature. Additional details about the quality assurance and control protocols are available in the [Supporting information](#).

The design and effective wind speeds of the RBC and SFTC setups were selected to maximize  $J_{gw}$ . Eq. (3) indicates that a

maximum value of  $J_{gw} = K_{gw}k_H C$  occurs when  $C_g \approx 0$ . Under these conditions the effects of solution chemistry on equilibrium partitioning ( $k_H C$ ) at the air–liquid interface can be readily examined. Low values of  $C_g$  were achieved in RBC and SFTC setups because of short air phase residence times of 0.2 and 0.9 min, respectively. The equilibrium value for  $C_g = k_H C / (RT_k)$  was calculated to be around  $1688$  mg NH<sub>3</sub>-N m<sup>-3</sup> when TAN =  $2500$  mg NH<sub>4</sub>-N m<sup>-3</sup>; where  $R$  is the gas constant and  $T_k$  is the temperature in degrees Kelvin. Experimental results presented below were several orders of magnitude lower than this equilibrium prediction. Most of the RBC and SFTC experiments were replicated and some were run in triplicate.

### 3. Results and discussion

#### 3.1. Performance of the experimental chambers

The performance of the RBC and SFTC setups were assessed by measuring the flux of NH<sub>3</sub> from pure AS solutions under differing TAN concentrations ( $100$ – $500$  mg NH<sub>4</sub>-N L<sup>-1</sup>). Period average measurements of ambient NH<sub>3</sub> concentrations (during  $\sim 1$  h duration) were obtained by dividing the total mass of NH<sub>3</sub> emitted from the chamber by the total air volume passed through the acid scrubbers. The final calculated ammonia concentration values were standardized to units of mg NH<sub>3</sub>-N m<sup>-3</sup>. Fig. 2 presents a plot of the period average NH<sub>3</sub> concentration as a function of the TAN concentration in pure AS solutions under constant temperature ( $20$  °C) and pH (pH = 7.6) conditions. As predicted by Eq. (2), the measured NH<sub>3</sub> concentration increases linearly with an increase in TAN concentration of the AS solutions. The RBC and SFTC setups gave nearly identical results, demonstrating that findings from the smaller RBC were comparable with the larger SFTC. The corresponding linear regression equations are presented in the figure. At concentrations of TAN >  $500$  mg NH<sub>4</sub>-N L<sup>-1</sup> there appears to be a definite linear relationship between the calculated NH<sub>3</sub> concentration and TAN. Failure of the regression equations to have a zero intercept is probably due to experimental error in the measurement of low NH<sub>3</sub> concentrations at low TAN. Error bars derived from replicate experiments were very small (Fig. 2). For simplicity, subsequent figures do not include error bars.

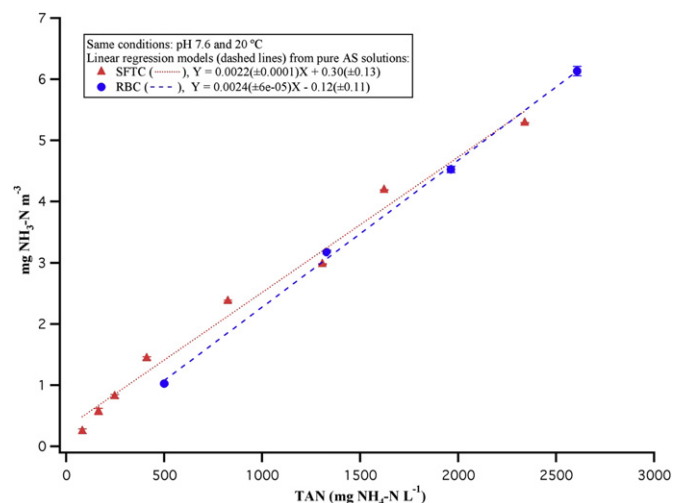


Fig. 2. Measurements of atmospheric NH<sub>3</sub> release from ammonium sulfate (AS) solutions as a function of TAN (pH 7.6,  $T = 20$  °C, SFTC and RBC). Dashed line represents linear regression model ( $X = \text{TAN}$ ,  $Y = \text{Air Concentration}$ ). Error bars on data points were derived from replicate experiments.

### 3.2. Emissions of $\text{NH}_3$ from AS solutions as a function of pH and temperature

Eq. (2) suggests a dependency of  $\text{NH}_3$  emission on pH and temperature (reaction rates and diffusion coefficients are dependent on temperature). Fig. 3 presents the period averaged  $\text{NH}_3$  concentration emitted from pure AS solutions in the RBC system at different pH (7.0–8.0) and temperature (10–25 °C) conditions. The TAN concentration of the AS solutions equaled 370 and 412  $\text{mg NH}_4\text{-N L}^{-1}$  for temperature and pH experiments, respectively. Manual measurements of temperature during these experiments showed temperature control of the test solutions within  $\pm 0.1$  °C. Measurements of pH and TAN concentration before and after the measurement period showed little to no change, demonstrating that the volume of the test solution used and the duration of the test period resulted in no chemical changes. The  $\text{NH}_3$  loss from the AS solution increases exponentially with water temperature and pH (Fig. 3). These observations are consistent with expectations based on Eq. (2) and the Arrhenius equation (Stumm and Morgan, 1995).

### 3.3. Emissions of $\text{NH}_3$ from animal wastewater

Fig. 4 presents plots of the period averaged  $\text{NH}_3$  concentrations from AS and AFO wastewaters (PL, LL, DML and DLL) at constant temperature (20 °C) and pH (pH = 7.6) as a function of TAN. Typically, the TAN of the DML and PL samples was higher than the DLL and LL samples. The extended TAN concentrations for the PL, LL, DML and DLL samples were obtained by spiking with solid reagent-grade AS. The response in the emitted  $\text{NH}_3$  concentration with changes in TAN of the wastewaters was linear over the range of tested TAN. This result indicates that spiking the wastewaters with AS was a valid approach. Regression equations for the emitted  $\text{NH}_3$  concentration and the TAN concentration are also presented in the figure. The calculated uncertainty associated with the intercept values was consistent with zero  $\text{NH}_3$  emissions as TAN approaches zero.

Although the response in the emitted  $\text{NH}_3$  concentration with TAN was linear for the PL, LL, and DML wastewater solutions, the absolute magnitude of the calculated  $\text{NH}_3$  concentration differed

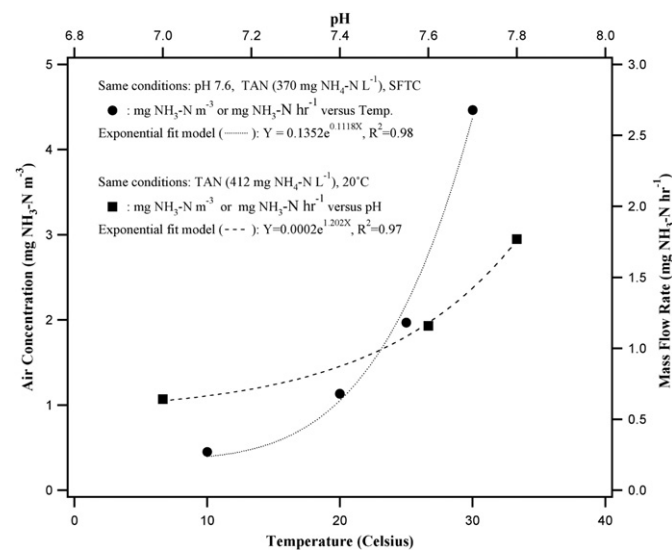


Fig. 3. Measurements of atmospheric  $\text{NH}_3$  release from pure ammonium sulfate (AS) solutions as a function of temperature (TAN 370  $\text{mg NH}_4\text{-N L}^{-1}$ , pH 7.6) and pH (TAN 412  $\text{mg NH}_4\text{-N L}^{-1}$ ,  $T = 20$  °C) using SFTC. Dashed lines represent linear regression models.

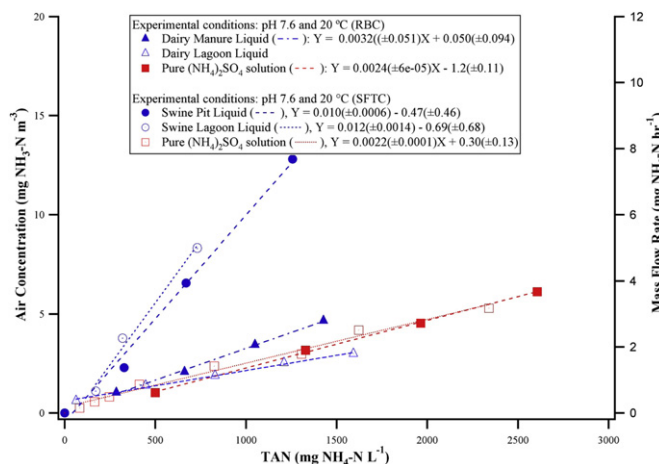


Fig. 4. Comparison of the measured atmospheric  $\text{NH}_3$  release from pure  $(\text{NH}_4)_2\text{SO}_4$  solutions, swine pit liquid (PL), swine lagoon liquid (LL) solutions, dairy manure liquid (DML) solutions, and dairy lagoon liquids (DLL) solutions. Dashed lines represent the respective linear regression models. Original TAN content for DLL was  $<126$   $\text{mg NH}_4\text{-N L}^{-1}$  and for DML was  $<570$   $\text{mg NH}_4\text{-N L}^{-1}$ .

significantly from that observed with the pure AS solutions. The emitted  $\text{NH}_3$  concentration from the swine wastewater sample solutions was  $\sim 5$  times higher than the values from pure AS solutions. Similarly, the emitted  $\text{NH}_3$  concentration from the DML sample was  $\sim 1.4$  times higher than the value from pure AS solutions.

In contrast to DML, DLL samples only demonstrated a slight enhancement in  $\text{NH}_3$  concentrations compared to the pure AS solutions by  $\sim 900$   $\text{mg NH}_4\text{-N L}^{-1}$ . Spiking DLL with solid AS in excess of 900  $\text{mg NH}_4\text{-N L}^{-1}$  actually resulted in a suppression of the  $\text{NH}_3$  emissions. The linearity of  $\text{NH}_3$  concentrations from the DML and DLL samples was investigated at higher TAN concentrations. At higher TAN, the increase in  $\text{NH}_3$  concentrations from DLL appears to once again to become linear with an increase in the TAN. The difference in the absolute magnitude of  $\text{NH}_3$  concentrations for DML and DLL samples at the higher TAN concentrations were directly related to the original TAN concentration in the wastewater samples. Hence, differences in  $\text{NH}_3$  release behavior of DML and DLL samples were attributed to the original TAN content of these samples; e.g., the TAN was relatively low ( $<126$   $\text{mg NH}_4\text{-N L}^{-1}$ ) for DLL compared to that of DML ( $<570$   $\text{mg NH}_4\text{-N L}^{-1}$ ).

A primary focus of this study is to better quantify the contributing reactions that enhance  $\text{NH}_3$  emissions from animal wastewater compared to AS. Results summarized in Fig. 4 suggest that, in general, wastewaters have greater  $\text{NH}_3$  emissions. This suggests that other reactions and processes are influencing  $\text{NH}_3$  emissions and release in wastewater systems. Similar to AS solutions (see Fig. 2), Eq. (2) appears to be the dominant reaction driving  $\text{NH}_3$  emissions in the wastewater at higher TAN concentrations, without any corresponding increase of other constituents in the liquid waste. When compared with AS solutions at similar TAN levels, the enhanced emission of  $\text{NH}_3$  in wastewaters indicates that at additional reactions are contributing to the higher  $\text{NH}_3$  emissions.

### 3.4. Impact of $\text{NaHCO}_3$ on $\text{NH}_3$ emissions

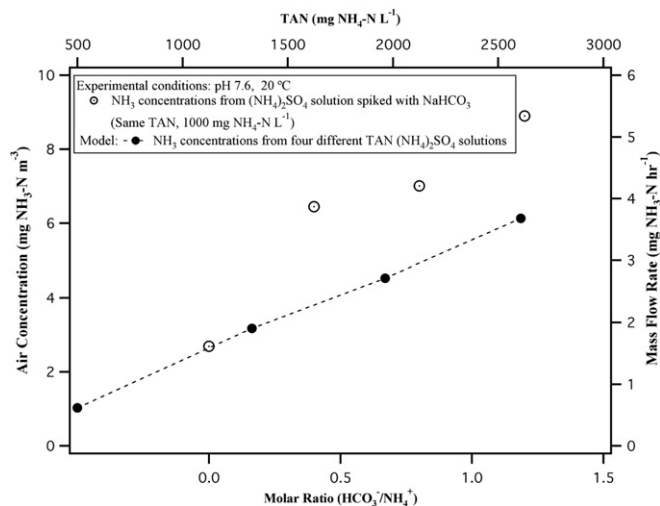
The challenge is to determine why compositional differences in wastewater and pure AS solutions could lead to differences in  $\text{NH}_3(\text{aq})$  concentration near the air–liquid interface. This requires that other reactions are occurring in solution and contributing to dissolved  $\text{NH}_3(\text{aq})$  beyond the basic relationship between  $\text{NH}_4^+$  and dissolved  $\text{NH}_3(\text{aq})$  given in Eq. (2). Animal wastewaters are complex



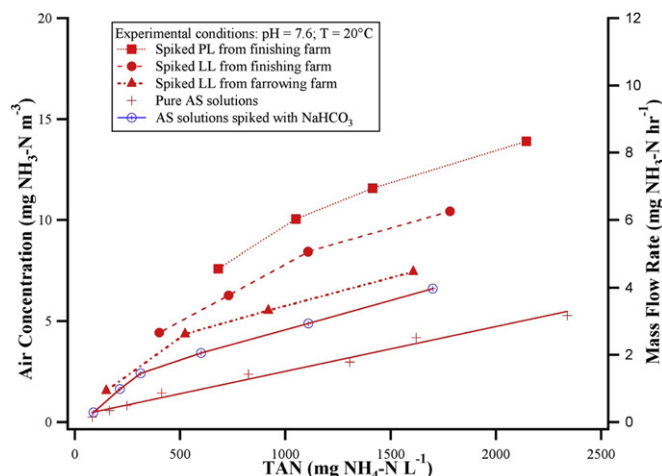
mixtures containing relatively high concentrations of bacteria, dissolved organic compounds, salts, and various fats and oils excreted by the animals or from feed spillage. These wastes also emit  $H_2S$ ,  $CO_2$  and VOCs as well as  $NH_3$  across the air–liquid interface. In particular, elevated levels of dissolved  $CO_2$  could be present in wastewater due to microbial respiration, and  $CO_2$  is expected to play a role in a variety of biogeochemical reactions. Hence, contributing reactions to  $NH_3$  emission could be dependent on dissolved sulfide, carbonate and organic acid. A series of screening tests was therefore conducted to determine whether various agents (surfactant, organic acid, methanol, and sodium bicarbonate) could promote or depress  $NH_3$  emissions from either the pure AS solutions or wastewater. Of the agents used in the screening trials, only bicarbonate produced a response in emissions similar to that observed for PL, LL, DML and DLL (Table S1 in the SI).

Additional RBC experiments were therefore conducted to test whether  $CO_2$  could promote  $NH_3$  emissions from pure AS solutions and animal wastewaters at constant pH ( $pH = 7.6$ ) and temperature ( $T = 20\text{ }^\circ\text{C}$ ), and if a quantitative response could be determined. Sodium bicarbonate was added to simulate the potential influence of elevated partial pressure of  $CO_2$  on emissions. Bicarbonate anion is the dominant form of dissolved carbonate species in solution at  $pH = 7.6$  (Stumm and Morgan, 1995). Results for spiking  $NaHCO_3$  into AS solutions and animal wastewaters at different TAN are presented in Figs. 5 and 6, respectively. Regression equations are summarized in Table S2 in the SI. Enhanced  $NH_3$  emissions were observed in Fig. 5 from AS solutions at the same TAN concentration when spiked with bicarbonate (0.036, 0.056, 0.084 M) (Table S3 in the SI). When these results are plotted as a function of  $HCO_3^-/NH_4^+$  molar ratio, the observed response showed  $\sim 2$  times higher  $NH_3$  emissions than the AS solutions without  $HCO_3^-$ . The rate of increase in  $NH_3$  emissions (derivative of the curve) was higher at lower values of  $HCO_3^-/NH_4^+$  and then decreased at higher TAN. Similarly, addition of  $HCO_3^-$  to the animal wastewaters yielded an initial enhancement in the  $NH_3$  emissions that decreased with increasing TAN (Fig. 6). This would explain why the derivative of the curve in  $NH_3$  emissions at higher TAN concentrations in the animal wastewater approaches that observed for the pure AS solutions.

These observations suggest that the enhancement mechanism/reaction of  $NH_3$  emissions is strictly related to the presence of bicarbonate. In other words, despite the complexity of animal

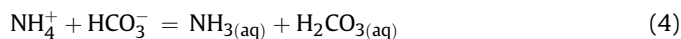


**Fig. 5.** Measurements of atmospheric  $NH_3$  release from pure ammonium sulfate (AS) solutions. AS solutions spiked with  $NaHCO_3$  as a function of TAN ( $pH = 7.6$ ,  $T = 20\text{ }^\circ\text{C}$ ) using RBC. Spiked solutions were generated by adding reagent-grade  $NaHCO_3$  (0, 0.4, 0.8, 1.2 molar ratio of  $HCO_3^-/NH_4^+$ ) to fresh 2 L portions of pure AS solutions of 1000 mg  $NH_4-N\text{ L}^{-1}$  TAN. Dashed lines are added to better visualize trends in  $NH_3$  emissions.



**Fig. 6.** Comparison of the measured atmospheric  $NH_3$  release between original and spiked test solutions of liquid animal waste and pure AS solutions as a function of increasing TAN ( $pH = 7.6$ ,  $T = 20\text{ }^\circ\text{C}$ ). Spiked solutions were generated by adding reagent-grade ammonium sulfate to fresh 10 L portions of liquid animal waste to obtain desired TAN. Dashed lines and zero TAN points added to better visualize trend in  $NH_3$  emissions. Solid line through AS data represents linear regression model.

wastewater, this enhancement is due to the influence of  $HCO_3^-$  either directly or indirectly on the production of  $NH_{3(aq)}$ , which at lower concentrations of TAN, is more efficient in producing  $NH_{3(aq)}$  than Eq. (2). Eq. (2) and the following chemical reactions provide a partial explanation for the observed dependence of  $NH_3$  emission on  $NH_4^+$ ,  $pH$  and  $HCO_3^-$  as:



It should be mentioned that other reactions that involve carbonate ( $CO_3^{2-}$ ) are also possible. However, at  $pH 7.6$  bicarbonate is the dominant carbonate ion ( $\sim 95\%$ ) and dissolved carbonic acid makes up only around 5% of the carbonate ions (Stumm and Morgan, 1995).

Changes in  $NH_3$  flux observed with additions of bicarbonate to the pure AS solutions (Fig. 5) can be explained in part by considering Eqs. (2) and (4)–(6). Similar to Eq. (2), Eq. (4) predicts that  $NH_{3(aq)}$  is related to TAN in a linear fashion. However, the decreasing slope in  $NH_3$  emissions with increasing TAN suggests that Eq. (4) may not be solely involved in enhancing the production of  $NH_{3(aq)}$ , or that Eq. (4) becomes kinetically limited possibly due to transport limitations near the air–liquid interface. For example, loss of  $CO_2$  across the air–liquid interface consumes  $H_2CO_{3(aq)}$  and this would tend to enhance  $NH_3$  emissions. The amount of bicarbonate added to the pure AS solutions (equivalent concentration = 0.02 M) was not limiting at the calculated  $NH_3$  fluxes measured with the RBC. Thus with increasing TAN, the destruction of  $H_2CO_{3(aq)}$  would become the rate-limiting step effectively maximizing the production of  $NH_{3(aq)}$  by Eq. (4). Further production of  $NH_{3(aq)}$  would continue via Eq. (2) with an increase in TAN, but at relatively lower amounts of  $NH_{3(aq)}$  per unit increase in TAN.

The influence of  $HCO_3^-$  on  $NH_3$  emissions from the various wastewaters (Figs. 4 and 6) was even more complicated than in pure AS solutions (Fig. 5). Enhanced emissions from the animal wastewater were related to the initial TAN concentration in the test

solution, with the limit in enhanced  $\text{NH}_3$  emissions substantially exceeding that observed in the spiked pure AS solutions, especially for the LL, PL, and DML samples. If the rate of loss of  $\text{CO}_2$  across the air–liquid interface (which consumes  $\text{H}_2\text{CO}_{3(\text{aq})}$ ) was the limiting step, it is not clear why this restriction was present in the pure AS solutions, but not in the LL, PL, DML and DLL solutions at the same TAN concentration. The apparent dependence on the concentration of TAN and the enhanced  $\text{NH}_3$  emissions suggests a link between TAN and the enhancement factor in the wastewater. For example, higher TAN in the wastewater could be associated with higher dissolved solids and higher respiration rates of bacteria in solution. Higher production of  $\text{CO}_2$  in solution would drive aqueous carbonate chemistry to produce higher concentrations of bicarbonate in solution. However, this would not necessarily increase the production of  $\text{NH}_{3(\text{aq})}$  if the rate of loss of  $\text{CO}_2$  across the air–liquid interface is the rate limiting step. It would therefore appear that a number of factors are influencing the extent of the reaction for Eq. (4) that eventually limits the production of  $\text{NH}_{3(\text{aq})}$  as a function of TAN in solution. The factors could include the overall energetics of Eq. (4) as well as possible transport of reactive species from the bulk solution to the region near the air–liquid interface. Additional research studies are needed to fully resolve these issues.

### 3.5. Modeling implications

Ammonia emissions increased in a linear fashion with increasing TAN for the pure AS solutions (Fig. 2). This was consistent with the expected results (Eq. (2)) and the assumption that the Henry's Law coefficient is indeed constant over a range of dissolved  $\text{NH}_{3(\text{aq})}$  in solution. The linearity in  $\text{NH}_3$  emissions from spiked animal wastewater (Fig. 3) also suggests that the value of the Henry's Law coefficient remained unchanged. Consequently, it is logical to assume that the enhanced emission of  $\text{NH}_3$  from wastewater compared with pure AS solutions (Fig. 4) was due to differences in the concentration of dissolved  $\text{NH}_{3(\text{aq})}$  near the air–liquid interface as a result of  $\text{HCO}_3^-$ .

## 4. Conclusions

The results from this study suggest that current models such as thin-film and process-based models being developed to predict  $\text{NH}_3$  emissions from AFOs waste liquid system could be in error if it is assumed a single reaction is controlling  $\text{NH}_3$  emissions across the air–liquid interface (i.e. Eq. (2)). While any model can be made to fit observed data, failure to account for the apparent complexity in reactions controlling  $\text{NH}_3$  emissions from animal wastewater have the potential to generate misleading results when used to test the effects on  $\text{NH}_3$  emissions from possible remediation strategies or alternative management practices to handle the waste.

## Appendix A. Supplementary data

Supplementary data related to this article can be found at <http://dx.doi.org/10.1016/j.atmosenv.2013.01.046>.

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