

Chemically synthesized antimicrobial peptides inhibit *in vitro* growth of *Campylobacter* spp.





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Introduction and Hypothesis

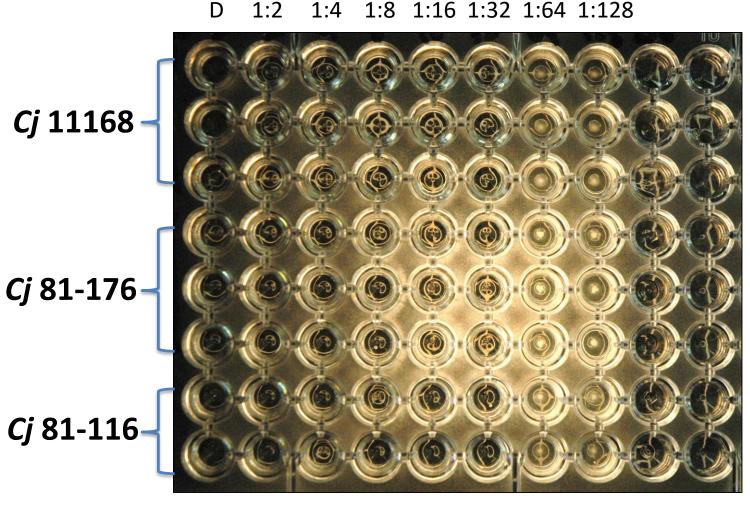
Novel alternatives to traditional antibiotics are needed for foodanimal production. One goal of our laboratories is to discover and evaluate antimicrobial peptides (AMP) to reduce foodborne bacterial pathogens during poultry production. AMPs permeabilize membranes and are found in most every class of living organism where they have evolved as a defense mechanism against invading microorganisms. Our working hypothesis is that AMPs can be identified that inhibit growth of *Campylobacter jejuni* then subsequently utilized to reduce gastrointestinal *Campylobacter* among commercially produced chickens.

RESULTS CONTINUED

Table 1. Formation of zones ofinhibitionbyAMPagainstvarious target bacteria.

	1. C12K-2β12	2. Cecropin A-	3. RL-37	4. Water-	
et Bacteria		Magainin 2		Control	

Figure 2. Typical MIC response for AMP RL-37 in microtiter well assay.

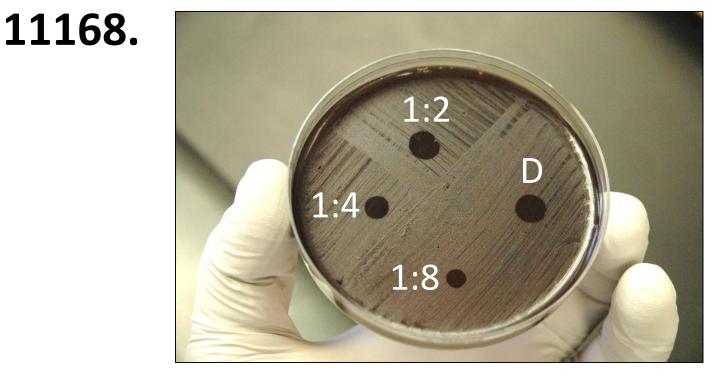


METHODS AND SUMMARY OF RESULTS

A set of 11 unique AMPs chemically synthesized commercially and evaluated for ability to inhibit growth of two *C. jejuni* strains. Six of the AMPs assayed produced zones of inhibition on lawns of *C. jejuni*. These included: NRC-13, a variant of pleurocidin isolated from the American plaice-flounder; RL-37, a **37-residue AMP of the cathelicidin family which is expressed in bone** marrow of the rhesus monkey; temporin, from the frog, Rana temporaria; a potent hybrid AMP (Cec-Mag) composed of residues 1-8 of cecropin A (from the *Cecropia* moth) fused to residues 1-12 of magainin 2 (from the African clawed frog, *Xenopus laevis*); dermaseptin from the skin of *Phyllomedusa* frogs; and the synthetic OAK, C12K-2b12. Three AMPs were chosen for further investigation on the basis of reported reduced cytotoxicity to mammalian cells: Cec-Mag, RL-37 and C12K-2b12. These AMPs produced zones of inhibition on lawn assays against 19 different bacteria, including C. *jejuni, C. coli* and *C. lari* as well as two strains of *Salmonella* and *Lactobacillus*. Modifications of the NCCLS M26A and Hancock assays were utilized to determine minimum inhibitory concentrations (MIC) in microtiter plates for these AMPs against three strains of *C. jejuni*. MICs were approximately 3.1 ug/ml for the AMP RL-37 and C12K-2b12, while the MIC for Cec-Mag was in the range of 12.5 to 50 ug/ml.

Cj 14118	Pos	Pos	Pos	Neg	
Cj 81-116	Pos	Pos	Pos	Neg	
Cj 81-176ª	Pos	Pos	Pos	Neg	
Cj 11168^a	Pos	Wk	Pos	Neg	
Cj RM1221ª	Pos	Pos	Pos	Neg	
Cj A74C	Pos	Pos	Pos	Neg	(
Cj A49943*	Pos	Pos	Pos	Neg	
Cj A33250*	Pos	Pos	Pos	Neg	
Cj A29428*	Pos	Pos	Pos	Neg	
Cc Epi 33-WT	Pos	Pos	Pos	Neg	
Cc A49941*	Pos	Pos	Pos	Neg	
Cc A33559*	Pos	Pos	Pos	Neg	
CI RM2100	Pos	Pos	Pos	Neg	
Cl A35221*	Pos	Pos	Pos	Neg	
Cl "slaughter beach"	Pos	Pos	Pos	Neg	
S. Typhimurium Epi 3	Pos	Pos	Pos	Neg	
S. Heidelberg Epi 42	Pos	Pos	Pos	Neg	
L. acidophilus-WT	Pos	Pos	Pos	Neg	
L. helveticus-WT	Pos	Neg	Neg	Neg	

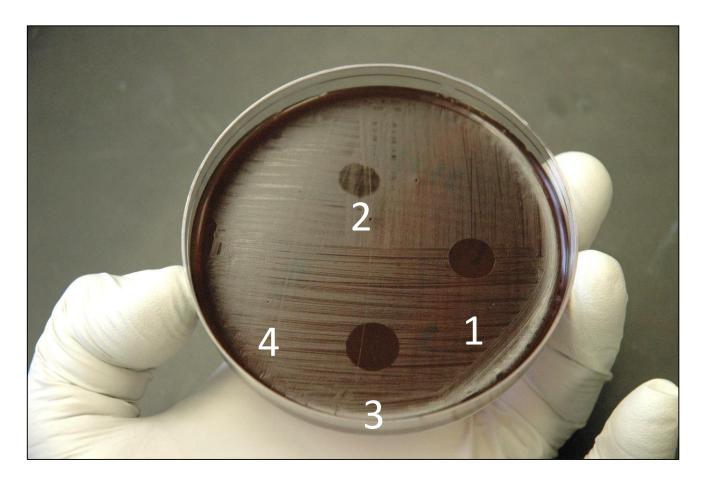
Pos = inhibition zone acquired Neg = no zone of inhibition Wk = weak activity without full zone of inhibition ^a Alternate ATCC designations [^]National Collection of Type Cultures (NCTC) isolate *American Type Culture Collection (ATCC) isolate Cj = Campylobacter jejuni Cc = Campylobacter coli Cc Epi 33 (3309-61099A hog) Cl = Campylobacter lari L = Lactobacillus Salmonella enterica serotype Typhimurium Salmonella enterica serotype Heidelberg WT = wild type Epi = Acquired from epidemiological surveillance Figure 3. Typical response for diluted AMP C12K-2β12 against *Cj*



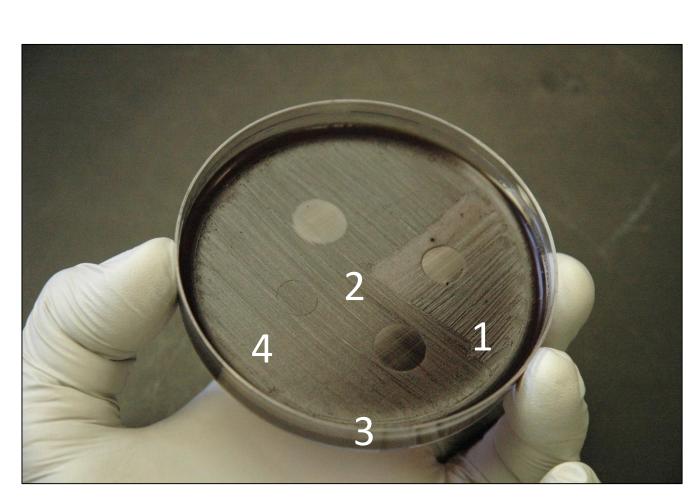
CONCLUSIONS

• The selected AMPs produced obvious zones of inhibition against growth of *C. jejuni*, *C. coli* and *C. lari* isolates in the classic spot-on lawn plating assay.

RESULTS

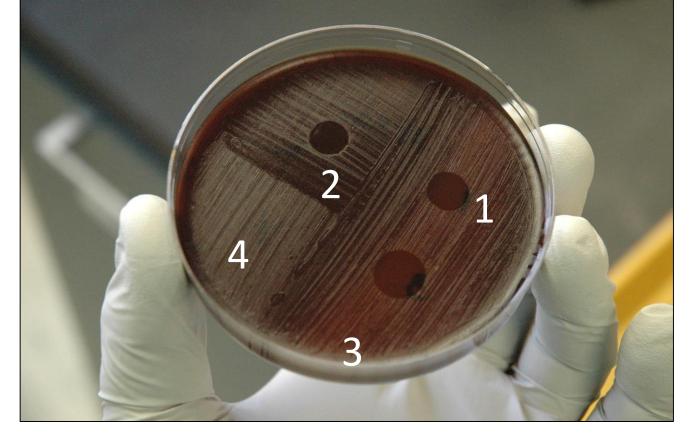


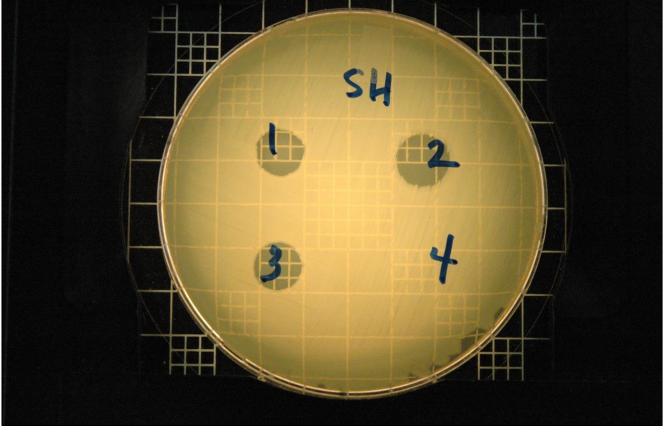
Campylobacter jejuni A74C



Campylobacter coli 49941

- MICs were approximately 3.1 ug/ml for the AMP RL-37 and C12K-2b12.
- MICs were slightly higher for the Cec-Mag AMP in the range of 12.5 to 50 ug/ml.
- The selected AMPs also inhibited growth of a pathogenic Salmonella isolate.
- Research must be done to determine the effect of AMP on the non-pathogenic natural flora of broiler chickens.
- Our next approaches are to express AMPs in yeast and explore encapsulation technologies to stabilize the AMP for *in vivo* trials in broiler chickens.





Campylobacter lari A35221

Salmonella enterica serotype Heidelberg

Figure 1. Examples of AMP zones of inhibition in a spot-on-lawn assay of 1) C12K-2β12; 2) Cecropin A-Maganin 2; 3) RL-37 and 4) sterile water.

All AMPs examined produced zones of inhibition on lawns of *Campylobacter* spp. and *Salmonella enterica* type Heidelberg. Sterile water and PBS did not cause zones of inhibition.

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