

# Global transmission of influenza viruses from humans to swine

Martha I. Nelson<sup>1\*</sup>, Marie R. Gramer<sup>2</sup>, Amy L. Vincent<sup>3</sup>, & Edward C. Holmes<sup>1,4</sup>

<sup>1</sup>Fogarty International Center, National Institutes of Health, Bethesda, MD 20892, USA.

<sup>2</sup>University of Minnesota Veterinary Diagnostic Laboratory, St. Paul, MN 55108, USA.

<sup>3</sup>Virus and Prion Diseases Research Unit, National Animal Disease Center, USDA-ARS,  
Ames, IA 50010, USA.

<sup>4</sup>Center for Infectious Disease Dynamics, Department of Biology, The Pennsylvania State  
University, University Park, PA 16802, USA.

\*Corresponding author. Fogarty International Center, National Institutes of Health, 16 Center  
Drive, Building 16, Room 202, Bethesda, MD 20892. Phone: (301) 402-5203. Fax: (301) 496-  
8496. E-mail: [nelsonma@mail.nih.gov](mailto:nelsonma@mail.nih.gov).

Abstract = 136 words, Main text = 3,326 words, 2 Tables, 1 Figure, 2 Supplementary Tables, 7  
Supplementary Figures

Key words = Influenza, swine, evolution, pandemic, cross-species transmission

23 **To determine the extent to which influenza viruses jump between humans and swine**  
24 **hosts, we undertook a large-scale phylogenetic analysis of pandemic A/H1N1/09**  
25 **(H1N1pdm09) influenza virus genome sequence data. From this we identified at least 49**  
26 **human-to-swine transmission events that occurred globally during 2009-2011, thereby**  
27 **highlighting the ability of the H1N1pdm09 virus to repeatedly transmit from humans to**  
28 **swine, even following adaptive evolution in humans. Similarly, we identified at least 23**  
29 **separate introductions of human seasonal (non-pandemic) H1 and H3 influenza viruses**  
30 **into swine globally since 1990. Overall, these results reveal the frequency with which**  
31 **swine are exposed to human influenza viruses, indicate that humans make a substantial**  
32 **contribution to the genetic diversity of influenza viruses in swine, and emphasize the**  
33 **need to improve biosecurity measures at the human-swine interface, including influenza**  
34 **vaccination of swine workers.**

## 35 INTRODUCTION

36 The 2009 pandemic H1N1 virus (H1N1pdm09) represents the best documented emergence of a  
37 swine pathogen in humans, particularly as the virus was associated with widespread global  
38 morbidity, mortality, and life-years lost in humans (Viboud *et al.*, 2010). The H1N1pdm09 virus  
39 was generated by a reassortment event between Eurasian swine H1N1 influenza viruses and  
40 North American triple reassortant H1 viruses, with the former contributing the N1 and M  
41 segments and the latter donating the PB2, PB1, PA, H1, NP, and NS segments (Garten *et al.*,  
42 2009). Although similar influenza virus reassortants containing segments of both Eurasian  
43 swine virus and triple reassortant swine virus origins have been detected in Asia (Lam *et al.*,  
44 2010; Smith *et al.*, 2009), no ‘smoking gun’ viruses that are closely related progenitors have  
45 been detected in swine in any locality.

46 Following the identification of H1N1pdm09 in humans in April 2009, the virus was rapidly  
47 transmitted from humans back into swine. The first isolation of a H1N1pdm09 virus in swine  
48 was from a pig farm in Alberta, Canada in May 2009 (Howden *et al.*, 2009), and the  
49 H1N1pdm09 virus was subsequently isolated from outbreaks in swine in all other major  
50 continents: Africa (Njabo *et al.*, 2011), Asia (Kim *et al.*, 2011; Sreta *et al.*, 2010), Australia  
51 (Holyoake *et al.*, 2011), Europe (Hofshagen *et al.*, 2009; Howard *et al.*, 2011), and South  
52 America (Pereda *et al.*, 2010). Upon introduction into the swine population, H1N1pdm09  
53 viruses co-circulated with endemic swine influenza viruses and exchanged genome segments  
54 via reassortment (Ducatez *et al.*, 2011; Lam *et al.*, 2010; Vijaykrishma *et al.*, 2010). In 2011  
55 twelve reassortant H3N2 (H3N2v) swine influenza viruses with matrix (M) segments of  
56 pandemic H1N1pdm09 origin were isolated from humans in the United States (Anonymous  
57 2012).

58 Seasonal human influenza viruses have also been isolated periodically from swine, with  
59 a few major lineages becoming endemic in swine: most notably the human-origin H3N2 viruses  
60 that proliferated in European swine in the 1970s (Ottis *et al.*, 1982), the H1 $\delta$  lineage of human

61 origin that emerged in North American swine in 2003 (Karasin *et al.*, 2006), and the human H3,  
62 N2, and PB1 segments that were associated with the triple reassortant viruses that were  
63 identified in 1998 in North America swine (Zhou *et al.*, 1999). In addition, singletons and smaller  
64 clusters of swine influenza viruses have been identified that contain one or more segments of  
65 human seasonal influenza virus origin in several countries, including Argentina (Cappuccio *et*  
66 *al.*, 2011; Pereda *et al.*, 2011) and Italy (Moreno *et al.*, 2012).

67 The increased global surveillance of influenza in pigs provides the first opportunity to  
68 estimate the extent of human-to-swine transmission of the H1N1pdm09 virus. The main aim of  
69 our study was therefore to estimate, using a simple phylogenetic approach, the total number of  
70 introductions of the H1N1pdm09 virus from humans into swine using a data set of H1N1pdm09  
71 viruses collected globally in swine during 2009-2011. We then compared this estimate with the  
72 equivalent transmission frequency of human seasonal (non-pandemic) H1 and H3 into swine  
73 since 1990.

74

## 75 **RESULTS**

76 **Frequent transmission of H1N1pdm09 virus from humans into swine.** Across the HA, NA,  
77 and concatenated internal gene phylogenies we identified a total of 49 discrete introductions of  
78 H1N1pdm09 influenza virus from humans into swine during 2009-2011 (Table 1, Figs. 1, S1-  
79 S3). These introductions were observed in 12 countries and semi-autonomous regions:  
80 Australia, Canada, China, Costa Rica, Cuba, Hong Kong (SAR), Italy, Singapore, South Korea,  
81 Taiwan, Thailand, and the United States. The majority of viral introductions into swine were  
82 identified in the United States and Canada (19 and 8 introductions, respectively, Table 1),  
83 reflecting the higher number of H1N1pdm09 influenza virus sequences available from North  
84 American swine. Given our strict criteria for defining viral introductions (bootstrap values  $\geq 70\%$ )  
85 and limited global sampling, these numbers are likely to significantly underestimate the extent of  
86 global spillover from humans to swine of the H1N1pdm09 virus. Our conservative methods are

87 particularly prone to miss introductions involving single isolates. For example, single  
88 H1N1pdm09 isolates from Mexico, Japan, England, Cameroon, Brazil, and Colombia  
89 (A/sw/4/Mexico/2009, A/sw/Osaka/1/2009, A/sw/England/73690/2010,  
90 A/sw/Cameroon/11rs149-198/2010, A/sw/Brazil/12A/2010, A/sw/Colombia/1403/2010) are likely  
91 to represent additional introductions. These additional probable introductions are identified by #  
92 symbols on the detailed HA, NA, and concatenated phylogenies presented in Figs. S1-S3.

93

94 **Frequency of human-to-swine transmission of seasonal influenza viruses.** Across the H1,  
95 H3, N1, and N2 phylogenies, at least 23 separate introductions of seasonal influenza virus from  
96 humans to swine were observed during the period 1990-2011 (Figs. S4-S7). The vast majority  
97 of these introductions were identified after 1996, when surveillance of influenza virus in swine  
98 increased. Six human-to-swine transmission events involved the human seasonal H1 segment,  
99 eight involved the human H3 segment, six involved the human N1 segment, and 16 involved the  
100 human N2 segment (Table 2). The higher number of introductions of the human N2 segment is  
101 likely related to several factors, including (a) the overall higher number of H3N2 virus  
102 introductions (n = 12) due to the dominance of the H3N2 subtype in humans during the period of  
103 this study, (b) the frequency of introductions (n = 3) of the human H1N2 variant that circulated  
104 globally in humans from 2001-2003, and (c) the frequency of reassortment involving N2  
105 segments of human origin and HA segments of swine origin. For example, the 20 H1N2  
106 reassortant viruses from Italy that are associated with introduction #11 have N2 sequences that  
107 are most closely related to the N2 of human seasonal H3N2 isolates collected in 1997-1998, but  
108 H1 sequences related to European swine influenza viruses.

109 Twelve of the introductions of seasonal influenza virus were first identified in Asia, six  
110 were first identified in the United States, three in Canada, and two in Europe (Table 2). As with  
111 the H1N1pdm09 viruses, the methods we used to define a human-to-swine transmission event  
112 are highly conservative, and numerous probable introductions were excluded from our final

113 estimate, particularly those involving singletons. Six singleton swine isolates, including three  
114 from Argentina, are highlighted that were identified to be of human origin on both the HA and  
115 NA trees but which were not supported by high bootstrap values (letters a-g, Table 2, Figs. S4-  
116 S7). Additionally, eight swine H3N2 isolates collected in Hong Kong during 2000-2002 are  
117 interspersed with human isolates collected from that same time period on both the H3 and N2  
118 phylogenies, suggesting additional introductions of human H3N2 viruses into swine in Hong  
119 Kong, although again lacking the phylogenetic resolution required to be defined as discrete  
120 introductions.

121

122 **Estimated time periods of human-to-swine transmission.** The long branch lengths  
123 associated with several introductions of human-origin influenza viruses into swine indicate that  
124 numerous viral introductions circulated in swine for many years before being detected. Given  
125 the high intensity of sampling of influenza virus in humans, the duration of unsampled circulation  
126 in swine can be estimated simply and directly by the differences in time between when the  
127 isolates were collected in swine and when the most closely related human isolates were  
128 collected. For example, the eight H3N2 viruses associated with introduction #13 were collected  
129 in swine in Thailand in 2004-2009, but are most closely related to A/Wuhan/359/1995(H3N2)-  
130 like viruses from 1995-1996 on the H3 and N2 phylogenies (Figs. S4 and S6, respectively),  
131 representing eight years of unsampled circulation in swine. The time difference between the  
132 2008 H3N2 and 2010 H1N2 isolates collected in swine in Argentina and the most closely related  
133 human seasonal isolates is approximately 6-8 years. Likewise, the isolate  
134 A/sw/Argentina/CIP051-StaFeN2/2010(H1N2) is most closely related to the reassortant H1N2  
135 viruses that circulated in humans during 2002-2003 (Figs. S5 and S6), suggesting that this virus  
136 has circulated undetected in swine in Argentina or elsewhere since at least 2003, when the  
137 H1N2 viruses disappeared globally in humans. Incorporating more viral sequence data from  
138 swine by also including partial HA (HA1) sequences filled in some of these gaps in surveillance

139 (H1-HA1 and H3-HA1 trees available from the authors upon request). For example, the USA  
140 2007-2011 ( $\delta$ -1) introduction (#18) is detected two years earlier in 2005 when HA1 data is  
141 included. Finally, eleven years separate the three H3N2 swine isolates collected in South Korea  
142 in 2007 (introduction #17) from the most phylogenetically related human seasonal H3N2 viruses  
143 collected in 1995-1996. However, the relatively short branch lengths associated with these  
144 2007 South Korean isolates raises the question of whether the evolutionary rate was particularly  
145 low on this branch, or whether these sequences could be erroneous. The short branch length  
146 that separates A/sw/Fujian/0325/2008(H1N1) (introduction #19) from the most closely related  
147 human seasonal H1N1 viruses collected in 2000-01, relative to the ~7-8 year difference in  
148 collection date, also raises the question of possible sequencing error.

149

150 **Spatial patterns of human-origin influenza viruses in swine.** The number of swine  
151 H1N1pdm09 isolates associated with each introduction (ranging from 1-10 isolates, Table 1)  
152 represents the extent of onward swine-to-swine transmission that can be inferred from our  
153 phylogenetic analysis. Although the vast majority of introductions of human H1N1pdm09  
154 viruses into swine exhibit local onward transmission in pigs, reflected in introductions that are  
155 associated with >1 swine isolate, we infer that global viral movement has not yet been important  
156 in the dissemination of the H1N1pdm09 virus in swine. Specifically, each cluster of swine  
157 H1N1pdm09 influenza viruses that are associated with a human-to-swine transmission event is  
158 highly spatially structured, with no swine introduction including viruses collected in more than  
159 one country. However, within-country spread of H1N1pdm09 viruses has occurred in the United  
160 States, as six viral US introductions include multiple states, representing viral dissemination  
161 within the Midwestern US (Table 1). Limited spread within Canada between Alberta and  
162 Manitoba was also detected (introduction #2, Table 1). Within-country spread was not observed  
163 in other countries where information on the province or region of viral collection was also  
164 available, including China and Australia.

165 Similarly, 18 of the 23 introductions of human seasonal H1 and H3 influenza viruses into  
166 swine involve a single country (Table 2). Six of these single-country introductions involve  
167 singleton swine influenza viruses, with no evidence of onward transmission. In contrast, four  
168 viral introductions include isolates collected in different countries, indicating viral migration within  
169 North American and into and within Asia. Aside from the major introduction of triple reassortant  
170 H3N2 viruses that spread from North America to Asia (introduction #4) and from the United  
171 States to Canada (introduction #10), possible limited viral migration occurred between Taiwan  
172 and Indonesia (introduction #9) and the US and South Korea (introduction #8).

173

174

## 175 **DISCUSSION**

176 Despite evolving the capability to transmit efficiently in humans, our results indicate that  
177 the H1N1pdm09 influenza virus has retained its ability to transmit back into swine and to co-  
178 circulate with other swine influenza viruses, increasing genetic diversity and providing the  
179 opportunity for reassortment. The large number of H1N1pdm09 transmission events observed  
180 in this study, even using conservative methods, highlights the frequency of human-swine  
181 contact rates globally and the continual exposure of swine to human influenza viruses. Indeed,  
182 this transmission rate is considerably higher than would be inferred from the relatively lower  
183 number ( $n = 23$ ) of human-to-swine transmission events of seasonal H1 and H3 influenza  
184 viruses that were identified during the past two decades. Importantly, detection of seasonal  
185 human influenza viruses in swine did not increase during 2009-2011, despite widespread global  
186 circulation of seasonal H3N2 viruses in humans during 2010-2011. Hence, the different  
187 transmission frequencies observed are not explained by the increase in swine influenza  
188 surveillance since 2009 and are unlikely to be an artifact of sampling. Furthermore, the low  
189 phylogenetic resolution of the H1N1pdm09 phylogeny, due to the virus's recent emergence and



190 relatively low genetic diversity, impeded the identification of what are likely to be many more  
191 introductions of this virus into swine than could be defined here.

192         Although it is tempting to compare the frequencies of human-to-swine transmission with  
193 the 27 swine-origin influenza viruses that were identified in humans in the United States over a  
194 similar time period as our study (1990-2010) (Shu *et al.*, 2012), such a comparison is greatly  
195 biased by the substantially higher levels of global surveillance of influenza in humans than  
196 consistently occurs in swine. However, the intensity of influenza surveillance in humans does  
197 facilitate estimation of the time of emergence of the 23 introductions of human seasonal  
198 influenza viruses into swine. For example, the introductions of H1N1pdm09 viruses were  
199 generally detected in swine within a year, whereas several of the human seasonal influenza  
200 viruses that were introduced into swine were not detected for 5-10 years. While the time to  
201 detection in swine may relate to intrinsic viral characteristics (e.g., the virus must first adapt,  
202 then replicate competently in the new host, then spread to many hosts before being detected), it  
203 is also possible that the difference in time between when a human-to-swine transmission event  
204 is detected in swine and the year of collection of the most closely related human influenza  
205 viruses provides an indication of intensity of swine surveillance in a given region. The smallest  
206 gaps in detection are in Canada, the US, and Hong Kong, where ongoing viral surveillance is  
207 conducted in swine, while larger time gaps occurred in countries such as South Korea, Italy,  
208 Thailand, and Argentina, where sampling appears to be more sporadic, based on publicly  
209 available sequence data.

210         The global frequency of introductions of H1N1pdm09 viruses into swine also provides an  
211 opportunity to track, in detail, the onward transmission and spatial dissemination of these new  
212 viral clades and how they relate to swine movements (Nelson *et al.*, 2011). It will be particularly  
213 important to determine which of these H1N1pdm09 lineages sustains transmission in swine over  
214 the long term, to observe their population dynamics in a new swine host, and to identify which  
215 lineages spread globally. The multiple introductions of human influenza viruses into swine

216 provide a natural experiment for observing adaptive evolution of influenza from human to swine  
217 hosts, in identifying instances of parallel adaptive evolution, and in the comparison of rates of  
218 evolutionary change and selection pressures along the branches that define human-to-swine  
219 transmission events. Clearly, the ability to trace such dynamics relies upon the continued  
220 surveillance of influenza in swine globally, even after the initial interest in the 2009 H1N1  
221 pandemic has subsided.

222

## 223 **METHODS**

224 **Sequence data used in this study.** To estimate the number of human-to-swine transmission  
225 events of the H1N1pdm09 virus, we compiled gene sequence data from 263 H1N1pdm09  
226 influenza viruses that were collected in swine during 2009-2011 in 18 countries: Argentina,  
227 Australia, Brazil, Cameroon, Canada, China, Colombia, Costa Rica, Cuba, England, Hong  
228 Kong, Japan, Mexico, Singapore, South Korea, Taiwan, Thailand, and the United States (Table  
229 S1). Due to the frequency of reassortment between H1N1pdm09 viruses and other swine  
230 influenza viruses involving the HA and NA segments, concatenated internal gene segments  
231 (PB2, PB1, PA, NP, M, and NS) and the HA and NA segments were studied separately.  
232 Separate phylogenies were also inferred for each of the internal gene segments to identify and  
233 remove any viruses with reassorted internal genes. As background data, 1,008 complete  
234 genome sequences from H1N1pdm09 influenza viruses that were collected in humans during  
235 2009-2011 were downloaded from the Influenza Virus Resource on GenBank (due to the size of  
236 the original data set ( $n = 2,718$  genomes), 50 genome sequences were randomly selected from  
237 highly sampled countries (i.e. those with  $>50$  genome sequences)) (Bao *et al.*, 2008).

238 To identify swine influenza viruses related to human seasonal H1 and H3 influenza  
239 viruses, four phylogenetic trees were inferred using all full-length H1 ( $n = 991$ , excluding  
240 pandemic viruses), H3 ( $n = 326$ ), N1 ( $n = 858$ ), and N2 sequences ( $n = 559$ ) collected from  
241 swine since 1990 that are available on GenBank (accession numbers available from the

242 authors). Due to the frequency of reassortment and low availability of whole-genome sequence  
243 data for these swine viral isolates, alignments of the concatenated internal genome segments  
244 were not included for this part of the study. These data came from Argentina, Belgium, Canada,  
245 China, Denmark, France, Germany, Hong Kong, Hungary, Italy, Japan, Spain, South Korea,  
246 Sweden, Taiwan, Thailand, the United Kingdom, and the United States of America. From these  
247 phylogenies, 152 human-origin swine H1 sequences, 221 human-origin swine H3 sequences,  
248 18 human-origin swine N1 sequences, and 430 human-origin swine N2 sequences were  
249 identified globally and used in the final analysis. This analysis also included, as background  
250 data, 1,000 randomly selected human seasonal H1, H3, N1, and N2 segments collected from  
251 1990-2011. A large clade of European sequences (e.g. A/sw/Scotland/410440/1994) that were  
252 of human seasonal H1 virus origin were not included because they clearly were introduced from  
253 humans into swine at least a decade prior to 1990, the beginning of our study period.

254 To identify any additional human-to-swine transmission events that could not be  
255 detected from trees inferred with full-length HA sequence data, two additional phylogenies were  
256 inferred using all partial HA1 sequence data (918 nt) from the swine H1 (n = 1,200 sequences)  
257 and H3 (n = 1,290 sequences) subtypes available since 1990, including HA1 sequence data  
258 from the same 1,000 human influenza viruses that were selected as background in the full-  
259 length HA phylogenies. Using HA1 data, we identified 50 additional swine H1 sequences of  
260 human origin and 70 additional swine H3 sequences of human origin. However, all but two  
261 isolates (A/sw/Obihiro/3/1993(H3N2) and A/sw/Obihiro/1/1994(H3N2)) were associated with  
262 introductions that are already identified in the analysis of full-length HA and NA sequences.

263

264 **Phylogenetic analysis of H1N1pdm09.** Alignments were manually constructed using the SeAl  
265 program (Rambaut *et al.*, 2002) for three sets of swine H1N1pdm09 virus sequence data, with  
266 the 1,008 human H1N1pdm09 isolates included as background: (i) the HA segment (n = 232  
267 swine H1N1pdm09 isolates, 199 of which were downloaded from GenBank, and 33 provided by

268 the University of Minnesota Veterinary Diagnostic Laboratory (UMVDL) from samples collected  
269 from their routine veterinary diagnostic laboratory submission and/or tested via the US  
270 Department of Agriculture (USDA) National Animal Health Laboratory Network (NAHLN Swine  
271 Influenza Surveillance System); (ii) the NA segment (n = 229 swine H1N1pdm09 isolates, 197  
272 from GenBank and 32 from UMVDL); and (iii) concatenated internal gene segments (n = 100  
273 swine H1N1pdm09 isolates, all downloaded from GenBank, with all reassortants identified from  
274 individual gene trees and excluded). For each alignment a maximum likelihood (ML) tree was  
275 inferred using the PhyML v3.0 program (Guindon *et al.*, 2010), employing SPR branch-  
276 swapping and a general time reversible model (GTR) model of nucleotide substitution with  
277 gamma-distributed rate variation among sites. Statistical support for individual nodes was  
278 estimated using 1000 replicate neighbor-joining trees inferred using PAUP\* and assuming the  
279 GTR + gamma model of nucleotide substitution (Swofford 2003). Well-supported nodes ( $\geq 70\%$   
280 bootstrap support) defining human-to-swine viral introductions were identified visually on each  
281 phylogeny. To mitigate possible effects of sequencing error, the several isolates that were  
282 separated by extremely long branch lengths were only categorized as discrete introductions  
283 when multiple isolates from the same country exhibited the same phylogenetic pattern.

284

285 **Phylogenetic analysis of seasonal H1 and H3.** Alignments were manually constructed using  
286 SeAl for four sets of swine influenza virus sequences that were identified as of human seasonal  
287 (non-pandemic) influenza virus origin: (i) 152 H1 sequences, (ii) 221 H3 sequences, (iii) 18 N1  
288 sequences, and (iv) 430 N2 sequences. As background, 1,000 human influenza viruses were  
289 randomly selected for each segment and downloaded from GenBank. For each alignment a  
290 maximum likelihood (ML) tree was inferred as described above to identify well-supported nodes  
291 ( $\geq 70\%$  bootstrap support) defining introductions of influenza viruses from humans to swine. The  
292 estimated time periods of human-to-swine transmission were inferred directly from the  
293 phylogeny by identifying the most closely related human viruses (the sampling date of which is

294 known). This simple phylogenetic method produced similar results (i.e. times to common  
295 ancestry) to those obtained using the Bayesian Markov chain Monte Carlo approach available in  
296 the BEAST program (Drummond & Rambaut 2007) and performed during a previous analysis of  
297 human influenza viruses that were transmitted to swine (Nelson *et al.*, 2011).

298

299

300

301 **ACKNOWLEDGEMENTS**

302 This research was conducted within the context of the Multinational Influenza Seasonal Mortality  
303 Study (MISMS), an on-going international collaborative effort to understand influenza  
304 epidemiology and evolution, led by the Fogarty International Center, NIH, with funding from the  
305 Office of Global Affairs at the Department of Health and Human Services (DHHS).

306

## REFERENCES

1. **Anonymous (2012).** Update: influenza A (H3N2)v transmission and guidelines – five states, 2011. *MMWR Morb Mortal Wkly Rep* **60**, 1741-1744.
2. **Bao Y., Bolotov, P., Dernovoy, D., Kiryutin, B., Zaslavsky, L., Tatusova, T., Ostell, J. & Lipman, D. (2008).** The influenza virus resource at the National Center for Biotechnology Information. *J Virol* **82**, 596-601.
3. **Cappuccio, J.A., Pena, L., Diárbora, M., Rimondi, A., Piñeyro, P., Insarralde, L., Quiroga, M.A., Machuca, M., Craig, M.I. & other authors (2011).** Outbreak of swine influenza in Argentina reveals non-contemporary human H3N2 virus highly transmissible among pigs. *J Gen Virol* **92**, 2871-2878.
4. **Drummond, A. J. & Rambaut, A. (2007).** BEAST: Bayesian evolutionary analysis by sampling trees. *BMC Evol Biol* **7**, 214.
5. **Ducatez, M.F., Hause, B., Stigger-Rosser, E., Darnell, D., Corzo, C., Juleen, K., Simonson, R., Brockwell-Staats, C., Rubrum, A. & other authors (2011).** Multiple reassortment between pandemic (H1N1) 2009 and endemic influenza viruses in pigs, United States. *Emerg Infect Dis* **17**, 1624-1629.
6. **Garten, R.J., Davis, C.T., Russell, C.A., Shu, B., Lindstrom, S., Balish, A., Sessions, W.M., Xu, X., Skepner, E. and other authors (2009).** Antigenic and genetic characteristics of swine-origin 2009 A(H1N1) influenza viruses circulating in humans. *Science* **325**,197-201.
7. **Guindon, S., Dufayard, J.F., Lefort, V., Anisimova, M., Jordijk, W. & Gascuel, O. (2010).** New algorithms and methods to estimate maximum-likelihood phylogenies: assessing the performance of PhyML 3.0. *Syst Biol* **59**, 301-321.
8. **Hofshagen, M., Gjerset, B., Er, C., Tarpai, A., Brun, E., Dannevig, B., Bruheim, T., Fostad, I.G., Iversen, B. & other authors (2009).** Pandemic influenza A(H1N1)v: human to pig transmission in Norway? *Euro Surveill* **14**, 19406.

9. **Holyoake, P.K., Kirkland, P.D., Davis, R.J., Arzey, K.E., Watson, J., Lunt, R.A., Wang, J., Wong, F., Moloney, B.J. & other authors (2011).** The first identified case of pandemic H1N1 influenza in pigs in Australia. *Aust Vet J* **89**, 427-431.
10. **Howard, W.A., Essen, S.C., Strugnell, B.W., Russell, C., Barass, L., Reid, S.M. & Brown, I.H. (2011).** Reassortant pandemic (H1N1) 2009 virus in pigs, United Kingdom. *Emerg Infect Dis* **17**, 1049-1052.
11. **Howden, K.H., Borckhoff, E.J., Vaya, F.D., McLeod, L.J., Lavole, M., Ding, J., Bystrom, J.M., Alexandersen, S., Pasick J.M. & other authors (2009).** An investigation into human pandemic influenza virus (H1N1) 2009 on an Alberta swine farm. *Can Vet J* **50**, 1153-1161.
12. **Karasin, A.I., Carman, S. & Olsen, C.W. (2006).** Identification of human H1N2 and human-swine reassortant H1N2 and H1N1 influenza A viruses among pigs in Ontario, Canada (2003 to 2005). *J Clin Microbiol* **44**, 1123-1126.
13. **Kim, S.H., Moon, O.K., Lee, K.K., Song, Y.K., Yeo, C.I., Bae, C.W., Yoon, H., Lee, O.S., Lee, J.H. & other authors (2011).** Outbreak of pandemic influenza (H1N1) 2009 in pigs in Korea. *Vet Rec* **169**, 155.
14. **Lam, T.T., Zhu, H., Wang, J., Smith, D.K., Holmes, E.C., Webster, R.G., Webby, R., Peiris, J.M. & Guan, Y. (2010).** Reassortment events among swine influenza A viruses in China: implications for the origin of the 2009 influenza pandemic. *J Virol* **85**, 10279-10285.
15. **Moreno, A., Chiapponi, C., Boniotti, M.B., Sozzi, E., Foni, E., Barbieri, I., Zanoni, G.M., Faccini, S., Lelli, D. & other authors (2012).** Genomic characterization of H1N2 swine influenza viruses in Italy. *Vet Microbiol* **156**, 265-276.
16. **Nelson, M.I., Lemey, P., Tan, Y., Vincent, A., Lam, T., Detmer, S., Viboud, C., Suchard, M.A., Rambaut, A., & other authors (2011).** Spatial dynamics of human-origin H1 influenza A virus in North American swine. *PLoS Pathog* **7**, e1002077.



17. **Njabo, K.Y., Fuller, T.L., Chasar, A., Pollinger, J.P., Cattoli, G., Terregino, C., Monne, I., Reynes, J.M., Njouom, R. & other authors (2011).** Pandemic A/H1N1/2009 influenza virus in swine, Cameroon, 2010. *Vet Microbiol* **156**, 189-192.
18. **Ottis, K., Sidoli, L., Bachmann, P.A., Webster, R.G. & Kaplan, M.M. (1982).** Human influenza A viruses in pigs: isolation of a H3N2 strain antigenically related to A/England/42/72 and evidence for continuous circulation of human viruses in the pig population. *Arch Virol* **73**, 103-108.
19. **Pereda, A., Cappuccio, J., Quiroga, M., Baumeister, E., Insarralde, L., Ibar, M., Sanguinetti, R., Cannilla, M.L., Franzese, D. & other authors (2010).** Pandemic (H1N1) 2009 outbreak on a pig farm, Argentina. *Emerg Infect Dis* **16**, 304-307.
20. **Pereda, A., Rimondi, A., Cappuccio, J., Sanguinetti, R., Angel, M., Ye, J., Sutton, T., Dibárbora, M., Olivera, V. & other authors (2011).** Evidence of reassortment of pandemic H1N1 influenza virus in swine in Argentina: are we facing the expansion of potential epicenters of influenza emergence? *Influenza Other Respi Viruses* **5**, 409-412.
21. **Rambaut, A. (2002).** Sequence alignment editor, version 2.0. Available: <http://tree.bio.ed.ac.uk/software/seal/>. Accessed 11 Dec 2011.
22. **Smith, G.J., Vijaykrishna, D., Bahl, J., Lycet, S.J., Worobey, M., Pybus, O.G., Ma, S.K., Cheung, C.L., Raghvani, J., Bhatt, S. & other authors (2009).** Origins and evolutionary genomics of the 2009 swine-origin H1N1 influenza A epidemic. *Nature* **459**, 1122-1125.
23. **Shu, B., Garten, R., Emery, S., Balish, A., Cooper, L., Sessions, W., Deyde, V., Smith, C., Berman, L., Klimov, A. & other authors (2012).** Genetic analysis and antigenic characterization of swine origin influenza viruses isolated from humans in the United States, 1990-2010. *Virology* **422**, 151-60.
24. **Sreta, D., Tantawet, S., Na Ayudhya S.N., Thontiravong, A., Wongphatcharachai, M., Lapkuntod, J., Bunpapong, N., Tuanudom, R., Suradhat, S. & other authors (2010).**

Pandemic (H1N1) 2009 virus on commercial swine farm, Thailand. *Emerg Infect Dis* **16**, 1587-1590.

25. **Swofford, D.L. (2003).** PAUP\*. Phylogenetic analysis using parsimony (\*and Other Methods). Version 4. Sinauer Associates, Sunderland, Massachusetts.
26. **Viboud, C., Miller, M., Olson, D., Osterholm, M. & Simonsen, L. (2010).** Preliminary estimates of mortality and years of life lost associated with the 2009 A/H1N1 pandemic in the US and comparison with past influenza seasons. *PLoS Curr* RRN1153.
27. **Vijaykrishna, D., Poon, L.L.M., Zhu, H.C., Ma, S.K., Li, O.T.W., Cheung, C.L., Smith, G.J.D., Peiris, J.S.M. & Guan, Y. (2010).** Reassortment of pandemic H1N1/2009 influenza A virus in swine. *Science* **328**,1529.
28. **Zhou, N.N., Senne, D.A., Landgraf, J.S., Swenson, S.L., Erickson, G., Rossow, K., Liu, L., Yoon, K., Krauss, S. & other authors (1999).** Genetic reassortment of avian, swine, and human influenza A viruses in American pigs. *J Virol* **73**, 8851-8856.

## FIGURE LEGENDS

**FIG 1.** Phylogenetic relationships of the six concatenated internal gene segment (PB2, PB1, PA, NP, M, NS) sequences from 100 H1N1pdm09 influenza virus isolates collected in swine globally during 2009-2011, and 1,008 H1N1pdm09 influenza viruses collected globally in humans during the same time period (total data set n = 1,108 sequences; color-coded black). Each swine H1N1pdm09 isolate is color-coded by the country of origin: United States (US) = brown, Canada (CAN) = light blue, Thailand (THD) = purple, South Korea (KOR) = dark green, China (CHN) = pink, Hong Kong (HKG) = red, Taiwan (TWN) = orange, Italy (ITL) = olive, Mexico (MEX) = light green, Singapore (SNG) = yellow, and Australia (AUS) = dark blue. The first introduction of H1N1pdm09 identified in swine, in May 2009 in Canada, is denoted with an asterisk. An identical phylogeny with all viral introductions labeled in detail is provided in Fig. S1.

**Table 1.** Country and state/province of viral collection, month/year of collection, number of swine isolates, and bootstrap support  $\geq 70$  for 49 introductions of H1N1pdm09 influenza virus from humans into swine during 2009-2011.

Introduction	Country of collection	State or province of collection (if available) <sup>#</sup>	Month, Year of collection	Number of isolates (swine)	Bootstrap support for viral introductions <sup>&amp;</sup>		
					<i>Internal gene phylogeny</i>	<i>HA phylogeny</i>	<i>NA phylogeny</i>
1	Canada	ALB	May 2009	2	85*	95	
2	Canada	ALB, MB	Aug 2009	2	94		
3	Australia	VIC	Aug 2009	6	95*	86	
4	Singapore		Aug 2009	1	72		
5	USA	MN	Aug-Sep 09	4	100		
6	Canada	MB	Sep-Oct 09	3		92	73
7	HK		Oct 2009	2	100		
8	Canada	QC	Nov 2009	1	85		
9	Canada	SK	Nov 2009	1	95		
10	Canada		Nov-Dec 09	3		84	72
11	Canada	QC	Nov-Dec 09	4		97	
12	Italy		Nov 2009	1	92		
13	S Korea		Nov-Dec 09	3	100*	98	70*
14	Canada	QC	Nov-Dec 09	5		90	84*
15	USA	IL	Nov 2009	1		71	
16	Taiwan		Nov 09 - Sep 10	4	98		82*
17	Thailand		Nov 09 - Jan 10	6	100		
18	China	Guangdong	Dec 2009	3	100	89	
19	China	Guangdong	Dec 2009	3	100	85	
20	HK		Dec 2009	2	100	94	
21	HK		Dec 2009	5	100		96
22	Italy		Dec 2009	2	100	98	
23	S Korea		Dec 2009	8	100	84	93
24	S Korea		Dec 09 - Jan 10	9	100	72	90
25	USA	IL, MN	Dec 09 - Feb 10	8		87	72*
26	USA	NC	Dec 09 - Mar 10	2		90	

27	USA	IL	Dec 09 - Jan 10	3		94*	76
28	China	Nanchang	Jan 2010	4	100	81	
29	China	Guangdong	Jan 2010	5	100		
30	Thailand		Feb - Jun 2010	4	97		
31	USA	IL, MO	Mar 2010	3		98	
32	China		May 2010	10	100		
33	USA	IL	May 2010	3		92*	97
34	Thailand		Sep 2010	2	100	80	
35	Costa Rica		Nov 2010	6		100	100
36	Cuba		Nov 2010	3		98*	100
37	Thailand		Nov 2010	2			99
38	USA	IA, IL	Nov - Dec 2010	3		98	97*
39	USA	IA	Nov 2010	2		100	100
40	USA	IL	Nov 10 - Mar 11	3			100
41	USA	MN, NE	Nov 10 - May 11	3		99	92
42	USA	NC, PA	Nov 10 - Feb 11	2		100	100
43	USA	IL	Nov 10 - Feb 11	3			94
44	USA	MN	Dec 10 - Jan 11	2			100
45	USA	IL	Dec 10 - Mar 11	2		100	
46	USA	MN	Dec 10 - Jan 11	2		100	98
47	USA	IL, KY, OH	Jan - Mar 2011	9		95	92
48	USA	MN	Feb - Apr 2011	2		100	99
49	USA	TX	Jul 2011	2	100	100	100

#For Canadian provinces, ALB = Alberta, MB = Manitoba, QC = Quebec, and SK = Saskatchewan; for Australia, VIC=Victoria; for the United States, IA = Iowa, IL = Illinois, KY = Kentucky, MN = Minnesota, MO = Missouri, OH = Ohio, NC = North Carolina, PA = Pennsylvania, and TX = Texas.

&Boxes that are empty indicate that no sequences were available for these genome segments or that bootstrap support was < 70.

\*Bootstrap support was  $\geq 70$  but only for a subset of the isolates in a given cluster.

**Table 2.** Country and year of collection, viral subtype, year of collection of the most phylogenetically related human influenza viruses, number of swine isolates, and bootstrap support  $\geq 70$  for 23 introductions of seasonal influenza viruses (H1, H3, N1, and N2 segments) from humans into swine during 1990-2011.

Introduction	Country & year of collection	Subtype	Related human viruses (year)	Number of isolates (swine)	Bootstrap support for viral introductions			
					H1	H3	N1	N2
1	Japan 1992	H1N1	1991	1			79	
2	Canada 1997	H3N2	1997	1		\$		98
3	Japan 1997	H3N2	1997	4		\$\$		84
4	USA 1998-1999 (triple reassortant I), S Korea 2005-2006, China 2007	H3N2	1995-96	10		100		97
5	HK 1999-2000, China 2003-2008	H3N2	1999	26		93		94*
6	France 1999	H3N2	1998	1		\$		99
7	HK 1999-2004, China 2004-2010	H1N2 <sup>##</sup>	1996	18				100
8	USA 1999, S Korea 2004	H3N2	1999	2		\$		95
9	Taiwan 2002, Indonesia 2004	H3N1 H3N2	1990	2		96		
10	USA 2003-2011, Canada 2005-2011 (triple reassortant II)	H3N2	1998	126 <sup>#</sup>		100		97,70 <sup>&amp;</sup>
11	Italy 2003-2010	H1N2 <sup>##</sup>	1997-98	20				100
12	Canada 2004	H1N2	2002-03	2				92
13	Thailand 2004-2009	H3N2	1995-96	8		100		100
14	China 2004-2006	H1N1	1997	2	93		100	
15	USA 2005-2011 ( $\delta$ -2)	H1N1 H1N2	2002-03	43 <sup>#</sup>	97		100*	97,70 <sup>&amp;</sup>
16	HK 2006	H1N1	2006-07	1	100		88	
17	S Korea 2007	H3N2	1995-96	3		100		97
18	USA 2007-2011 ( $\delta$ -1)	H1N2	2002-03	97	97			97,70 <sup>&amp;</sup>
19	China 2008	H1N1	2000-01	1	\$		80	
20	USA 2009	H1N1	2007	1	90			
21	Canada 2009	H1N1	2008-09	2	81		90	
22	China 2010, HK 2011	H3N2	2005	3		100		100

23	Vietnam 2010	H3N2	2005	6		100	100
a	HK 1999	H3N2	1999-00	1		-	-
b	HK 2000-2002	H3N2	2000-02	8		-	-
c	Canada 2003	H1N2	2002-03	1	-		-
d	USA 2003	H3N2	2001-02	1		-	-
e	Argentina 2008	H3N2	2001-02	1		-	-
f	Argentina 2009	H1N1	2005-07	1	-		-
g	Argentina 2010	H1N2	2002-03	1	-		-

\*Bootstrap support was  $\geq 70$  but only for a subset of the isolates in a given cluster.

#In cases of reassortment, the number of isolates on HA phylogeny is provided.

&In cases of reassortment, bootstrap values for both NA clades that are associated with an introduction on the HA tree are provided.

\$The HA associated with this introduction was identified to be of human seasonal influenza virus origin on the HA phylogeny (or the HA1 phylogeny<sup>\$\$</sup>), but was not associated with a significant bootstrap value.

##These H1N2 viruses are not related to the seasonal H1N2 viruses that circulated globally in humans 2001-2003 but rather are human-swine reassortants.

