

Adjustments of net photosynthesis in *Solanum tuberosum* in response to reciprocal changes in ambient and elevated growth CO₂ partial pressures

Richard C. Sicher and James A. Bunce

U.S.D.A., Agricultural Research Service, Climate Stress Laboratory, Bldg. 046-A, 10300 Baltimore Avenue, Beltsville Agricultural Research Center – West, Beltsville, MD 20705-2350, USA

*Corresponding author, e-mail: sicherr@ba.ars.usda.gov

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Single leaf photosynthetic rates and various leaf components of potato (*Solanum tuberosum* L.) were studied 1–3 days after reciprocally transferring plants between the ambient and elevated growth CO₂ treatments. Plants were raised from individual tuber sections in controlled environment chambers at either ambient (36 Pa) or elevated (72 Pa) CO₂. One half of the plants in each growth CO₂ treatment were transferred to the opposite CO₂ treatment 34 days after sowing (DAS). Net photosynthesis (P_n) rates and various leaf components were then measured 34, 35 and 37 DAS at both 36 and 72 Pa CO₂. Three-day means of single leaf P_n rates, leaf starch, glucose, initial and total Rubisco activity, Rubisco protein, chlorophyll (*a* + *b*), chlorophyll (*a*/*b*), α -amino N, and nitrate

levels differed significantly in the continuous ambient and elevated CO₂ treatments. Acclimation of single leaf P_n rates was partially to completely reversed 3 days after elevated CO₂-grown plants were shifted to ambient CO₂, whereas there was little evidence of photosynthetic acclimation 3 days after ambient CO₂-grown plants were shifted to elevated CO₂. In a four-way comparison of the 36, 72, 36 to 72 (shifted up) and 72 to 36 (shifted down) Pa CO₂ treatments 37 DAS, leaf starch, soluble carbohydrates, Rubisco protein and nitrate were the only photosynthetic factors that differed significantly. Simple and multiple regression analyses suggested that negative changes of P_n in response to growth CO₂ treatment were most closely correlated with increased leaf starch levels.

Introduction

Prior studies have established that growth in elevated atmospheric CO₂ usually resulted in increased rates of P_n and of biomass production (Bowes 1991, Stitt 1991). Diminished photosynthetic capacity that often occurred during long-term growth in elevated CO₂ was associated with modifications in both plant growth and in the chemical components of leaves and other plant parts. Among the many reported physiological and constitutive changes of plants in response to elevated growth CO₂ were a build-up of leaf starch (Poorter et al. 1997), decreased soluble proteins and total foliar N (Wong 1979, Nie et al. 1995), chlorosis and other forms of visible leaf damage (Tripp et al. 1991), premature senescence (Sicher and Bunce 1997, 1998) and decreased Rubisco activity and Rubisco protein levels (Wong 1979, Van Oosten and Besford 1995). There was also evidence that temporarily enhanced plant growth rates under elevated CO₂ could result in various nutrient limitations, particularly

N and P insufficiencies (Arp 1991). Nutrient deficiencies would adversely affect rates of protein synthesis and thereby inhibit P_n indirectly (Stitt and Krapp 1998).

Large numbers of studies have investigated acclimation of P_n in response to CO₂ enrichment (e.g. Stitt and Krapp 1998). Photosynthetic acclimation has been defined as differing rates of P_n for ambient and elevated CO₂-grown plants measured at the same CO₂ partial pressure (Long 1991). Direct comparisons of plants grown continuously in ambient and elevated CO₂ have generated diverse and often conflicting observations. This inherent complexity has made it difficult to identify mechanisms responsible for photosynthetic acclimation. An alternative and less frequent approach to studying growth CO₂ effects on P_n was to examine the induction and/or the reversibility of photosynthetic acclimation after transferring plants between atmospheres having either enriched or ambient CO₂. Prior studies

Abbreviations – Chl (*a* + *b*), chlorophyll *a* plus *b*; Chl (*a*/*b*), chlorophyll *a* to *b* ratio.; PAR, photosynthetically active radiation; p_i(36) and p_i(72), estimated internal leaf CO₂ partial pressures measured at 36 and 72 Pa external CO₂, respectively; P_n(36) and P_n(72), net photosynthesis rates measured at 36 and 72 Pa external CO₂, respectively; Rubisco, ribulose 1,5-bisphosphate carboxylase/oxygenase.

using this approach have been performed with cotton (Sasek et al. 1985), rice (Gesch et al. 1998), clover (Morin et al. 1992), tobacco (Sicher et al. 1994) and soybean (Sicher et al. 1995). Most of these earlier investigations showed that photosynthetic acclimation was almost completely reversed within 1–3 days after plants were transferred from elevated to ambient CO₂. Leaf starch levels (Sasek et al. 1985, Sicher et al. 1995), Rubisco activity, and Rubisco protein and soluble carbohydrate concentrations (Gesch et al. 1998) also were altered within days after plants were transferred from elevated to ambient CO₂.

Most previous CO₂ enrichment studies have attributed observed decreases of photosynthetic capacity to changes of Rubisco activity or Rubisco protein concentration (Bowes 1991, Stitt 1991). Our earlier field CO₂ enrichment experiment with potato was consistent with this generalization (Sicher and Bunce 1999). However, it has not been established that reversible changes of P_n in response to transitions between elevated and ambient CO₂ growth conditions were the result of altered Rubisco activity. The extent to which leaf constituents and other factors associated with photosynthetic acclimation are reversed upon changing growth CO₂ treatments is also uncertain. Our objective in the current study was to use quantitative statistical approaches to elucidate the role of Rubisco activity and possibly other leaf factors in altering P_n in response to transitions between ambient and elevated growth CO₂.

The hypothesis in the present study, was that the reversible changes of P_n in *Solanum tuberosum* L. within 1–3 days after changing the ambient and elevated growth CO₂ conditions would be attributable to altered Rubisco activity. We further assumed that leaf constituents affecting the inhibition of P_n during growth in elevated CO₂ would be reversible concomitant with changes of P_n.

Materials and methods

Plant material

Potato plants (*Solanum tuberosum* L. cv. Atlantic) were grown from tuber sections planted in 3-l pots filled with vermiculite. Plants were raised in controlled environment chambers (model M-3, EGC Corporation, Chagrin Falls, OH, USA) with a 14-h/10-h light/dark photoperiod and at a 22°C/16°C day/night temperature. Relative humidity was never less than 50% and irradiance was 1.05 mmol m⁻² s⁻¹ PAR (Sicher and Bunce 1997). Between 10 and 20 plants were grown continuously at either 36 (ambient) or 72 (elevated) Pa CO₂. One-half of the ambient CO₂-grown plants were transferred to 72 Pa CO₂ (shifted up) beginning 34 days after sowing (DAS), and simultaneously, one-half of the elevated CO₂-grown plants were transferred to 36 Pa CO₂ (shifted down). Results are representative of 3 experiments using a total of 4 growth chambers. Gas-exchange and Rubisco activity data were combined from the results of two experiments. Data for all other measurements were from one experiment (n = 4–5) to reduce variability.

Gas-exchange measurements

Net photosynthesis (P_n) rates were measured 3-h after the start of the light period with a portable infrared gas-analysis system (CIRAS-1, PP Systems, Haverhill, MA, USA) using the terminal leaflet of the uppermost fully expanded leaf (Bunce 1998). Gas-exchange measurements were performed 34, 35 and 37 DAS at both 36 and 72 Pa external CO₂ using plants in all 4 CO₂ treatments. These are referred to as the P_n(36) and the P_n(72) measurements, respectively. Other than CO₂, all measurement conditions were essentially as for plant growth. Measurements were repeated until consecutive values varied by less than 5%. Internal CO₂ partial pressures at both measurement CO₂ levels [p_i(36) and p_i(72)] and P_n rates were obtained following von Caemmerer and Farquhar (1981) and were expressed as means ± SE (n = 6).

Leaf components

Immediately after performing the gas-exchange measurements, 3 pairs of leaf discs (3.6 cm²) were removed from the penultimate leaflets of the same leaf used for gas-exchange analysis. Foliar samples were used to measure pigments, carbohydrates, protein levels, Rubisco activity and other leaf constituents. One-third of the samples were extracted with methanol:chloroform:water (5:3:1, v/v) and chlorophyll (Chl) *a* and *b* were determined in 80% acetone after partitioning the solvent extracts (Lichtenthaler 1987). Leaf starch was obtained from the pellet fraction and was quantified enzymically according to Hendrix (1993). The aqueous fraction was concentrated to a minimum volume and was diluted to 1.0 ml with deionized H₂O. Sucrose, glucose and fructose were measured in coupled enzyme assays according to Bergmeyer et al. (1974). The aqueous concentrates were diluted 1:4 and nitrate was measured by isocratic high-pressure liquid chromatography (HPLC) using an anion exchange column (Whatman Partisil-10 SAX, Clifton, NJ, USA) according to Thayer and Huffaker (1980). The column was eluted with 50 mM phosphate buffer pH 3.0, at 1.0 ml min⁻¹ and nitrate was detected at 210 nm using a multi-wavelength detector (Waters 490E, Milford, MA, USA). Recovery of standard nitrate was greater than 90%. α-Amino N also was measured in the aqueous concentrates by a ninhydrin procedure using glycine as a standard (Sicher and Bunce 1998). Initial and total Rubisco activity was measured using a separate pair of leaf discs. Assays were performed before and after activating the enzyme with CO₂ and Mg²⁺ using a radiometric procedure (Sicher and Bunce 1999). Rubisco protein was quantified by a dye-binding method after separation by denaturing gel electrophoresis (Sicher et al. 1994).

Statistical treatments

Significant differences for mean values (n = 13–18) of photosynthetic factors measured 34, 35 and 37 DAS were compared using a repeated measures analysis of variance (ANOVA) procedure (Statview 5.0, SAS Inst., Raleigh, NC, USA) with date and treatment as conditionals. Mean differences were tested at *P* ≤ 0.05 (*) or *P* ≤ 0.001 (**). Factors

that differed significantly were tested for reversibility in reciprocal transfer experiments based on measurements performed 34 and 37 DAS ($n = 4-9$). An overall CO₂ treatment effect was identified using a one-way ANOVA procedure and individual treatment means that differed significantly were assigned probabilities using Fisher's protected LSD test. In addition, correlation coefficients and probability values for photosynthetic responses measured at 36 and 72 Pa CO₂ and various leaf constituents measured 34, 35 and 37 DAS were calculated by linear and multiple regression analyses using the same computer program.

Results

Physiological responses to CO₂ enrichment

Prior to examining the reversibility of P_n upon switching CO₂ treatments, we first sought to establish which photosynthetic parameters in *S. tuberosum* responded to continuous CO₂ enrichment. Three-day means measured 34, 35 and 37 DAS and corresponding significance levels of the various factors measured in this study are shown in Table 1. Mean single leaf P_n rates determined at both 36 and 72 Pa CO₂ were 37 and 25% lower for plants grown in the continuous elevated compared with the continuous ambient CO₂ treatments, respectively. Differences between means for both P_n(36) and P_n(72) were highly significant. Similarly, leaflet starch levels were almost threefold greater in elevated compared with ambient CO₂-grown plants. The only other factor that was highly significantly different between the continuous ambient and continuous elevated CO₂ treatments was total Rubisco activity, which was about 21% lower in elevated compared with ambient CO₂-grown plants. In contrast to the above, p_i(36), p_i(72), sucrose, fructose, soluble protein and percent Rubisco activation did not differ between the ambient and elevated CO₂ treatments. Rubisco activation also did not differ between CO₂ treatments after the data were transformed with a trigonometric function

prior to statistical analysis. Leaflet sucrose levels were unaffected but glucose was decreased by about 32% in response to CO₂ enrichment. Leaflet Chl ($a + b$) levels, the Chl (a/b) ratio, Rubisco protein, α -amino N, nitrate, and initial Rubisco activity were all lower in elevated compared with ambient CO₂-grown plants ($P \leq 0.05$). Averaged over CO₂ treatments, the Chl (a/b) ratio increased and α -amino N levels decreased significantly with leaf age. Soluble protein differed by date because leaf concentrations were slightly greater 35 DAS compared with 34 and 37 DAS (data not shown). A CO₂ by date interaction was detected for P_n(36) and p_i(72). However, these were minor and did not affect the overall differences in photosynthetic rates because of the growth CO₂ treatments.

Reversibility of photosynthetic gas-exchange rates

The responses of net leaflet P_n rates of ambient and elevated CO₂-grown potato plants to reciprocal transfers in CO₂ treatment were dependent upon the direction of transfer and on the measurement CO₂ partial pressure (Fig. 1A,B). Leaflet P_n rates of elevated CO₂-grown plants, determined 72 h after the plants were shifted down (72 to 36 Pa), were either partially or completely de-acclimated under the P_n(36) and P_n(72) measurement conditions, respectively. Comparable leaflet P_n(72) rates measured 37 DAS of plants that were shifted up (36 to 72 Pa) were unchanged in comparison with the ambient controls.

However, plants that were shifted up (36 to 72 Pa) in the P_n(36) measurement were partially acclimated to elevated CO₂ after 72 h of treatment. A CO₂ treatment effect on p_i(36) was detected ($P \leq 0.001$) in the shifted experiments 37 DAS (Fig. 1C). However, the changes in p_i(36) were too small to fundamentally alter the response of P_n(36) to the switched treatments discussed above. No significant CO₂ treatment effects were detected for p_i(72) when measured 37 DAS.

Table 1. Three-day means and significance levels of gas-exchange parameters and leaf components of ambient (36 Pa) and elevated (72 Pa) CO₂-grown *Solanum tuberosum*. Values are means for samples measured 34, 35 and 37 DAS. Probabilities are for $P \leq 0.05$ (*), $P \leq 0.001$ (**), and $P \geq 0.05$ (NS) based on a repeated measures analysis of variance procedure.

Measurement	Growth CO ₂		Probability		
	36 Pa	72 Pa	CO ₂	Date	CO ₂ × date
P _n (36), $\mu\text{mol m}^{-2} \text{s}^{-1}$	27.2	17.5	**	NS	*
P _n (72), $\mu\text{mol m}^{-2} \text{s}^{-1}$	50.2	37.6	**	NS	NS
p _i (36), Pa	23.8	23.2	NS	NS	NS
p _i (72), Pa	47.1	45.2	NS	NS	*
Starch, g m ⁻²	70.1	208.1	**	NS	NS
Sucrose, g m ⁻²	6.3	8.3	NS	NS	NS
Glucose, g m ⁻²	0.44	0.24	*	NS	NS
Fructose, g m ⁻²	0.52	0.34	NS	NS	NS
Chl ($a + b$), g m ⁻²	0.44	0.41	*	NS	NS
Chl (a/b), ratio	4.8	4.6	*	**	NS
Soluble protein, g m ⁻²	11.3	11.2	NS	*	NS
Rubisco protein, g m ⁻²	3.5	3.1	*	NS	NS
α -amino N, mmol m ⁻²	7.2	6.3	*	*	NS
Nitrate, g m ⁻²	0.69	0.46	*	NS	NS
Initial Rubisco activity, $\mu\text{mol m}^{-2} \text{s}^{-1}$	53.2	43.1	*	NS	NS
Total Rubisco activity, $\mu\text{mol m}^{-2} \text{s}^{-1}$	73.6	58.2	**	NS	NS
Percent activation	74	73	NS	NS	NS

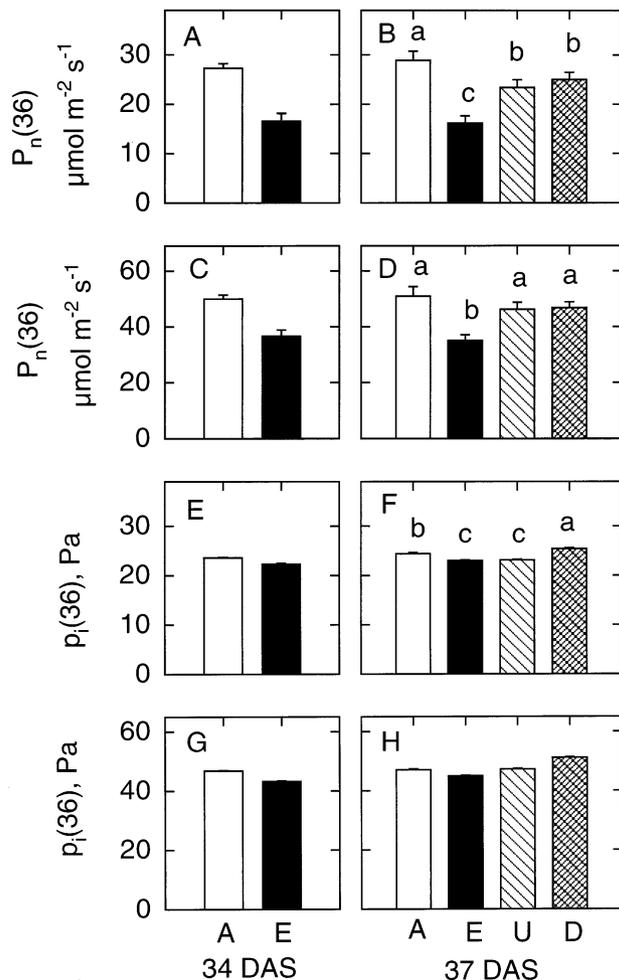


Fig. 1. Effects of a reciprocal transfer in growth CO_2 treatments on gas-exchange rates of *Solanum tuberosum* (L.). (A–D) Net rates of photosynthesis (P_n) and (E–H) internal leaf CO_2 partial pressures (p_i) were measured at 36 Pa (A,B,E,F) and 72 Pa (C,D,G,H) CO_2 . Values are means \pm SE for $n=4$ for measurements performed 34 and 37 DAS. Separate letters designate significant differences at $P \leq 0.05$. Specific CO_2 treatments are the following: A, continuous ambient (open bars); E, continuous elevated (filled bars); U, shifted up 36 to > 72 Pa CO_2 (single hatch); and D, shifted down 72 to > 36 Pa CO_2 34 (cross hatch).

Reversibility of leaf carbohydrate levels

Starch levels in the shifted up and shifted down CO_2 treatments measured 37 DAS were not significantly different from the respective elevated and ambient CO_2 controls (Fig. 2A). Therefore, leaflet starch concentrations were fully acclimated and fully de-acclimated 72-h after a reciprocal shift in the ambient and elevated CO_2 treatments. The response of soluble carbohydrate concentrations to reciprocal shifts in the CO_2 treatment differed from the observations for starch (Fig. 2B–D). Sucrose and fructose, but not glucose, exhibited increased leaflet concentrations 72 h after ambient CO_2 -grown plants were shifted up. Increased fructose and sucrose levels in the shifted up treatment measured 37 DAS were probably transitory. None of the three soluble carbohydrates measured in potato leaflets responded significantly to the shifted down treatment when determined 37 DAS.

Reversibility of leaflet N-constituents

There were no changes of leaflet Rubisco protein levels measured 37 DAS when ambient CO_2 -grown plants were shifted up (Fig. 3A). However, Rubisco protein increased 11.5% in the shifted down treatment when compared with elevated CO_2 -grown plants 37 DAS. Rubisco protein levels were fully de-acclimated in potato leaflets 3-days after plants were transferred from elevated to ambient CO_2 . A CO_2 treatment effect on leaflet nitrate concentrations also was observed (Fig. 3B). Values for nitrate in the shifted up plants were intermediate between the ambient and elevated CO_2 treatments. However, no significant differences were detected in leaflet nitrate levels in the elevated and the shifted down plants 37 DAS. Changes of Chl ($a+b$), Chl (a/b) ratio and α -amino N were all non-significant with respect to all 4 of the CO_2 treatments 37 DAS (data not shown). The shifted up and shifted down CO_2 treatments also had no effect on initial or total Rubisco activity or on percent Rubisco activation (data not shown).

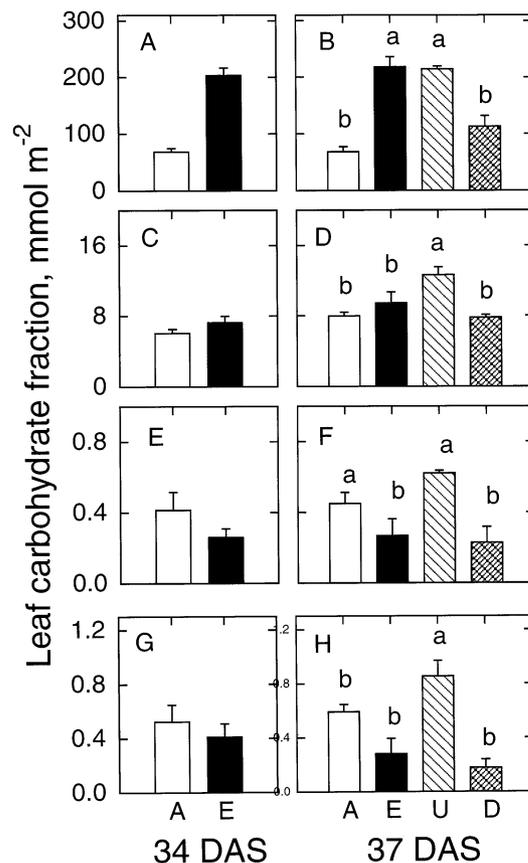


Fig. 2. Effects of a reciprocal transfer in growth CO_2 treatments on leaflet non-structural carbohydrate levels of *Solanum tuberosum* (L.). (A,B) Leaf starch, (C,D) sucrose, (E,F) glucose, and (G,H) fructose concentrations in leaflets of potato plants were measured 34 and 37 DAS. CO_2 treatments and other conditions were as in Fig. 1.

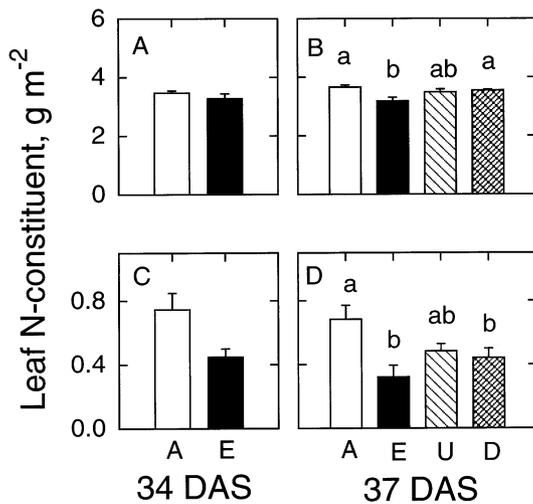


Fig. 3. Effects of a reciprocal transfer in growth CO₂ treatments on nitrate and Rubisco protein levels in leaflets of *Solanum tuberosum* (L.). (A,B) Rubisco protein and (C,D) nitrate concentrations in leaflets of potato plants were measured 34 and 37 DAS. CO₂ treatments and other conditions were as in Fig. 1.

Regression analyses of CO₂-dependent acclimation responses

Relationships between P_n measured 34, 35 and 37 DAS versus various potato leaf components in response to reciprocal transfers between ambient and elevated CO₂ growth conditions were analyzed by linear regression (Fig. 4a–c). Three leaf factors, leaf starch, initial Rubisco activity and nitrate concentration, generated statistically significant ($P < 0.05$) regressions versus both P_n(36) and P_n(72). Leaf starch was negatively correlated with P_n(36), whereas both initial Rubisco activity and leaf nitrate levels were positively correlated with P_n(36). There was a better correlation between P_n(36) and leaf starch [$y = -0.059(x) + 31.84$ ($R^2 = 0.701$, $P \leq 0.003$)] than there was for initial Rubisco activity [$y = 0.789(x) + 29.01$ ($R^2 = 0.517$, $P \leq 0.019$)] or for leaf nitrate concentration [$y = 0.248(x) + 9.25$ ($R^2 = 0.562$, $P \leq 0.013$)]. Multiple regression analysis for P_n(36) versus leaf starch and initial Rubisco activity also was significant ($R^2 = 0.756$, $P \leq 0.006$), and indicated a significant P -value for starch ($P = 0.048$) but not for initial Rubisco activity ($P = 0.381$). A comparable analysis for P_n(36) versus leaf starch and nitrate also was statistically significant ($R^2 = 0.765$, $P \leq 0.006$). The P -value for starch was significant ($P = 0.044$), whereas that for nitrate was not ($P = 0.210$).

Discussion

The physiological responses of *S. tuberosum* to CO₂ enrichment were consistent with prior reports for a number of other C₃ crop species, including tomato (Van Oosten and Besford 1995), wheat (Nie et al. 1995, Sicher and Bunce 1997, 1998), rice (Gesch et al. 1998), tobacco (Sicher et al. 1994) and cotton (Sasek et al. 1985). Single leaf rates of P_n in potato were inhibited 25–37% on average when compared at the same measurement CO₂ partial pressure. In spite of this observed photosynthetic acclimation, rates of P_n

measured at the same growth CO₂ concentration were always greater under CO₂ enrichment. Leaf starch was almost threefold greater in leaflets of elevated compared with ambient CO₂, whereas total and initial Rubisco activity, Rubisco protein, Chl ($a + b$), the Chl (a/b) ratio and nitrate levels were all decreased in leaflets of potato plants grown at

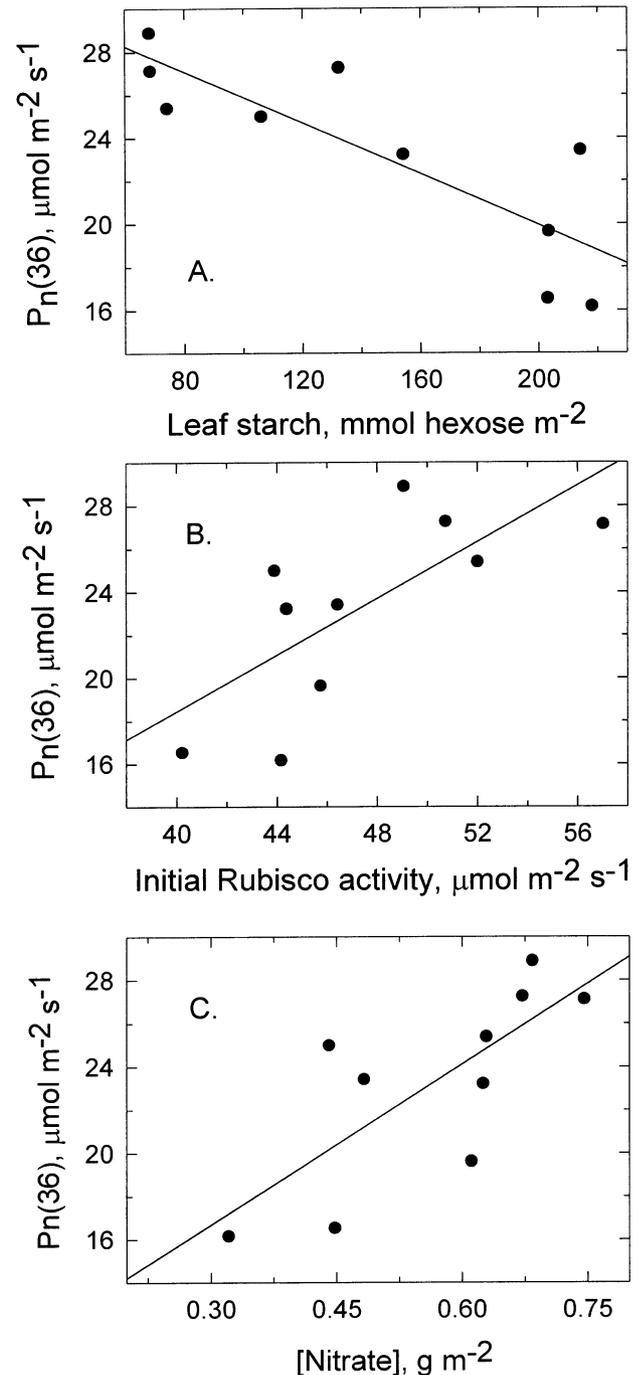


Fig. 4. Simple linear regression analyses of potato leaflet biochemical factors versus P_n(36) in response to reciprocal transfers between ambient and elevated growth CO₂ treatments. Regressions were for (A) leaf starch, (B) initial Rubisco activity and (C) leaflet nitrate concentration. Individual values were means ($n = 4-6$) measured 34, 35 and 37 DAS.

elevated compared with ambient CO₂. Current results showing that total Rubisco activity was reduced by CO₂ enrichment also were in agreement with a prior field study using potato grown in open-topped field chambers (Sicher and Bunce 1999). Although some of the physiological responses of potato to elevated CO₂ were not as great as reports for other species, we concluded that photosynthetic acclimation in potato involved conventional changes of leaf components and of related photosynthetic factors.

Reciprocal changes of growth CO₂ concentrations demonstrated that the inhibition of P_n in *S. tuberosum* under CO₂ enrichment was partially to completely reversed 3 days after plants were shifted down from elevated to ambient CO₂. This was consistent with prior reports for soybean (Sicher et al. 1995), cotton (Sasek et al. 1985) and mature rice leaves (Gesch et al. 1998). Conversely, little or no acclimation of P_n was observed 3 days after potato plants were shifted up from ambient to elevated CO₂. Sicher et al. (1994) reported that P_n rates of mature tobacco leaves required a week or more to acclimate to elevated CO₂ after being shifted up from ambient CO₂. In comparison, Morin et al. (1992) observed that P_n rates of clover acclimated to elevated CO₂ within 24 h, although plants in this experiment were transferred to an atmosphere having a twelvefold increase in ambient CO₂. Gesch et al. (1998) observed an apparent 15% inhibition of P_n (i.e. maximum acclimation) in mature rice leaves 3 days after plants were shifted up from ambient to elevated CO₂. Measurements of p_i(36) and p_i(72) in the current study suggested that changes of P_n in response to shifting the ambient and elevated CO₂ treatments were the result of non-stomatal factors (cf. Sasek et al. 1985). At least for the species examined thus far, transfer studies using 2–3 times ambient CO₂ demonstrated that acclimation and de-acclimation of P_n can occur at different rates. This finding may have potential implications for understanding the biochemical basis for photosynthetic acclimation to elevated CO₂.

The above findings demonstrated that P_n rates and various potato leaf constituents responded to reciprocal transfers between the ambient and elevated growth CO₂ treatments. Regression analyses were used to quantitatively assess which leaf constituents responded to CO₂-dependent changes of P_n. Leaf starch levels, initial Rubisco activity and leaf nitrate concentrations were most closely correlated with responses of P_n to changes in growth CO₂. A build-up of leaf starch and soluble carbohydrates has been frequently associated with feedback-inhibited photosynthesis (Azcón-Bieto 1983). Recent efforts have identified an end product synthesis limitation (Ludewig et al. 1998), sucrose cycling (Moore et al. 1998) and carbohydrate regulated gene expression (Jang and Sheen 1997) as mechanisms that carbohydrates modify the photosynthetic capacity of leaves. Sasek et al. (1985) observed that excess leaf starch was completely mobilized 3 days after elevated CO₂-grown cotton was transferred to ambient CO₂. This closely coincided with the de-acclimation of P_n in cotton and suggested to these authors that a CO₂-dependent build-up of starch was responsible for the inhibition of P_n. Rey and Jarvis (1998) similarly concluded that decreases of photosynthetic capacity in response to elevated growth CO₂ were associated with in-

creased leaf starch levels in *Betula pendula*. Leaflet starch levels were threefold greater in elevated than in ambient CO₂-grown potato plants. Moreover, starch levels were completely acclimated and de-acclimated within 3 days after potato plants were shifted up or down. Not surprisingly, there was a close correlation between leaf starch levels and the acclimation of P_n in *S. tuberosum* in the present study (R² = 0.701). The mechanism by which elevated leaf starch resulted in a decrease of P_n in response to CO₂ enrichment is unknown. In contrast to starch, there was no direct correlation between changes on total foliar soluble carbohydrate pools and photosynthetic acclimation to elevated CO₂ in potato.

Decreased Rubisco activity and Rubisco protein levels frequently have been observed in association with photosynthetic acclimation to elevated CO₂ (Bowes 1991, Stitt 1991). Results with *S. tuberosum* grown continuously with ambient and elevated CO₂ both here and in open-topped field chambers (Sicher and Bunce 1999) were in general agreement with this conclusion. Initial and total Rubisco activities remained unchanged 3 days after potato plants were reciprocally transferred between the ambient and elevated CO₂ treatments. However, a 12% increase of Rubisco protein was observed 3 days after plants were shifted from elevated to ambient CO₂. As the changes of Rubisco protein and of Rubisco activity in the current study were small, it was not obvious that Rubisco was responsible for the 25–37% inhibition of photosynthesis that occurred in this species as a result of CO₂ enrichment.

Growth in elevated CO₂ can create an imbalance between nutrient supply and utilization in roots and leaves (Stitt and Krapp 1998, Geiger et al. 1999). In agreement with this observation, foliar nitrate levels were significantly lower in elevated than in ambient CO₂-grown potato plants both here and previously (Ludewig et al. 1998). This occurred in spite of the fact that the nutrient solution in this study was 12 mM in nitrate and 3.5 mM in ammonium. However, leaflet nitrate levels were unchanged 3 days after elevated CO₂-grown plants were shifted down to ambient CO₂. Nitrate concentrations in these plants were 10–25 times greater than those observed in potato raised in open-topped field chambers (Sicher and Bunce 1999). Consequently, it cannot be concluded that reduced leaflet nitrate levels during growth in elevated CO₂ created an N-limited condition. It was not obvious why nitrate levels were decreased by growth in elevated CO₂, although lower evapotranspiration rates and a greater dependence on reduced N sources could be factors.

In summary, reciprocal transfer experiments with *S. tuberosum* showed that the inhibition of P_n during growth in elevated CO₂ was partially to completely reversed 3 days after plants were transferred to ambient CO₂. However, little or no evidence of photosynthetic acclimation was observed 3 days after plants were shifted up from ambient to elevated growth CO₂. Leaf starch, soluble carbohydrates, Rubisco activity, Rubisco protein, Chl (*a + b*) and nitrate were all affected by CO₂ enrichment in this study. Simple and multiple regression analyses indicated that growth CO₂-dependent changes of P_n in potato were most closely correlated with leaf starch levels. The mechanism whereby leaf starch affected P_n has not been identified.

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References

- Arp WJ (1991) Effects of source-sink relations on photosynthetic acclimation to elevated CO₂. *Plant Cell Environ* 14: 869–875
- Azcón-Bieto J (1983) Inhibition of photosynthesis by carbohydrates in wheat leaves. *Plant Physiol* 73: 681–686
- Bergmeyer HU, Brent E, Schmidt F, Stock H (1974) D-Glucose. Determination with hexokinase and glucose 6-phosphate dehydrogenase. In: Bergmeyer HU (ed) *Methods of Enzymatic Analysis*, Vol 3, 2nd Edn. Academic Press, New York, NY, pp 1196–1198
- Bowes G (1991) Growth at elevated CO₂: Photosynthetic responses mediated through Rubisco. *Plant Cell Environ* 14: 795–806
- Bunce JA (1998) Effects of humidity on short-term response of stomatal conductance to an increase in carbon dioxide concentration. *Plant Cell Environ* 21: 115–120
- Geiger M, Haake V, Ludewig F, Sonnewald U, Stitt M (1999) The nitrate and ammonium nitrate supply have a major influence on the response of photosynthesis, carbon metabolism, nitrogen metabolism and growth to elevated carbon dioxide in tobacco. *Plant Cell Environ* 22: 1177–1199
- Gesch RW, Boote KJ, Vu JCV, Allen LH Jr, Bowes G (1998) Changes in growth CO₂ result in rapid adjustments of ribulose 1,5-bisphosphate carboxylase/oxygenase small subunit gene expression in expanding and mature leaves of rice. *Plant Physiol* 118: 521–529
- Hendrix DL (1993) Rapid extraction and analysis of nonstructural carbohydrates in plant tissues. *Crop Sci* 33: 1306–1311
- Jang J-Y, Sheen J (1997) Sugar sensing in higher plants. *Trends Plant Sci* 2: 208–214
- Lichtenthaler HK (1987) Chlorophylls and carotenoids: Pigments of photosynthetic biomembranes. *Methods Enzymol* 148: 350–382
- Long SP (1991) Modification of the response of photosynthetic productivity to rising temperature by atmospheric CO₂ concentrations. Has its importance been underestimated? *Plant Cell Environ* 14: 729–739
- Ludewig F, Sonnewald U, Kauder F, Heineke D, Geiger M, Stitt M, Müller-Robert BT, Gillisen B, Kühn C, Frommer WB (1998) Role of transient starch in acclimation to elevated CO₂. *FEBS Lett* 429: 147–151
- Moore B, Cheng S-H, Seemann JR (1998) Sucrose cycling, Rubisco expression, and prediction of photosynthetic acclimation to elevated atmospheric CO₂. *Plant Cell Environ* 21: 905–916
- Morin F, André M, Betsche T (1992) Growth kinetics, carbohydrate, and leaf phosphate content of clover (*Trifolium subterraneum* L.) after transfer to a high CO₂ atmosphere or to high light and ambient air. *Plant Physiol* 99: 89–95
- Nie GY, Long SP, Garcia RL, Kimball BA, La Morte RL, Pinter PJ, Wall GW, Webber A (1995) Effects of free-air CO₂ enrichment on the development of the photosynthetic apparatus in wheat, as indicated by changes in leaf proteins. *Plant Cell Environ* 18: 855–864
- Poorter H, van Berkel Y, Baxter R, den Hertog J, Dijkstra P, Gifford RM, Griffin KL, Roumet C, Roy J, Wong SC (1997) The effect of elevated CO₂ on the chemical composition and construction costs of leaves of 27 C₃ species. *Plant Cell Environ* 20: 472–482
- Rey A, Jarvis PG (1998) Long-term photosynthetic acclimation to increased atmospheric CO₂ concentration in young birch (*Betula pendula*) trees. *Tree Physiol* 18: 441–450
- Sasek TW, De Lucia EH, Strain BR (1985) Reversibility of photosynthetic inhibition in cotton after long-term exposure to elevated CO₂ concentrations. *Plant Physiol* 78: 619–622
- Sicher RC, Bunce JA (1997) Relationship of photosynthetic acclimation to changes of Rubisco activity in field-grown winter wheat and barley during growth in elevated carbon dioxide. *Photosynth Res* 52: 27–38
- Sicher RC, Bunce JA (1998) Evidence that premature senescence affects photosynthetic decline of wheat flag leaves during growth in elevated carbon dioxide. *Int J Plant Sci* 159: 798–804
- Sicher RC, Bunce JA (1999) Photosynthetic enhancement and conductance to water vapor of field-grown *Solanum tuberosum* (L.) in response to CO₂ enrichment. *Photosynth Res* 62: 155–163
- Sicher RC, Kremer DF, Rodermeil SR (1994) Photosynthetic acclimation to elevated CO₂ occurs in transformed tobacco with decreased ribulose-1,5-bisphosphate carboxylase/oxygenase content. *Plant Physiol* 104: 409–415
- Sicher RC, Kremer DF, Bunce JA (1995) Photosynthetic acclimation and photosynthate partitioning in soybean leaves in response to carbon dioxide enrichment. *Photosynth Res* 46: 409–417
- Stitt M (1991) Rising CO₂ levels and their potential significance for carbon flow in photosynthetic cells. *Plant Cell Environ* 14: 741–762
- Stitt M, Krapp A (1998) The interaction between elevated carbon dioxide and nitrogen nutrition: The physiological and molecular background. *Plant Cell Environ* 22: 583–621
- Thayer JR, Huffaker RC (1980) Determination of nitrate and nitrite by high-pressure liquid chromatography: Comparison with other methods for nitrate determination. *Anal Biochem* 102: 110–119
- Tripp KE, Peet MM, Pharr DM, Willits DH, Nelson PV (1991) CO₂-enhanced yield and foliar deformation among tomato genotypes in elevated CO₂ environments. *Plant Physiol* 96: 713–719
- Van Oosten JJ, Besford RT (1995) Some relationships between the gas exchange, biochemistry and molecular biology of photosynthesis during leaf development of tomato plants after transfer to different carbon dioxide concentrations. *Plant Cell Environ* 18: 1253–1266
- von Caemmerer S, Farquhar GD (1981) Some relationships between biochemistry of photosynthesis and gas exchange of leaves. *Planta* 153: 376–387
- Wong SC (1979) Elevated atmospheric partial pressure of CO₂ and plant growth. I. Interactions of nitrogen nutrition and photosynthetic capacity in C₃ and C₄ plants. *Oecologia* 44: 68–74

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