Response of Four Apple Cultivars to 1-Methylcyclopropene Treatment and Controlled Atmosphere Storage

Jinhe Bai
Oregon State University, Mid-Columbia Agricultural Research and Extension Center, Hood River, OR 97031

Elizabeth A. Baldwin1 and Kevin L. Goodner
U.S. Department of Agriculture, Agricultural Research Service, Citrus and Subtropical Products Laboratory, Winter Haven, FL 33881

James P. Mattheis
U.S. Department of Agriculture, Agricultural Research Service, Tree Fruit Research Laboratory, Wenatchee, WA 98801

Jeffrey K. Brecht
Horticultural Sciences Department, University of Florida, Gainesville, FL 32611

Additional index words. Malus sylvestris var. domestica, firmness, titratable acidity, volatile abundance, 1-MCP, CA

Abstract. Apples [Malus sylvestris (L.) Mill var. domestica (Borkh.) Mansf. (‘Gala’, ‘Delicious’, ‘Granny Smith’ and ‘Fuji’)], pretreated or nontreated with 1-methylcyclopropene (1-MCP, 0.6 to 1.0 µL·L–1 for 18 hours at 20 °C), were stored in controlled atmosphere (CA, 1 to 1.5 kPa O2; 1 to 2 kPa CO2) or in regular atmosphere (RA) for up to 8 months at 1 °C. Firmness, titratable acidity (TA), soluble solids content (SSC), and volatile abundance were analyzed every month directly or after transfer to air at 20 °C for 1 week to determine effect of 1-MCP, storage atmosphere and storage time on apple quality immediately after cold storage and after simulated marketing conditions at 20 °C. The 1-MCP + CA treatments delayed ripening and prolonged storage life as indicated by delayed loss of firmness and TA in all four cultivars during storage. The 1-MCP + CA also slightly delayed loss of SSC for ‘Gala’ but had no effect on SSC levels for the other cultivars. There were differences among treatments for firmness and TA content [(1-MCP + RA) > CA] for ‘Gala’, ‘Delicious’, and ‘Granny Smith’ apples, but not for ‘Fuji’. These differences were generally exacerbated after transfer of fruit to 20 °C for 1 week. A combination of 1-MCP + CA was generally best [(1-MCP + CA) > (1-MCP + RA) or CA] for maintaining ‘Delicious’ firmness and TA. However, the treatments that were most effective at retaining TA and firmness also retained the least volatiles. The results indicate that the efficacy of 1-MCP and CA in maintaining apple quality factors is cultivar dependent and that 1-MCP + RA may be a viable alternative to optimal eating quality for some cultivars.

The compound 1-methylcyclopropene (1-MCP) has been discovered to be a very effective inhibitor of ethylene action due to competitive binding on the ethylene receptor in plant tissue (Sisler et al., 1996; Sisler and Serek, 1999; Sisler and Wood, 1988). 1-MCP has been recently approved by the U.S. Environmental Protection Agency (2002) as a reduced-risk product for use on produce. Much research has been conducted in the last few years, which showed that 1-MCP decreased ethylene production; reduced respiration, softening, loss of titratable acidity, color change, and some physiological disorders; and increased storage life of apples (Blankenship and Dole, 2003; Watkins and Miller, 2004). 1-MCP has been considered a major breakthrough in apple storage technology.

The efficacy of 1-MCP depends on cultivar and storage condition of apple (Fan et al., 1999; Watkins et al., 2000) as well as harvest maturity (Mir et al., 2001). For example, the general range of 0.5 to 2.0 µL·L–1 1-MCP is effective to retard flesh softening in apples.

Table 1. Treatment and storage conditions of four apple cultivars. Fruit were pretreated directly after harvest with 1-MCP for 18 h at 20 °C then stored at 1 °C.

<table>
<thead>
<tr>
<th>Cultivar</th>
<th>Harvest date</th>
<th>1-MCP concn (µL·L–1)</th>
<th>CA storage condition (kPa)</th>
<th>Experiment month</th>
</tr>
</thead>
<tbody>
<tr>
<td>Gala</td>
<td>4 Sept.</td>
<td>0.625</td>
<td>1</td>
<td>2</td>
</tr>
<tr>
<td>Delicious</td>
<td>16 Oct.</td>
<td>1.0</td>
<td>1</td>
<td>2</td>
</tr>
<tr>
<td>Granny Smith</td>
<td>16 Oct.</td>
<td>1.0</td>
<td>1.5</td>
<td>1</td>
</tr>
<tr>
<td>Fuji</td>
<td>31 Oct.</td>
<td>1.0</td>
<td>1.5</td>
<td>1</td>
</tr>
</tbody>
</table>

For ‘Empire’ and ‘Delicious’ cultivars, softening was retarded when treated by 1-MCP in the lower end of this range, while the higher end was necessary to get the same effect for ‘McIntosh’ and ‘Law Rome’ apples (Watkins et al., 2000). Inconsistencies in EthylBlock powder that occurred during the early years of production may make the concentration incorrect (Blankenship and Dole, 2003; Zanella, 2003), however, recent studies in apples indicate that 1.0 µL·L–1 1-MCP is effective for most of the cultivars (Blankenship and Dole, 2003; Watkins and Miller, 2004).

Controlled atmosphere (CA) storage has been used for apple commercial storage for decades, and has been recognized as the most practical and effective supplement to proper temperature management for extending the shelf life of intact and fresh-cut fruits and vegetables. About 50% of the U.S. apple production is held in CA storage, with recommended CA conditions of 1 to 3 kPa O2 and 1 to 5 kPa CO2 at 0 °C depending on the cultivar (Watkins et al., 2004).

It is not clear how use of 1-MCP on apples will interact with CA storage, and how CA storage of apples, with or without 1-MCP, compares with 1-MCP and subsequent air storage for optimal eating quality. Watkins et al. (2000) reported that the effects of 1-MCP are greater in CA than in regular air (RA) with ‘McIntosh’, ‘Empire’, ‘Delicious’, and ‘Law Rome’ apples, as also observed by DeLong et al. (2004) in ‘Cortland’ and ‘McIntosh’ apples. However, 1-MCP treatment maintained greater firmness and controlled scald of ‘Granny Smith’ apples regardless if in CA or RA (Zanella, 2003). Furthermore, a disorder, resembling external CO2 injuries, was observed on fruit treated with 1-MCP and subsequently stored in CA (Zanella, 2003). DeEll et al. (2005) compared the effects of standard CA and low O2 CA on storage quality of apples pretreated with or without 1-MCP. The results indicate that for ‘Empire’, ‘Gala’, and ‘Delicious’, there was no further quality benefit from combining low O2 CA with 1-MCP. In addition, several physiological disorders developed with the combination (DeEll et al., 2005).

High firmness, appropriate sugar to acid ratio, and aroma profile are considered important parameters for apple eating quality. Since sugars, the major component in the soluble solids, do not change much during storage, delayed loss of firmness and acidity are the most important indicators for evaluating apple storage practices. Volatile compounds produced by apples constitute aroma, however instrumental analysis of volatiles has indicated that there is a loss of aroma after long-term storage and due to treatment by 1-MCP (Fan and Mattheis, 1999;
cultivars. Preliminary work with 1-MCP on maintenance of apple firmness and sometimes acidity, and volatile losses when the fruit are stored in air (DeEll et al., 2002; Fan and Mattheis, 1999; Fan et al., 1999; Jaing and Joyce, 2002; Lurie et al., 2002; Mir et al., 2001; Rupasinghe et al., 2000). Some reports are of fruit in cold storage, ambient or combined cold followed by a simulated ambient marketing period. A few studies compared 1-MCP treatment of fruit stored in CA versus RA after cold storage compared to after a marketing period, for firmness and acidity, but not volatile losses (DeEll et al., 2005; DeLong et al., 2004; Watkins et al., 2000; Zanella, 2003).

In this research, we investigated the responses of major northwest apple cultivars to 1-MCP, CA, or 1-MCP + CA compared to apples stored in air. The objective of this study was to determine whether 1-MCP pretreatment and CA storage interact in terms of factors that affect apple eating quality, as determined by changes in firmness, titratable acidity, soluble solids content, and volatile abundance measured immediately after cold storage and after a week under simulated marketing conditions. Results will help industry make informed choices about use of 1-MCP in combination with storage atmosphere for optimal apple eating quality.

Materials and Methods

Early cultivar ‘Gala’ (harvested on 4 Sept.), midseason cultivars ‘Delicious’, ‘Granny Smith’ (harvested 16 Oct.) and late-ripening cultivar ‘Fuji’ (harvested 31 Oct.) apples (Malus sylvestris (L.) Mill var. domestica (Borkh.) Mansf) were harvested randomly from commercial orchards located in Wenatchee, Wash., in 2001 at the pre- to early-climacteric stage, as determined by internal ethylene concentration (Williams and Patterson, 1962) except for ‘Delicious’. Ethylene means ± standard deviation (n = 10) at harvest were 2.0 ± 1.4 for ‘Gala’, 2.3 ± 1.6 for ‘Fuji’, 19 ± 17 for ‘Delicious’, and undetectable for ‘Granny Smith’. Starch means (on a 1 to 6 scale) were 3.7 ± 1.2 for ‘Gala’, 4.6 ± 0.6 for ‘Fuji’, 2.2 ± 0.3 for ‘Delicious’, and 2.0 ± 0.1 for ‘Granny Smith’. For every cultivar, defect-free fruit were divided into two groups, one for 1-MCP treatment and another for nontreated control. Pretreatment of 1-MCP was immediately performed by exposing fruit to 1-MCP (SmartFresh, Rohm and Haas, Philadelphia, Pa.) at 20 °C for 18 h at USDA Tree Fruit Research Laboratory in Wenatchee using a 230-L steel chamber with sufficient headspace to prevent development of anaerobic respiration during treatment. The initial concentration of 1-MCP was determined by gas chromatography (GC) calibrated with 1-butene (Gong et al., 2002), and was 0.6 µL·L⁻¹ for ‘Gala’ and 1.0 µL·L⁻¹ for other cultivars. Preliminary work with 1-MCP on these cultivars had determined the effective concentrations used in this study and the fact that the European protocol for 1-MCP-treated fruit is 0.6 µL·L⁻¹.

Both 1-MCP-treated and nontreated groups were further divided to two batches, one for CA storage and another for regular atmosphere (RA) storage at 1 °C and 95% RH. The CA conditions were 1 to 1.5 kPa O₂ and 1 to 2 kPa CO₂, depending on cultivar (Table 1). Optimal CA conditions and length of storage have previously been established for these cultivars (Kupferman, 2003; Mattheis et al., 1998; Plotto et al., 1997).

Immediately after 1-MCP treatment, or every month after storage at 1 °C, fruit were shipped overnight to the USDA Citrus and Subtropical Products Laboratory in Winter Haven, Fla., with separate boxes for the 1-MCP-treated and nontreated fruit. Fruit temperatures were between 5 to 14 °C upon arrival. Every shipment included 40 fruit from each treatment and each cultivar, from which 20 fruit were immediately analyzed for the quality parameters upon arrival, and another 20 fruit were analyzed after storage at 20 °C for 1 week, to simulate the marketing conditions.

Measurements were obtained using five composite replicates of four fruit each. Flesh firmness and soluble solids content (SSC) were measured for each individual fruit and averaged per replicate. The SSC was measured using a digital, temperature-compensated refractometer (model PR-101; Atago Co. Tokyo, Japan) with freshly prepared juice.

Flesh firmness was assessed with a penetrometer (FT 327; McCormick, Facchini, Alfonsine, Italy), equipped with a cylindrical plunger 11 mm in diameter. The measurement was obtained from the equatorial area where 16 mm-diameter peel discs had been removed.

Titratable acidity (TA) and pH were measured from 50 g of flesh tissue obtained from four fruit per replicate and 50 mL deionized water, blended at speed 4 for 45 s with a homogenizer (model PT 10/35; Brinkmann Instruments, Co., Switzerland). The homogenate was vacuum-filtered through a 25- to 50-µm Büchner funnel. The filtrates (20 g) were measured for pH and then were titrated to pH 8.1 with 0.1 N NaOH using a Sage dispensing system (ATI Orion, Boston, Mass.), and the acidity was calculated as malic acid on a weight basis.

Volatile compounds were measured by headspace sampling [using an autosampler (NPS-2; Gerstel, Baltimore, Md.)] of tissue homogenate, and analyzed with a GC (model 6890; Agilent Technologies, Palo Alto, Calif.) equipped with a 0.53 mm × 30 m polar Stabil wax capillary column (1.0-µm film thickness, Restek, Bellefonte, Pa.) and a flame ionization detector (FID). Apple tissue (50 g) from four fruit per replicate was homogenized with 25 mL deionized water and 25 mL saturated NaCl solution (Bai et al., 2004). The homogenate (2

![](image)

**Fig. 1.** Firmness of four apple cultivars treated with 1-MCP or not, and stored in CA or RA at 1 °C (solid line); subsamples were transferred to 20 °C for an additional week (dotted line).
ml) was transferred to 20-ml glass vials sealed with a crimp-topped Teflon-silicone septum, flash frozen in liquid nitrogen and stored at –80 °C before analysis. Frozen samples were later thawed under running tap water, then were incubated in an agitator at 500 rpm and 40 °C for 2 min before the headspace sample (1 ml) was taken from the vial and injected into the GC. The analysis conditions were: oven temperature held at 40 °C for 6 min, then raised to 180 °C at a rate of 6 °C·min−1. The injection port and detector were kept at 250 and 280 °C, respectively. For a general picture of volatile production, including unknown compounds, total volatile abundance is reported in FID area response units rather than absolute amounts of individual analytes.

A split-plot analysis of variance (PROC ANOVA, Version 8, SAS Institute, Cary, N.C.) was used for statistical analysis to determine the effect of 1-MCP vs. non-1MCP, CA vs. RA, storage duration (0 to 8 months), and before vs. after transfer to 20 °C on SSC, flesh firmness, TA, and total volatile abundance. Means among each measurement were separated by the least significant difference at the 5% level (LSD 0.05). Regression analysis was carried out using PROC REG (SAS).

Results and Discussion

Firmness. 1-MCP and/or CA storage retarded the loss of firmness after the first 4 weeks of storage at 1 °C, compared with the control for all four cultivars (Fig. 1). Fruit treated with 1-MCP, regardless of storage atmosphere, remained firmer during storage at 1 °C; however, ‘Gala’ and ‘Delicious’ lost firmness over time in CA or air if not treated with 1-MCP. ‘Gala Smith’, stored in CA, lost firmness after 5 months storage. ‘Fuji’ is the only cultivar for which CA maintained fruit firmness similarly to 1-MCP in RA. However, over all of the four cultivars, fruit stored in CA generally maintained firmer flesh than that of control fruit stored in RA after the first 2 months (Fig. 1). These results are not surprising as it has been reported that 1-MCP-treated apples maintained firmness for ‘Redchief Delicious’ (Jayanty et al., 2004; Mir et al., 2001), ‘Fuji’, ‘Gala’, ‘Gingergold’, ‘Jonagold’, ‘Delicious’ (Fan et al., 1999), ‘Golden Delicious’ (Jiang and Joyce, 2002), ‘Cortland’, and ‘Empire’ apples (DeEll et al., 2002) in RA storage; for ‘Empire’, ‘Gala’ and ‘Delicious’ CA storage (DeEll et al., 2005); for ‘Gala Smith’ (Zanella, 2003), ‘McIntosh’, ‘Empire’, ‘Law Rome’ (Watkins et al., 2000), ‘Cortland’, and ‘McIntosh’ (DeLong et al., 2004) in CA and RA storage. DeLong et al. (2004) and Watkins et al. (2000) found that the combination of 1-MCP and CA was more effective in maintaining firmness than when 1-MCP-treated apples were stored in air as was found in this study.

When the fruit were transferred from 1 °C to 20 °C storage for 1 week, further firmness loss occurred in the control for all four cultivars, with average losses of 13%, 11%, 9%, and 4% in 1 week for ‘Gala’, ‘Delicious’, ‘Gala Smith’, and ‘Fuji’, respectively (Fig. 1). The firmness losses in CA-stored fruit after transfer to 20 °C were similar to those in control fruit for ‘Gala’, ‘Delicious’, and ‘Gala Smith’, but no significant decrease was observed for ‘Fuji’. Treatments with 1-MCP prevented the rapid firmness loss upon transfer of fruit to room temperature, regardless of storage atmosphere. Following reports support the 1-MCP prevention to firmness loss in shelf life at room temperatures: DeEll et al. (2005) with ‘Gala’ and ‘Delicious’ in CA, Watkins et al. (2000) with ‘Delicious’ and Zanella (2003) with ‘Gala Smith’ in RA and CA.

Table 2. F value, significant levels and CV (%) from analysis of variance performed on firmness, titratable acidity (TA), and volatile abundance (VA) of four apple cultivars pretreated or not with 1-MCP, stored in CA or RA at 1 °C (solid line); subsamples were transferred to 20 °C for 1 additional week (dotted line).

<table>
<thead>
<tr>
<th>Analysis</th>
<th>Firmness</th>
<th>TA</th>
<th>VA</th>
<th>Firmness</th>
<th>TA</th>
<th>VA</th>
<th>Firmness</th>
<th>TA</th>
<th>VA</th>
<th>Firmness</th>
<th>TA</th>
<th>VA</th>
</tr>
</thead>
<tbody>
<tr>
<td>T-MCP (M)</td>
<td>1</td>
<td>1035.31</td>
<td>284.11</td>
<td>394.22</td>
<td>1</td>
<td>1696.52</td>
<td>60.52</td>
<td>312.99</td>
<td>1</td>
<td>1816.08</td>
<td>137.89</td>
<td>25.23</td>
</tr>
<tr>
<td>CA (C)</td>
<td>1</td>
<td>102.57</td>
<td>111.17</td>
<td>212.17</td>
<td>1</td>
<td>171.86</td>
<td>17.45</td>
<td>128.77</td>
<td>1</td>
<td>455.36</td>
<td>2.88</td>
<td>116.16</td>
</tr>
<tr>
<td>Duration (D)</td>
<td>6</td>
<td>16.94</td>
<td>38.27</td>
<td>98.58</td>
<td>8</td>
<td>34.57</td>
<td>65.16</td>
<td>20.81</td>
<td>8</td>
<td>45.67</td>
<td>50.79</td>
<td>36.64</td>
</tr>
<tr>
<td>Transfer (T)</td>
<td>1</td>
<td>58.61</td>
<td>88.96</td>
<td>368.29</td>
<td>1</td>
<td>96.07</td>
<td>0.02</td>
<td>168.81</td>
<td>1</td>
<td>27.09</td>
<td>0.29</td>
<td>66.37</td>
</tr>
<tr>
<td>M × C</td>
<td>6</td>
<td>59.45</td>
<td>30.36</td>
<td>110.28</td>
<td>1</td>
<td>32.65</td>
<td>0.43</td>
<td>15.86</td>
<td>1</td>
<td>690.68</td>
<td>0.1</td>
<td>20.18</td>
</tr>
<tr>
<td>M × T</td>
<td>6</td>
<td>3.33</td>
<td>6.22</td>
<td>7.92</td>
<td>1</td>
<td>24.16</td>
<td>1.73</td>
<td>6.28</td>
<td>1</td>
<td>68.88</td>
<td>2.85</td>
<td>6.41</td>
</tr>
<tr>
<td>M × T + D</td>
<td>6</td>
<td>42.62</td>
<td>12.57</td>
<td>4.85</td>
<td>1</td>
<td>141.07</td>
<td>3.77</td>
<td>10.16</td>
<td>1</td>
<td>73.83</td>
<td>23.48</td>
<td>3.16</td>
</tr>
<tr>
<td>C × T</td>
<td>3</td>
<td>5.17</td>
<td>9.04</td>
<td>5.38</td>
<td>1</td>
<td>5.24</td>
<td>2.27</td>
<td>2.59</td>
<td>1</td>
<td>20.32</td>
<td>5.01</td>
<td>4.61</td>
</tr>
<tr>
<td>C × T + D</td>
<td>3</td>
<td>5.17</td>
<td>9.04</td>
<td>5.38</td>
<td>1</td>
<td>5.24</td>
<td>2.27</td>
<td>2.59</td>
<td>1</td>
<td>20.32</td>
<td>5.01</td>
<td>4.61</td>
</tr>
<tr>
<td>D × T</td>
<td>6</td>
<td>1.42</td>
<td>1.76</td>
<td>3.75</td>
<td>7</td>
<td>4.69</td>
<td>4.87</td>
<td>1.65</td>
<td>7</td>
<td>5.74</td>
<td>9.85</td>
<td>2.35</td>
</tr>
<tr>
<td>CV (%)</td>
<td>4</td>
<td>6.96</td>
<td>7.71</td>
<td>1.52</td>
<td>6.48</td>
<td>10.51</td>
<td>1.5</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

**Significant at p < 0.05 or 0.01.
An analysis of variance indicated that there were significant differences between 1-MCP pretreatment and no pretreatment, CA and RA, and among different storage durations for all of the four cultivars (Table 2). Comparisons of before and after transferring fruit to 20 °C for 1 week showed significant differences for ‘Gala’, ‘Delicious’, and ‘Granny Smith’, but not ‘Fuji’.

The interaction between pretreatments (1-MCP vs. non-1-MCP) and storage atmospheres (CA vs. RA) was significant statistically (Table 2), but practically the combination of 1-MCP + CA showed only a small difference in firmness compared to 1-MCP + RA for ‘Gala’, ‘Delicious’ and ‘Granny Smith’, and from 1-MCP + RA or CA for ‘Fuji’ (Fig. 1).

Titratable acidity and soluble solids. The TA decreased during 1 °C storage for ‘Gala’, ‘Delicious’, and ‘Fuji’, but increased slightly in first 2 months and then decreased for ‘Granny Smith’ regardless of treatment (Fig. 2). Both CA and 1-MCP retarded the decrease in TA, although 1-MCP was more effective than CA for ‘Gala’, and ‘Delicious’ toward the end of storage. 1-MCP + RA was similar to CA for retarding loss of acidity in all but ‘Fuji’, where the 1-MCP + RA was less effective. A synergistic reaction between 1-MCP and CA was observed for ‘Gala’ and ‘Delicious’, but not for ‘Granny Smith’ and ‘Fuji’ in terms of maintaining TA (Fig. 2, Table 2).

Transferring fruit to 20 °C for 1 week accelerated TA losses for most of the cultivars and treatments (Fig. 2). The average TA loss caused by transferring fruit to 20 °C for 1 week, for control fruit over the storage duration was 14%, 5%, 12%, and 11% for ‘Gala’, ‘Delicious’, ‘Granny Smith’, and ‘Fuji’, respectively. For ‘Gala’ and ‘Granny Smith’, the temperature-dependent TA loss was retarded by CA and/or 1-MCP, within which 1-MCP was more effective than CA. However, for ‘Delicious’ and ‘Fuji’, CA and 1-MCP did not significantly retard TA loss (Fig. 2, Table 2).

It has been reported that 1-MCP inhibited loss of TA in both pre-climacteric and climacteric apple fruit (‘Fuji’, ‘Gala’, ‘Ginger Gold’, ‘Jogold’ and minimally for ‘Delicious’) (Fan et al., 1998) reported similar patterns of ‘Gala’ volatile production during storage. However, there were significant reductions of total volatile abundance after transferring fruit from 1 °C to 20 °C for ‘Gala’, ‘Delicious’, and ‘Fuji’, but not ‘Granny Smith’ (Fig. 3). Volatile abundance of fruit in RA increased within the first 2 months then decreased in ‘Gala’, continued to increase until the sixth month in ‘Fuji’ (Fig. 3A and D) or remained at the same level in ‘Delicious’ during storage at 1 °C (Fig. 3B). Volatiles did not increase until the third to fourth month in ‘Granny Smith’ and remained level (Fig. 3C). The increase of volatiles indicates that the fruit continued ripening during storage, and the subsequent decrease indicates that the fruit were undergoing senescence. Mattheis et al. (1998) reported similar patterns of ‘Gala’ volatile production during storage. However, there were unique positive linear regressions of firmness and TA content for every cultivar (data not shown). For the same firmness level, ‘Granny Smith’ had higher TA content, and ‘Fuji’ had lower TA content than other cultivars (Figs. 1 and 2).

Not surprisingly, there was a negative regression between TA and pH. A small change in TA content led to a great change in pH for ‘Fuji’ fruit, but a big change in TA content led to a small response in pH for ‘Granny Smith’ apple (data not shown).

CA and 1-MCP slightly retarded the loss of SSC in ‘Gala’ during 1 °C storage and after transfer to 20 °C, but did not affect SSC in other cultivars (data not shown). Most of the reports show no major effects of 1-MCP or CA on SSC (DeLong et al., 2004; Fan et al., 1999; Rupasinghe et al., 2000; Zanella, 2003). However, Watkins et al. (2000) found that 1-MCP lowered SSC levels in ‘McIntosh’ and ‘Delicious’ apples in both air and CA storage, and in ‘Law Rome’ in CA, but had no effect on ‘Empire’.

Volatile abundance. There was much less total volatile production from fruit treated with 1-MCP and CA compared with that of the control (RA-stored fruit) throughout the entire 1 °C storage and after transfer to 20 °C for all four cultivars (Fig. 3). Volatile abundance of fruit in RA increased within the first 2 months then decreased in ‘Gala’, continued to increase until the sixth month in ‘Fuji’ (Fig. 3A and D) or remained at the same level in ‘Delicious’ during storage at 1 °C (Fig. 3B). Volatiles did not increase until the third to fourth month in ‘Granny Smith’ and remained level (Fig. 3C). The increase of volatiles indicates that the fruit continued ripening during storage, and the subsequent decrease indicates that the fruit were undergoing senescence. Mattheis et al. (1998) reported similar patterns of ‘Gala’ volatile production during storage. However, there were significant reductions of total volatile abundance after transferring fruit from 1 °C to 20 °C for ‘Gala’, ‘Delicious’, and ‘Fuji’, but not ‘Granny Smith’ (Fig. 3). Volatile accumulation, as measured in this study, depends on the production of volatiles by the fruit in relation to loss of volatiles due to off-gassing from (or diffusing out of) the fruit. Since transferring apples from 1 °C to 20 °C usually increases metabolism and the production of volatiles, the decreased volatile levels at 20 °C may have been due to increased diffusion of volatiles out of the fruit. ‘Gala’, ‘Delicious’, and ‘Fuji’ apples produce mostly esters, as opposed to ‘Granny Smith’, which produces a majority of alcohols (Bai et al., 2002). According to Henry’s Law, esters diffuse more readily than alcohols out of the fruit (Bai et al., 2002).
The magnitude of volatile inhibition by 1-MCP alone (1-MCP + RA) was generally similar to that of CA during storage at 1 °C, except for ‘Delicious’ until late storage (Fig. 3). However, total volatile abundance in 1-MCP-pretreated fruit was often similar to or lower than that of fruit stored in CA after transfer to 20 °C (Fig. 3). Inhibition of total volatile abundance by the 1-MCP + CA combination treatment was the most severe compared to either treatment alone (Fig. 3).

CA and modified atmosphere (MA) decrease volatile production of apples (Mattheis et al., 1998; Patterson et al., 1974). Production of lipid-derived volatiles is inhibited by reduced O₂ levels, while volatiles that arise from amino acid catabolism are negatively impacted by elevated CO₂ (Brackmann et al., 1993; Abdi et al., 1998). There were significant differences in volatile production among the apple cultivars treated with and without 1-MCP, and CA on volatile production in relation to apple cultivar, 1-MCP exposure (dosage, duration and temperature) and CA condition.

Conclusion

1-MCP with and without CA was tested on ‘Gala’, ‘Delicious’, ‘Granny Smith’, and ‘Fuji’ apples at levels now used commercially, to determine individual and interactive effects on eating quality factors during cold storage and subsequent simulated marketing conditions. There were significant differences found for firmness, TA and volatile abundance among the apple fruits treated with and without 1-MCP, stored in CA versus RA, over storage time, and before and after transfer to marketing temperature. In comparison with CA, 1-MCP + RA treatments were more effective in prolonging storage life as indicated by maintenance of firmness and TA in all cultivars except ‘Fuji’. In agreement with Watkins et al. (2000), results showed that a combination of 1-MCP + CA was more effective than either individual treatment alone for maintaining firmness, but our study showed this was at the expense of the highest aroma volatile losses. Therefore, 1-MCP + RA may be a practical alternative to CA storage of apples, while maintaining optimal flavor quality for some cultivars.

Literature Cited


