

Effects of nitrogen on *Pinus palustris* foliar respiratory responses to elevated atmospheric CO₂ concentration

R.J. Mitchell^{1,6}, G.B. Runion², S.A. Prior³, H.H. Rogers³, J.S. Amthor⁴ and F.P. Henning⁵

¹ Jones Ecological Research Center, RR2, Box 2324, Newton, GA 31770, USA

² School of Forestry, Auburn University, AL 36849, USA

³ USDA-ARS, National Soil Dynamics Laboratory, Auburn, AL 36849, USA

⁴ Global Climate Research Division, Lawrence Livermore National Laboratory, Livermore, CA 94550, USA

⁵ Agronomy Department, Auburn University, AL 36849, USA

Received 6 March 1995; Accepted 5 June 1995

Abstract

Indirect effects of atmospheric CO₂ concentration [CO₂], on longleaf pine (*Pinus palustris* Mill.) foliage respiration were studied by growing trees in a factorial arrangement of low and high [CO₂] (369 and 729 μmol CO₂ mol⁻¹) and low and high N (40 and 400 kg ha⁻¹ yr⁻¹). Direct effects of [CO₂] on leaf respiration were tested by measuring respiration rates of foliage from all treatments at two CO₂ levels (360 and 720 μmol CO₂ mol⁻¹) at the time of measurement. Elevated CO₂ did not directly or indirectly affect leaf respiration when expressed on a leaf area or mass basis, but a significant increase in respiration per unit leaf N was observed in trees grown in elevated [CO₂] (indirect response to elevated [CO₂]). The lack of a [CO₂] effect on respiration, when analysed on an area or mass basis, may have resulted from combined effects of [CO₂] on factors that increase respiration (e.g. greater availability of non-structural carbohydrates stimulating growth and carbon export from leaves) and on factors that decrease respiration (e.g. lower N concentration leading to lower construction costs and maintenance requirements). Thus, [CO₂] affected factors that influence respiration, but in opposing ways.

Key words: *Pinus palustris*, elevated CO₂, nitrogen, foliar respiration.

Introduction

Due to burning of fossil fuels and changes in land use world-wide, atmospheric CO₂ concentration [CO₂], has

increased during the past 100 years at a rate that may be unprecedented in the historical records for the past 160 000 years (Raynaud *et al.*, 1993; Sundquist, 1993). [CO₂] affects a wide range of plant functions, both directly and indirectly, and so increasing atmospheric [CO₂] may alter plant function and community structure (Field *et al.*, 1992). However, predicting the direction, much less the magnitude, of changes in plant function is difficult since there is a lack of understanding of the mechanisms that control overall plant response to [CO₂] (Mooney, 1991). Most studies of plant responses to elevated [CO₂] have focused on photosynthesis and growth of crops and trees (Strain, 1987; Bowes, 1993; Rogers *et al.*, 1994). Much less effort has been directed to understanding respiratory responses to elevated [CO₂], even though respiration may consume more than 50% of the carbon fixed at the whole plant level (Amthor, 1989), and despite evidence suggesting that CO₂ may directly and indirectly influence respiration (see Amthor, 1991, 1995; Ryan, 1991; Wullschleger *et al.*, 1994, for recent reviews). The extent to which respiration is increased or decreased by elevated [CO₂] is not known, however, extant results conflict, in part, because of non-conformity in methods used to quantify respiration and the manner in which respiration rate has been expressed (Ryan, 1991; Wullschleger *et al.*, 1994). Also, respiration is often measured only at one point in time, although variation in respiration has been observed due to plant ontogeny (Poorter *et al.*, 1992; Mousseau, 1993). Farrar and Williams (1991) suggest that solitary measurements of leaf or whole plant respiration are of little use in understanding respiratory responses and their relationship to growth; rather, time-course

⁶ To whom correspondence should be addressed. Fax: +1 912 734 4707.

studies are needed. Furthermore, the procedure by which respiration is measured may complicate comparisons of different studies. Several investigators have determined respiration rates by shading the plant or leaf during the day (Ziska *et al.*, 1990) or by extrapolating respiratory rates from light response curves (Hicklenton and Jolliffe, 1978; Nijs *et al.*, 1988). These estimates of respiratory rates may differ substantially from steady-state night-time respiration (Wullschleger *et al.*, 1994), possibly as a result of alteration in diurnal sink-source relationships. Additional complications can arise because the $[\text{CO}_2]$ during a measurement may affect respiration rate. For example, Amthor *et al.* (1992) found that apparent respiration rate was inhibited 20–30% by a $300 \mu\text{mol mol}^{-1}$ increase in $[\text{CO}_2]$ during a respiration measurement. This effect was readily reversible, i.e. a decrease in $[\text{CO}_2]$ during a measurement increased respiration rate. Although this response to short-term changes in $[\text{CO}_2]$ at the time of measurement has been commonly observed (Begg and Jarvis, 1968; Gale, 1982; Reuveni and Gale, 1985; Bunce, 1990, 1992; El Kohen *et al.*, 1991; Mousseau, 1993; Ziska and Bunce, 1993, 1994; Downton and Grant, 1994; Villar *et al.*, 1994), it is unknown whether changes in apparent respiration represent a direct effect of $[\text{CO}_2]$ on respiration *per se* or reflect the influence of $[\text{CO}_2]$ on other metabolic reactions such as dark fixation of HCO_3^- by PEPcase (Amthor, 1995).

Difficulty in comparing studies of respiratory responses to elevated $[\text{CO}_2]$ is exacerbated by variability in the manner in which respiration rates are expressed. That is, respiration rates have been expressed in numerous ways (e.g. per unit dry mass, leaf area, and protein content) and long-term elevated $[\text{CO}_2]$ can alter relationships between plant mass, leaf area, and protein content. For example, elevated $[\text{CO}_2]$ often increases plant C:N ratio (Norby *et al.*, 1986, 1992; Field *et al.*, 1992) and, thus, may alter relationships between respiration per unit dry mass and respiration per unit N or protein.

Although recent work has indicated that effects of $[\text{CO}_2]$ on photosynthesis and productivity may be mediated by nutrients (Larigauderie *et al.*, 1988; Eamus and Jarvis, 1989; Bazzaz, 1990; Fajer *et al.*, 1992; Griffin *et al.*, 1993), the extent to which elevated $[\text{CO}_2]$ increases N use efficiency, thus ameliorating N deficiencies, is controversial (see Norby *et al.*, 1986; Coleman *et al.*, 1991; Hilbert *et al.*, 1991, for contrasting views). Moreover, even though the availability of N has been shown to elicit strong respiratory responses (Ryan, 1991), little effort has been given to determining how soil fertility may influence respiratory responses to elevated $[\text{CO}_2]$.

This study examined the respiration response of long-leaf pine (*Pinus palustris* Mill.) foliage from trees grown in ambient and elevated atmospheric $[\text{CO}_2]$ and at two levels of soil N. All respiration measurements were conducted at night in the dark. Two $[\text{CO}_2]$ (360 and $720 \mu\text{mol}$

$\text{CO}_2 \text{ mol}^{-1}$) were used during measurements to assess any direct effects of $[\text{CO}_2]$ on respiration rate. Also, effects of N on respiration and on respiratory responses to $[\text{CO}_2]$ were quantified. Results are reported per unit dry mass, leaf surface area and foliar N content.

Materials and methods

Plant growth and exposure system

Longleaf pine (*Pinus palustris* Mill.) seedlings, from a wild seed source, were lifted from a Florida nursery in February 1993. Seedlings were stored (5°C) for less than 1 week, graded (root collar diameter mean = 13 mm; standard deviation = 2 mm; range = 9 to 21 mm), and planted (3 per pot) into each of 192, 45 l black plastic pots containing construction grade sand (pH 5.1) which was low in mineral elements. Seedlings were irrigated with deionized water as needed.

Seedlings were exposed to target $[\text{CO}_2]$ of $365 \mu\text{mol CO}_2 \text{ mol}^{-1}$ (ambient) or $720 \mu\text{mol CO}_2 \text{ mol}^{-1}$ (elevated) within an open top chamber system similar to that described by Rogers *et al.* (1983). The chambers, which are 3 m in diameter and 2.4 m in height, are constructed of a structural aluminium frame covered by a clear PVC plastic film (0.2 mm thickness). Carbon dioxide is supplied from a 12.7 Mg liquid CO_2 receiver through a high volume dispensing manifold and added to the chambers by injection into plenum boxes (Rogers *et al.*, 1983). Air is dispensed into each chamber through the bottom half of each chamber cover which is double-walled; the inside wall is perforated with 2.5 cm diameter holes to serve as ducts that distribute air uniformly into the chamber. Carbon dioxide concentrations in each chamber are monitored 24 h d^{-1} using a time-shared sampling manifold; a solenoid bank directed samples to an infra-red gas analyser which recorded an instantaneous value at the end of that sampling period (Rogers *et al.*, 1983). Carbon dioxide exposures were initiated on 30 March 1993, and $[\text{CO}_2]$ values were continuously recorded every 15–30 min for each chamber, depending upon whether or not an additional CO_2 study was on line. Average ambient $[\text{CO}_2]$ (\pm standard deviation) through the duration of the experiment was $368.6 \mu\text{mol mol}^{-1}$ (± 17.1) during the day (7 a.m. to 7 p.m.) and 403.1 (± 34.4) during the night. Average $[\text{CO}_2]$ in the elevated chambers was $728.9 \mu\text{mol mol}^{-1}$ (± 52.1) during the day and 792.9 (± 53.5) during the night. In order to maintain temperatures within 3°C of ambient, three chamber volumes were exchanged every minute.

Nitrogen treatments were similar to those described by Bazzaz and Miao (1993). They consisted of applying either 0.20 or 0.02 mg N g^{-1} soil per year (as sulphur-coated urea; 38–0–0 [N–P–K]) which correspond to high (400 kg $\text{N ha}^{-1} \text{ yr}^{-1}$) and low (40 kg $\text{N ha}^{-1} \text{ yr}^{-1}$) N treatments. Fertility of other nutrients was maintained at non-limiting levels in all pots by application of sulphur-coated potassium (0–0–47 [N–P–K]; 0.04 mg K g^{-1} soil yr^{-1}) and MicroMax™ Plus (0–4–0 [N–P–K]; P=0.14, Ca=0.57, Mg=0.28, and S=0.05 mg g^{-1} soil yr^{-1} , plus a complete complement of micronutrients) which was mixed into the sand at the time the pots were filled. Iron chelate (0.007 mg Fe g^{-1} soil) was applied once in April 1993.

Treatments were arranged in a 2×2 factorial design with six replications. Carbon dioxide treatments were randomly assigned to chambers while N treatments were randomly assigned to a total of 16 pots within each chamber. To avoid location effects within a chamber, pot locations were re-randomized monthly.

Respiration measurements

Respiration was measured four times over a 6-week period. Average daily temperature (range=27–30 °C) and solar radiation (range=5652–6521 W m⁻²) on the day of respiration measurement varied only slightly throughout this 6-week period. On each seedling, a cohort of actively growing needles (that expanded under previously described CO₂ treatments) was delimited with pipe cleaners. Starting 25 June 1993, and continuing every 2 weeks for a 6-week period, one fascicle from the delimited cohort of fascicles from each seedling was excised. All fascicles within a treatment were immediately enclosed in a 1 l cuvette and CO₂ efflux was measured, over a 30–90 s period, with a Li-Cor 6200 portable photosynthesis system (Li-Cor, Inc., Lincoln, NE). Measurements were started at 9 p.m. and continued until approximately 3 a.m. and were conducted in the dark. All measurements were conducted at two CO₂ partial pressures (360 and 720 μmol mol⁻¹, the order of which was randomly assigned) within the cuvettes. Respiration rate was determined by the amount of time required to change [CO₂] by 6 μmol mol⁻¹. After all respiration rate measurements during a single night were made, the average time per measurement was calculated and, in an empty (no fascicles) cuvette, the change in [CO₂] during the average measurement time (≈45 s) was determined at both 360 and 720 μmol mol⁻¹ CO₂. These values were used to correct the respiration rate measurements for any leaks in the system. Changes in cuvette [CO₂] due to leaks were always low compared to changes in [CO₂] with needles present; in most cases, leak rates were at least 10-fold less than respiration measurements.

After respiration rate determinations, needle surface area was measured. Needles were then dried at 55 °C for 4 d (to constant mass) and fascicle dry mass was recorded. Foliar N and C content per fascicle was then determined with a LECO CHN-600 analyser (LECO Corp. St Joseph, MI). Total non-structural carbohydrates (starch plus sugar) were analysed according to the methods described by Thomas *et al.* (1993). Construction costs (g glucose required to construct 1 g dry weight of needle tissue) were determined using a bomb calorimeter and adjusted for ash content and N concentration, as described by Williams *et al.* (1987), Griffin *et al.* (1993), and Amthor *et al.* (1994).

Data analysis

Treatment main effects and interactions were tested using a repeated measures multivariate analysis of variance. The main effects of [CO₂] and N were determined using a split-plot factorial analysis with CO₂ treatments as the whole plot and N treatments as the subplot. Trends through time and interactions of main effects with measurement dates were tested using the Wilk's lambda test.

Results and discussion

Increasing [CO₂] did not alter apparent respiration rate of *Pinus palustris* foliage either directly (cuvette [CO₂]) or indirectly (open top chamber [CO₂]) when expressed on a leaf area or dry mass basis (Table 1). Needle respiration rates averaged 1.04 μmol CO₂ m⁻² s⁻¹ and 10.8 nmol CO₂ g⁻¹ s⁻¹, respectively. Nitrogen supply strongly influenced respiration rate per unit needle mass, per unit leaf area, and per unit foliar N content (Table 1). Longleaf pine foliage from trees grown in the high N treatment showed a 45% increase in respiration rate on a leaf area basis and a 54% increase on a dry mass basis when compared to trees grown at the low N treatment. Although respiration (expressed either per unit leaf area or per unit leaf dry mass) demonstrated a significant N by measurement date interaction (data not shown), this interaction was one of magnitude rather than rank (Fig. 1). Respiration rates per unit foliar N were significantly influenced by [CO₂], N treatment, and their interaction (Fig. 2). Long-term elevated atmospheric [CO₂] resulted in greater respiration rates per unit foliar N;

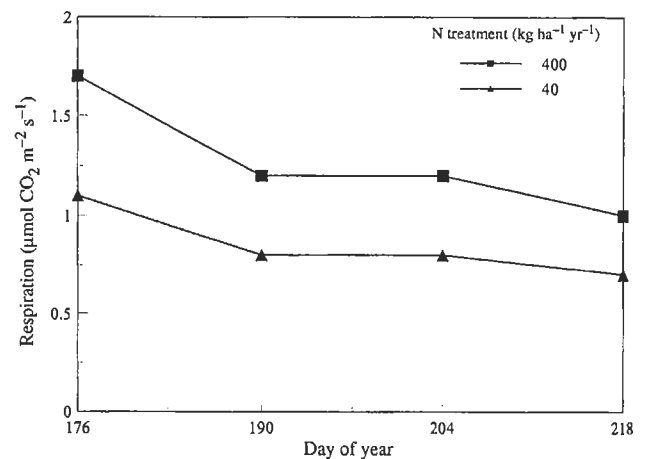


Fig. 1. Respiration of longleaf pine foliage per unit leaf area for the low (40 kg ha⁻¹ yr⁻¹) and high (400 kg ha⁻¹ yr⁻¹) nitrogen treatments across the four measurement dates ($Pr > F = 0.0001$ for the nitrogen treatment and $Pr > F = 0.0003$ for the date by N interaction).

Table 1. Effects of long-term (open top chamber) [CO₂] (indirect), nitrogen, and short-term (cuvette) [CO₂] (direct) treatments on CO₂ efflux from longleaf pine needles expressed per unit: needle area, needle dry mass, and needle N

Respiration measurement	Main effect treatment variable								
	Long-term [CO ₂] (μmol CO ₂ mol ⁻¹)			N (kg ha ⁻¹ yr ⁻¹)			Cuvette [CO ₂] (μmol CO ₂ mol ⁻¹)		
	360	720	Pr > F ^a	40	400	Pr > F	360	720	Pr > F
μmol CO ₂ m ⁻² s ⁻¹	1.04	1.05	0.937	0.85	1.24	<0.001	1.05	1.04	0.761
nmol CO ₂ g ⁻¹ s ⁻¹	10.90	10.66	0.584	8.50	13.06	<0.001	10.82	10.74	0.797
nmol CO ₂ mg ⁻¹ N s ⁻¹	1.04	1.22	0.007	1.21	1.06	<0.001	1.14	1.13	0.876

^a Probability of a greater *F* value by chance for the difference between the CO₂ or N treatments, and for the CO₂ by N interaction, where the interaction was significant.

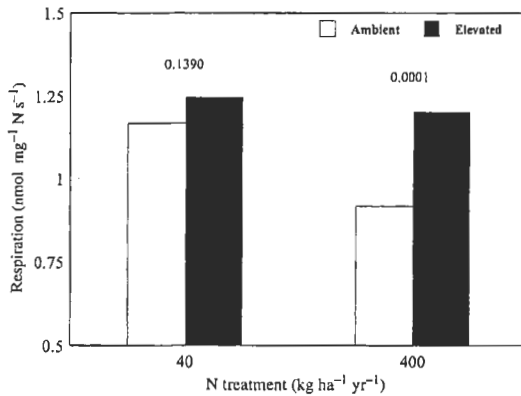


Fig. 2. Respiration of longleaf pine foliage per unit of foliar nitrogen for the ambient ($360 \mu\text{mol mol}^{-1}$) and elevated ($720 \mu\text{mol mol}^{-1}$) CO_2 treatments at the low ($40 \text{ kg ha}^{-1} \text{ yr}^{-1}$) and high ($400 \text{ kg ha}^{-1} \text{ yr}^{-1}$) nitrogen treatments. Values above each set of bars are probabilities of a greater F from contrasts run under General Linear Models. The CO_2 by N interaction was significant ($\text{Pr} > F = 0.0087$).

however, this effect was much greater in the high N treatment than in the low N treatment, even though increasing N availability reduced respiration rate per unit foliar N. Similarly, N had a greater influence on dark respiration rate of loblolly pine (*Pinus taeda* L.) shoots than did $[\text{CO}_2]$ under similar $[\text{CO}_2]$ and N supply and with a similar effect of N supply on tissue N concentration (Griffin *et al.*, 1993).

It has been suggested that increased atmospheric $[\text{CO}_2]$ can influence plant respiration directly (i.e. a response to $[\text{CO}_2]$ within minutes that is reversible) and indirectly (Amthor, 1991). The indirect response might be mediated by altered N or carbohydrate content which, in turn, regulate growth and respiration rates. A common response to short-term step changes in $[\text{CO}_2]$ has been a reduction in respiration rate (Amthor, 1995). Direct effects of $[\text{CO}_2]$ on respiration, however, may be affected by plant ontogeny. For example, El Kohen *et al.* (1991) observed an immediate and readily reversible decrease in springtime dark respiration rate of leaves and whole shoots of 2-year-old sweet chestnut (*Castanea sativa* Mill.) trees grown in 350 or $700 \mu\text{mol mol}^{-1} \text{CO}_2$, but the effect of $[\text{CO}_2]$ on respiration rate decreased through summer and was negligible in autumn. In addition, several reports suggest that plants grown in elevated $[\text{CO}_2]$ for long time periods are less responsive to short-term changes in $[\text{CO}_2]$ compared with plants grown in ambient $[\text{CO}_2]$ (Bunce, 1990; El Kohen *et al.*, 1991; Thomas and Griffin, 1994). Amthor (1995) suggests that the direct inhibition of apparent respiration from short-term night-time increases in $[\text{CO}_2]$ may be due to dark CO_2 fixation by PEP carboxylase activity, rather than a respiratory response *per se*. Further research is required to resolve this issue.

Indirect (long-term) effects of atmospheric $[\text{CO}_2]$ on plant respiration per unit dry mass have been variably reported to increase (Gifford *et al.*, 1985; Hrubec *et al.*,

1985; Kendall *et al.*, 1985; Williams *et al.*, 1992), decrease (Gifford *et al.*, 1985; Kendall *et al.*, 1985; Spencer and Bowes, 1986; Azcón-Bieto *et al.*, 1994), and remain constant (Gifford *et al.*, 1985; Hrubec *et al.*, 1985; Baker *et al.*, 1992; Azcón-Bieto *et al.*, 1994 [C_4 species]; Downton and Grant, 1994) in elevated $[\text{CO}_2]$ (reviewed in Amthor, 1995). In many cases, maintenance respiration rates have been shown to be correlated with tissue N content (Ryan, 1991), and elevated $[\text{CO}_2]$ often decreases N content of tissue (Field *et al.*, 1992). This decrease in N content of plants grown in elevated $[\text{CO}_2]$ has been suggested to be an important factor in decreasing respiration per unit dry mass in plants grown in elevated CO_2 (Amthor, 1991, 1995; Wullschlegel *et al.*, 1994). Yet several reports indicate that respiration rate per unit N (or protein) is depressed following long-term growth in elevated $[\text{CO}_2]$ (Ziska and Bunce, 1993; Ceulemans and Mousseau, 1994). The data presented here, and those of Griffin *et al.* (1993), are contrary to this trend. Foliar respiration of longleaf pine was strongly related to N concentration; however, greater respiration rates per unit N were observed when trees were grown in elevated $[\text{CO}_2]$, at least for the high N treatment. Griffin *et al.* (1993) also found a decrease in N concentration and an increase in apparent respiration rate due to elevated $[\text{CO}_2]$ with a high N treatment; thus, apparent respiration per unit N increased.

Atmospheric $[\text{CO}_2]$ may also influence apparent respiration rates by altering carbohydrate availability. Non-structural carbohydrate concentration of foliage in this study was increased by elevated $[\text{CO}_2]$ and decreased by increased N availability (Fig. 3); the interaction among main treatment variables was not significant. Several reports of increased respiration rates due to long-term elevated atmospheric $[\text{CO}_2]$ exposure have indicated that

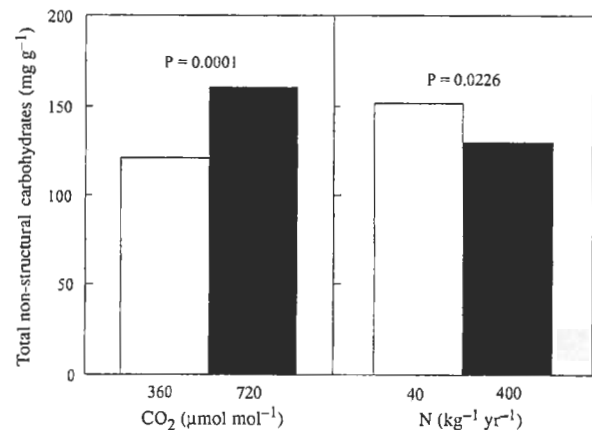


Fig. 3. Total non-structural carbohydrate content of longleaf pine foliage for the ambient ($360 \mu\text{mol mol}^{-1}$) and elevated ($720 \mu\text{mol mol}^{-1}$) CO_2 treatments and for the low ($40 \text{ kg ha}^{-1} \text{ yr}^{-1}$) and high ($400 \text{ kg ha}^{-1} \text{ yr}^{-1}$) nitrogen treatments. Values above each set of bars are probabilities of a greater F from contrasts run under General Linear Models. The CO_2 by N interaction was not significant ($\text{Pr} > F = 0.7456$).

increased availability of non-structural carbohydrates may stimulate respiration rates (Azcón-Bieto and Osmond, 1983; Hrubec *et al.*, 1985; Poorter *et al.*, 1992; Williams *et al.*, 1992). Azcón-Bieto *et al.* (1983) suggested that respiration is limited by the supply of glucose and fructose to mitochondria, although Amthor (1994) suggested that, with the exception of actively growing cells, respiration is regulated mainly by the rate at which respiratory products (ATP, NAD(P)H, and carbon skeletons) are used to meet the metabolic demands of growth, maintenance, transport, and ion uptake. Perhaps the increased photosynthate produced in these trees grown in elevated [CO₂] (data not shown) was—in addition to accumulating in leaves—being exported out of the leaves at greater rates and being used for additional leaf growth, both of which would result in increased respiration. The decrease in leaf N content, however, may have partially offset increased respiration as a result of reduced costs of growth and maintenance so that no net effect of [CO₂] on respiration rate was observed. In this study, the magnitude of the decrease in growth cost due to CO₂ enrichment was small (1.6%) while increasing N supply increased construction costs by a similar magnitude (2.5%). The interaction among main treatment variables was not significant (Fig. 4). The effects of [CO₂] on

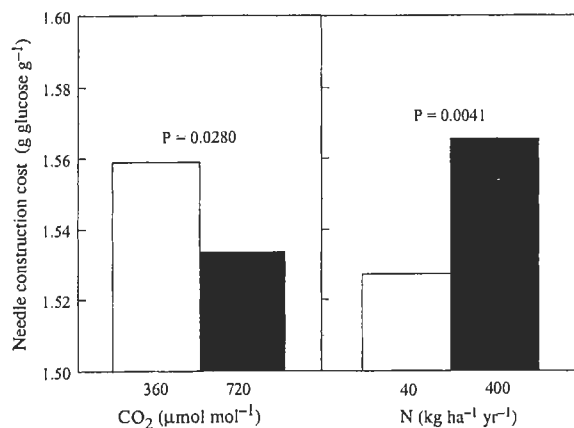


Fig. 4. Costs of constructing longleaf pine foliage for the ambient (360 $\mu\text{mol mol}^{-1}$) and elevated (720 $\mu\text{mol mol}^{-1}$) CO₂ treatments at the low (40 $\text{kg ha}^{-1} \text{yr}^{-1}$) and high (400 $\text{kg ha}^{-1} \text{yr}^{-1}$) nitrogen treatments. Values above each set of bars are probabilities of a greater *F* from contrasts run under General Linear Models. The CO₂ by N interaction was not significant ($\text{Pr} > F = 0.1757$).

growth cost that were estimated in this study are about the same as those calculated by Griffin *et al.* (1993) in both magnitude and direction. Thus, effects of [CO₂] on pine leaf growth respiration may be due mainly to increased growth rate rather than changes in costs of construction.

We emphasize that, in addition to [CO₂], N influenced apparent leaf construction cost (Fig. 4) and we reiterate the suggestion of Griffin *et al.* (1993) that with elevated [CO₂] and low N, pine growth may be sink limited, resulting in an increased non-structural carbohydrate concentration, and that with elevated [CO₂] and high N, pine growth may be source limited. Trends in leaf weight:area ratio, although not statistically significant (Table 2) are consistent with this view and indicate a greater accumulation of non-structural carbohydrates at low compared to high N. Carbohydrate data also follow this pattern (Fig. 3) which was also reported by Thomas and Strain (1991).

In summary, we did not observe a significant effect of [CO₂] on leaf respiration rate per unit leaf area or mass, but respiration per unit leaf N was greater in elevated CO₂. The lack of a respiratory response per unit leaf mass or area is likely due to indirect effects that influenced respiration in opposing ways. Increased carbohydrate content resulted in increased transport and/or increased growth, thus increasing respiration rate, while lower N content decreased maintenance respiration rate. The increased respiration per unit foliar N may reflect more growth per unit N or greater carbohydrate transport at a given level of foliar N. Although elevated [CO₂] increased growth rate, the effect of increased growth rates on growth respiration were partially offset by decreased costs of constructing and maintaining a unit of leaf tissue. Nitrogen supply had a greater effect on both maintenance respiration and cost of constructing needle tissue than did atmospheric [CO₂].

Acknowledgements

This material is based upon work supported through the Southeastern Regional Center of the National Institute for Global Environmental Change by the US Department of Energy under Co-operative Agreement No. DE-FC03-90ER61010 and through the Experimental Program to Stimulate

Table 2. Effects of long-term (open top chamber) [CO₂] and nitrogen treatments on construction cost variables for longleaf pine needles

Nitrogen (kg ha ⁻¹ yr ⁻¹)	Long-term [CO ₂] (μmol CO ₂ mol ⁻¹)	<i>n</i>	% N ^a	% Ash	Δ Heat of combustion (kJ g ⁻¹)	Leaf weight: area ratio (g m ⁻²)
40	360	6	0.77 c	3.25 a	20.13 b	116.31 a
40	720	6	0.60 d	2.97 ab	19.97 b	122.31 a
400	360	6	1.28 a	3.32 a	20.74 a	121.99 a
400	720	6	0.97 b	2.61 b	20.13 b	113.08 a

^a Letters which differ within a column indicate a significant difference ($\alpha = 0.05$) according to contrasts conducted under General Linear Models.

Competitive Research by the US Environmental Protection Agency under Contract No. R821826-01-1. Any opinions, findings, and conclusions or recommendations expressed in this publication are those of the authors and do not necessarily reflect the views of the US Department of Energy or the US Environmental Protection Agency. The authors wish to express their sincere appreciation to Mr Jack Jarrell, Mr Barry Dorman, Ms Tammy Counts, and Ms Trina Cagle for technical assistance provided during the course of this project. We also appreciate critical reviews of the manuscript provided by Drs SD Wullschlegel, RB Thomas, and BG Drake.

References

- Amthor JS. 1989. *Respiration and crop productivity*. Berlin: Springer Verlag.
- Amthor JS. 1991. Respiration in a future, higher-CO₂ world. *Plant, Cell and Environment* **14**, 13–20.
- Amthor JS. 1994. Plant respiratory responses to the environment and their effects on the carbon balance. In: Wilkinson RE, ed. *Plant-environment interactions*. New York: Marcel Dekker, 501–54.
- Amthor JS. 1995. Plant respiratory responses to elevated CO₂ partial pressure. In: Allen LH, Kirkham MB, Olszyk DM, Whitman CE, eds. *Advances in CO₂ effects research*. ASA Special Publication, Madison, WI: ASA, CSSA, and SSSA (in press).
- Amthor JS, Koch GW, Bloom AJ. 1992. CO₂ inhibits respiration in leaves of *Rumex crispus* L. *Plant Physiology* **98**, 757–60.
- Amthor JS, Mitchell RJ, Runion GB, Rogers HH, Prior SA, Wood CW. 1994. Energy content, construction cost and phytomass accumulation of *Glycine max* (L.) Merr. and *Sorghum bicolor* (L.) Moench grown in elevated CO₂ in the field. *New Phytologist* **128**, 443–50.
- Azcón-Bieto J, Gonzalez-Meler MA, Doherty W, Drake BG. 1994. Acclimation of respiratory O₂ uptake in green tissues of field-grown native species after long-term exposure to elevated atmospheric CO₂. *Plant Physiology* **106**, 1163–8.
- Azcón-Bieto J, Lambers H, Day DA. 1983. Effect of photosynthesis and carbohydrate status and the involvement of the alternative pathway in leaf respiration. *Plant Physiology* **72**, 598–603.
- Azcón-Bieto J, Osmond CB. 1983. Relationship between photosynthesis and respiration. The effect of carbohydrate status on the rate of CO₂ production by respiration in darkened and illuminated wheat leaves. *Plant Physiology* **71**, 574–81.
- Baker JT, Laugel F, Boote KJ, Allen Jr LH. 1992. Effects of daytime carbon dioxide concentration on dark respiration in rice. *Plant, Cell and Environment* **15**, 231–9.
- Bazzaz FA. 1990. The response of natural ecosystems to the rising global CO₂ levels. *Annual Review of Ecology and Systematics* **21**, 167–96.
- Bazzaz FA, Miao SL. 1993. Successional status, seed size, and responses of tree seedlings to CO₂, light, and nutrients. *Ecology* **74**, 104–12.
- Begg JE, Jarvis PG. 1968. Photosynthesis in Townsville lucerne (*Stylosanthes humilis* H.B.K.). *Agricultural Meteorology* **5**, 91–109.
- Bowes G. 1993. Facing the inevitable: plants and increasing atmospheric CO₂. *Annual Review of Plant Physiology and Plant Molecular Biology* **44**, 309–32.
- Bunce JA. 1990. Short- and long-term inhibition of respiratory carbon dioxide efflux by elevated carbon dioxide. *Annals of Botany* **65**, 637–42.
- Bunce JA. 1992. Stomatal conductance, photosynthesis and respiration of temperate deciduous tree seedlings grown outdoors at an elevated concentration of carbon dioxide. *Plant, Cell and Environment* **15**, 541–9.
- Coleman JS, Rochefort L, Bazzaz FA, Woodward FI. 1991. Atmospheric CO₂, plant nitrogen status and the susceptibility of plants to an acute increase in temperature. *Plant, Cell and Environment* **14**, 667–74.
- Ceulemans R, Mousseau M. 1994. Effects of elevated atmospheric CO₂ on woody plants. *New Phytologist* **127**, 425–46.
- Downton WJS, Grant WJR. 1994. Photosynthetic and growth responses to variegated ornamental species to elevated CO₂. *Australian Journal of Plant Physiology* **21**, 273–97.
- Eamus D, Jarvis PG. 1989. The direct effects of increase in the global atmospheric CO₂ concentration on natural and commercial temperate trees and forests. *Advances in Ecological Research* **19**, 1–55.
- El Kohen A, Pontailier J-Y, Mousseau M. 1991. Effet d'un doublement du CO₂ atmosphérique sur la respiration à l'obscurité des parties aériennes de jeunes châtaigniers (*Castanea sativa* Mill.). *Comptes Rendus de l'Académie des Sciences, Paris* **312**, Série III, 477–81.
- Fajer ED, Bowers MD, Bazzaz FA. 1992. The effect of nutrients and enriched CO₂ environments on production of carbon-based allelochemicals in *Plantago*: a test of the carbon/nutrient balance hypothesis. *American Naturalist* **140**, 707–23.
- Farrar JF, Williams ML. 1991. The effects of increased atmospheric carbon dioxide and temperature on carbon partitioning, source-sink relations and respiration. *Plant, Cell and Environment* **14**, 819–30.
- Field CB, Chapin FS, Matson PA, Mooney HA. 1992. Responses of terrestrial ecosystems to the changing atmosphere: a resource-based approach. *Annual Review of Ecology and Systematics* **23**, 201–35.
- Gale J. 1982. Evidence for essential maintenance respiration of leaves of *Xanthium strumarium* at high temperature. *Journal of Experimental Botany* **33**, 471–6.
- Gifford RM, Lambers H, Morison JIL. 1985. Respiration of crop species under CO₂ enrichment. *Physiologia Plantarum* **63**, 351–6.
- Griffin KL, Thomas RB, Strain BR. 1993. Effects of nitrogen supply and elevated carbon dioxide on construction cost in leaves of *Pinus taeda* (L.) seedlings. *Oecologia* **95**, 575–80.
- Hickleton PR, Jolliffe PA. 1978. Effects of greenhouse CO₂ enrichment on the yield and photosynthetic physiology of tomato plants. *Canadian Journal of Plant Science* **58**, 801–17.
- Hilbert DW, Larigauderie A, Reynolds JF. 1991. The influence of carbon dioxide and daily photon-flux density on optimal leaf nitrogen concentration and root:shoot ratio. *Annals of Botany* **68**, 365–76.
- Hrubec TC, Robinson JM, Donaldson RP. 1985. Effects of CO₂ enrichment and carbohydrate content on the dark respiration of soybeans. *Plant Physiology* **79**, 684–9.
- Kendall AC, Turner JC, Thomas SM, Keys AJ. 1985. Effects of CO₂ enrichment at different irradiances on growth and yield of wheat. *Journal of Experimental Botany* **36**, 261–73.
- Larigauderie A, Hilbert DW, Oechel WC. 1988. Effect of CO₂ enrichment and nitrogen availability on resource acquisition and resource allocation in a grass, *Bromus mollis*. *Oecologia* **77**, 544–9.
- Mooney HA. 1991. Biological response to climate change: an agenda for research. *Ecological Applications* **1**, 112–17.
- Mousseau M. 1993. Effects of elevated CO₂ on growth, photosynthesis and respiration of sweet chestnut (*Castanea sativa* Mill.). *Vegetatio* **104/105**, 413–19.
- Nijs I, Impens I, Behaeghe T. 1988. Effects of elevated

- atmospheric carbon dioxide on gas exchange of white clover. *Photosynthesis Research* **15**, 163–76.
- Norby RJ, Gunderson CA, Wullschleger SD, O'Neill EG, McCracken MK.** 1992. Productivity and compensatory responses of yellow-poplar trees in elevated CO₂. *Nature* **357**, 322–4.
- Norby RJ, O'Neill EG, Hood WG, Luxmoore RJ.** 1986. Effects of atmospheric CO₂ enrichment on the growth and mineral nutrition of *Quercus alba* seedlings in a nutrient poor soil. *Plant Physiology* **82**, 83–9.
- Poorter H, Gifford RM, Kriedemann PE, Wong SC.** 1992. A quantitative analysis of dark respiration and carbon content as factors in the growth response of plants to elevated CO₂. *Australian Journal of Botany* **40**, 501–13.
- Raynaud D, Jouzel J, Barnola JM, Chappellaz J, Delmas RJ, Lorius C.** 1993. The ice record of greenhouse gases. *Science* **259**, 926–34.
- Reuveni J, Gale J.** 1985. The effects of high levels of carbon dioxide on dark respiration and growth of plants. *Plant, Cell and Environment* **8**, 623–8.
- Rogers HH, Heck WW, Heagle AS.** 1983. A field technique for the study of plant responses to elevated carbon dioxide concentrations. *Air Pollution Control Association Journal* **33**, 42–4.
- Rogers HH, Runion GB, Krupa SV.** 1994. Plant responses to atmospheric CO₂ enrichment with emphasis on roots and the rhizosphere. *Environmental Pollution* **83**, 155–89.
- Ryan MG.** 1991. Effects of climate change on plant respiration. *Ecological Applications* **1**, 157–67.
- Spencer W, Bowes G.** 1986. Photosynthesis and growth of water hyacinth under CO₂ enrichment. *Plant Physiology* **82**, 528–33.
- Strain BR.** 1987. Direct effects of increasing atmospheric CO₂ on plants and ecosystems. *Tree* **2**, 18–21.
- Sundquist ET.** 1993. The global carbon dioxide budget. *Science* **259**, 934–41.
- Thomas RB, Griffin KL.** 1994. Direct and indirect effects of atmospheric carbon dioxide enrichment on leaf respiration of *Glycine max* (L.) Merr. *Plant Physiology* **104**, 355–61.
- Thomas RB, Reid CD, Ybema R, Strain BR.** 1993. Growth and maintenance components of leaf respiration of cotton grown in elevated carbon dioxide partial pressure. *Plant, Cell and Environment* **16**, 539–46.
- Thomas RB, Strain BR.** 1991. Root restriction as a factor in photosynthetic acclimation of cotton seedlings grown in elevated carbon dioxide. *Plant Physiology* **96**, 627–34.
- Villar R, Held AA, Merino J.** 1994. Comparison of methods to estimate dark respiration in the light in leaves of two woody species. *Plant Physiology* **105**, 167–72.
- Williams K, Percival F, Merino J, Mooney HA.** 1987. Estimation of tissue construction cost from heat of combustion and organic nitrogen content. *Plant, Cell and Environment* **10**, 725–34.
- Williams ML, Jones DG, Baxter R, Farrar JF.** 1992. The effect of enhanced concentrations of atmospheric CO₂ on leaf respiration. In: Lambers H, van der Plas LHW, eds. *Molecular, biochemical and physiological aspects of plant respiration*. The Hague: SPB Academic Publishing, 547–51.
- Wullschleger SD, Ziska LH, Bunce JA.** 1994. Respiratory responses of higher plants to atmospheric CO₂ enrichment. *Physiologia Plantarum* **90**, 221–9.
- Ziska LH, Bunce JA.** 1993. Inhibition of whole plant respiration by elevated CO₂ as modified by growth temperature. *Physiologia Plantarum* **87**, 459–66.
- Ziska LH, Bunce JA.** 1994. Direct and indirect inhibition of single leaf respiration by elevated CO₂ concentrations: interaction with temperature. *Physiologia Plantarum* **90**, 130–8.
- Ziska LH, Seemann JR, DeJong TM.** 1990. Salinity-induced limitations in photosynthesis in *Prunus salicina*, a deciduous tree species. *Plant Physiology* **93**, 864–70.