

Utilizing cover crop mulches to reduce tillage in organic systems in the southeastern USA

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Research Paper

Abstract

Organic systems in the southeastern USA offer unique challenges and solutions to crop production due to regional soil and climate characterized by highly weathered soil types, high precipitation and the capacity to grow cover crops in the winter. Recently, the interest of producers and researchers in high-residue cover crops and conservation tillage systems has increased. Various designs of the roller crimper to manage cover crops have been invented and demonstrated to growers in the southeastern region of the USA over the past 17 years. The impacts of high-residue cover crop mulches on the agronomic systems in the region are diverse. Legume cover crops assist with meeting N demand from cash crops though they decompose rapidly and are seldom sufficient for N demanding crops such as corn. Cereal cover crop mulches can have the opposite effect by immobilizing N and have a longer impact on soil moisture and weed dynamics. While undesirable for many crops, N immobilization is one possible mechanism for weed suppression in legume cash crops planted into cereal residues. Other cover crop weed suppression mechanisms include physical impedance, light availability, allelopathy and microclimate effects. Regardless of the cause, successful weed control by mulches is highly dependent on having substantial biomass. The southeastern region is capable of producing cover crop biomass in excess of 9000 kg ha⁻¹, which is sufficient for weed control in many cash crops, although supplementary weed control is sometimes necessary. Long-term data are needed to predict when farmers should add supplementary weed control. More work is also needed on how much additional N is required for the cash crops and how best to deliver that N in a high-residue environment using organic sources.

Key words: roller crimper, organic, no till

Roller–Crimpers and Terminating Cover Crops

Rolled cover crops have been a primary research focus as a means to increase cover crop adoption and reduce tillage on organic farms in the southeastern USA. Rolling technology originated in Brazil where producers have been using rollers successfully for decades in no-till conservation systems¹. The Brazilian-type roller/crimper design (Fig. 1a) did not result in wide adoption in the USA in part due to vibration transferred to the tractor and operator as speed increased. To increase adoption, research has been conducted by the USDA-ARS National Soil Dynamics Laboratory since 1994 on new roller designs that are effective in terminating cover crop, but generate less vibration (Fig. 1b, c). Based on the success of

the novel roller designs, new concepts for rollers/crimpers intended for vegetable production were developed, such as an elevated bed roller to terminate cover crops on row-tops and in furrows (Fig. 2).

Both general principles and soil-type-dependent practices are needed for establishing cash crops in thick cover crop mulches. One general principle is the need to roll crimp cover crops parallel to the cash crop planting direction because it minimizes or eliminates residue buildup on subsoiling and planting units. Previous research has documented that diagonal and perpendicular rolling directions will cause residue buildup, increase cleaning time of planting units from residue and decrease planting quality, i.e., skips in planting, ‘hair pinning’ and slow emergence². In addition, row cleaners mounted immediately prior to or after the cutting coulter (Fig. 3) can further enhance direct seeding and transplanting operations.



Figure 1. Different roller types: (a) original Brazilian type straight bar roller, (b) curved bar roller crimper and (c) smooth roller with crimping bar (U.S. patent no. 7,604,067 B1).



Figure 2. (a) Roller crimper for elevated beds: one row and two furrows. (b) Roller for elevated beds: two rows and three furrows. (c) Two stage roller/crimper designed to operate with smaller tractors (40 HP power source). The roller comprises a 30cm diameter smooth drum which rolls down the cover crop and second small 15cm diameter drum with six equally spaced 0.6cm thick crimping bars on the drum's perimeter. Depending on cover crop type and amount, crimping force can be adjusted by changing preloading force of two adjusting springs to obtain the best crimping characteristics for a particular cover crop. Patent pending U.S. Patent no. 7,562,517 B1.



Figure 3. (a) John Deere no till planter for planting cotton seeds into rye residue cover using DAWN™ row cleaners and (b) John Deere no till planter with attached Yetter™ row cleaners.

Rollers have been shown to be beneficial by flattening the cover crop to provide a mat over the surface of the field and prevent multiple-direction lodging of cereal cover crops³. Effectively killing cover crops without herbicides depends on the growth stage and species involved. The best growth stage for terminating cereal rye is from early milk to soft dough⁴, late flowering for crimson clover⁵ (*Trifolium incarnatum* L.) and when pods are first seen in the upper five nodes of hairy vetch⁶ (*Vicia villosa* Roth). Rye is typically greater than 90% terminated after 3 weeks

if rolling is performed at the optimal stage^{4,7,8}. Cover crops should be terminated 2–3 weeks prior to planting to restore soil moisture depleted by the cover crop^{9–11}.

Weed Management Impacts of Reducing Tillage

For growers transitioning to and maintaining organic production, identifying effective weed control strategies

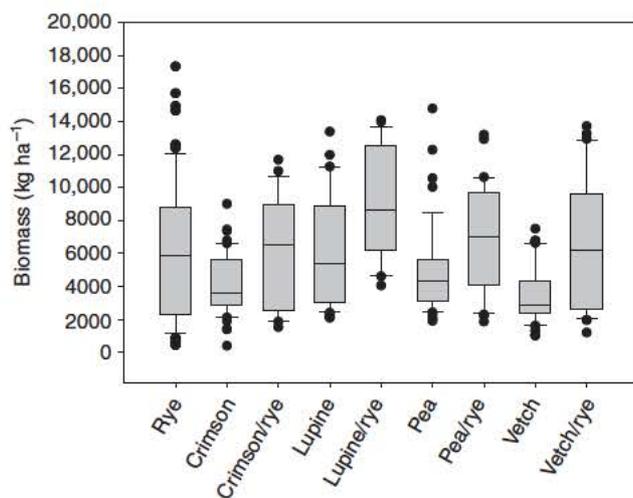


Figure 4. Productivity of legumes and legume/cereal mixes in the Southeast from 2008 to 2010. Figures come from a pooled analysis of trials in Georgia (GA) and North Carolina (NC). Box plots were created from nine site years for all cover crops except for lupine and lupine/rye, with four to six replicates per site year. Lupine data are from GA only (five site years) due to extremely low productivity in NC. Crimson clover varieties: *T. incarnatum* L. 'AU Sunrise' in NC; 'Dixie' in GA. Vetch varieties: *V. villosa* Roth 'AU Early Cover' in NC; *V. sativa* L. 'Cahaba White' in GA. Winter pea varieties: *P. sativum* L. subsp. *sativum* var. *arvense* (L.) Poir 'variety unstated' in NC and GA. Narrow leaf lupine varieties: *L. angustifolius* L. 'TifBlue 78' in NC and GA. NC data adapted from Parr et al.⁵. Rye (*Secale cereale* L.) at both locations was 'Wrens abruzzi' grown without added N fertility.

can be the most problematic issue faced¹²⁻¹⁵. As with conventional systems, adequate weed control is crucial for achieving optimal yields in organic systems. Traditionally, organic producers have primarily relied upon mechanical methods to maintain sufficient weed control due to the lack of synthetic herbicide options¹⁶. Whether through initial tillage, in-season cultivation, hand removal of weeds or flaming, mechanical means to inhibit weed growth are very effective tools for reducing weed pressure in organic systems^{16,17}. However, these practices degrade soil quality, require more labor, necessitate multiple field passes or may not fully control certain weed species. Several mechanical control tactics such as rotary hoeing or flex-tine harrowing can only destroy weeds at very early growth stages¹⁸. Extensive cultivation with implements such as the rotary hoe can also damage crops¹⁹. Weather conditions, such as prolonged periods of precipitation, can also complicate mechanical control by delaying cultivation and allowing weeds to reach a growth stage that precludes effective control. Moreover, continuous tillage practices can increase soil erosion through water runoff in an agricultural system. Organic growers need additional weed control methods that are less time sensitive, less labor intensive and that enhance soil conservation and soil quality.

Mechanisms of weed control by mulches

Cover crops can stress weeds at multiple points in their life cycle, including reducing seedling establishment, minimizing growth and competitive ability and reducing or preventing production of propagules²⁰. Emergence of small-seeded annual weeds is reduced with increasing rates of mulch residue²¹⁻²³. Some of the troublesome small-seeded broadleaf weeds common to the southeastern region include Palmer amaranth (*Amaranthus palmeri* S. Wats.), spiny amaranth (*Amaranthus spinosus* L.) and redroot pigweed^{24,25} (*Amaranthus retroflexus* L.). Physical impedance and light availability are important factors influencing weed establishment, especially with small-seeded weeds^{23,26,27}, though other factors likely contribute to weed suppression, including allelopathy^{22,28-30} and microclimate effects³¹. Nitrogen immobilization may also be playing a role in soybean/rye systems because low N conditions can favor growth of N-fixing crops over non-N fixing weeds³².

Cover crop productivity impacts on weed management

Hairy vetch produced 2340 kg ha⁻¹ of dry biomass in Mississippi, but had little effect on reducing densities of smooth pigweed³³ (*Amaranthus hybridus* L.). This level of biomass residue was likely insufficient to suppress weeds and likely degraded rapidly. In the northeastern USA, weed densities were reduced with cover crop residues between 6000 and 8500 kg ha⁻¹, double the amount of documented growth in that region²¹ (plots were supplemented with additional residues). Legume monocultures generally do not achieve that level of productivity in the southeastern USA³⁴ (Fig. 4). However, rye/legume mixes can produce significantly more biomass than legume monocultures (Fig. 4). Rye monocultures produce more biomass than other cover crops in Alabama³⁵, Georgia³⁶ and North Carolina³⁴. Narrowleaf lupin (*Lupinus angustifolius* L.) is the only other winter cover crop that can reach biomass levels similar to rye but its sensitivity to frost damage limits its usefulness to the most southern parts of the region (Fig. 4). Mixtures of rye with winter legumes have shown increased weed suppression compared to winter legumes alone in Arkansas and may extend weed suppression benefits, as legume residues decompose more rapidly than rye³⁷.

Traditionally, rye cover crops in the southeastern US are planted at relatively low seeding rates (60 kg ha⁻¹) in late autumn, with minimal inputs, and are mowed prior to planting the summer crop. To maximize the weed control provided by a rye mulch, a more intensive cover crop management system is needed. Planting early, with higher seeding rates and adequate N fertility,^{7,38} is needed to maximize the amount of rye biomass. Appropriate cultivar selection, long an under-studied component of cover crop systems, can also contribute to biomass

Table 1. Earliest roll times and productivity for rye (*Secale cereale* L.) cover crops in North Carolina. Rye was grown with 90 lb N applied to simulate maximal potential rye growth for each location.¹

Maturity	Estimated roll time ²	Cultivar	Biomass ³ (kg ha ⁻¹)
Early	Late April	Wrens Abruzzi	11,500 a ⁴
		Maton II	11,300 ab
		Wrens 96	10,800 ab
Mid	Early May	Aroostook	10,300 ab
Late	Mid to late May	Wheeler	10,200 b
		Rymin	8600 c

¹ Adapted from Wells *et al.*⁶²

² Estimate based on average time each cultivar reached Feeke's stage 11.1 (milk).

³ Biomass values are averaged over three site years in North Carolina.

⁴ Means followed by the same letter are not significantly different according to Fisher's Protected LSD.

production and affect the timing of the system (Table 1). While intensive management for a cover crop is uncommon, the potential economic value from improved weed control and higher crop yields would justify such investments. Relative to multiple spring tillage passes and repeated cultivations of the cash crop, more intensive cover crop management can be economically beneficial by comparison^{6,39}. With adequate fertility, rye biomass yields can exceed 9000 kg ha⁻¹; a level of biomass that affords excellent weed control^{21,40} (Fig. 5). However, substantial year-to-year variation in biomass production suggests that supplementary weed control will be necessary in some years.

Supplementary weed control

Supplemental weed control options are limited when utilizing high-residue cover crop mulches. Most cultivators will not work in such a high-residue environment. One type of cultivator that is reportedly successful on a variety of soil types is the wide sweep cultivator mounted on a parallel linkage^{40,41}. Most models consist of a large, smooth coulter mounted ahead of a wide sweep cultivator that has one or two depth wheels located near each sweep. The angle of the sweep relative to the soil surface is generally adjustable. Running the cultivator completely flat, or parallel to the soil surface, severs weed root systems minimally disturbing residue and soil. However, rerooting is possible with wet conditions, particularly with smaller weeds.

Herbicide options are limited in number and efficacy in organic systems^{42,43}. Available herbicides used in the southeastern USA largely contain either clove oil or citric acid and are non-selective in nature¹⁶. In addition to being costly, previous research has shown that these natural herbicides can be inefficient weed control methods⁴³.

Under-leaf band spraying of nonselective herbicides would fit well into mulch systems, but the high cost and only partial effectiveness make them economically infeasible for corn and soybeans, though they may have a role in higher-value crops.

Fertility impacts of rolled cover crops

In the Southeast, the ideal time for successful roll kill of cover crops often occurs late in the spring and is not ideal for timely corn planting^{6,44,45}. It is now well understood that rolled termination of cover crops is most effective after plant anthesis⁴. Susceptibility of legume cover crops to rolling has been shown to be significantly later than the traditional termination time for incorporated legumes, with rolling occurring in late-April for crimson clover varieties, mid-May for hairy vetch varieties and winter peas (*Pisum sativum* L. subsp. *sativum* var. *arvense* (L.)), and late-May for common vetch (*Vicia sativa* L.) and berseem clover (*Trifolium alexandrinum* L.)⁵. While late termination can delay corn planting, it can also result in significant increases in legume biomass N returned to the system due to having a longer period of time for spring growth and N accumulation⁴⁶⁻⁴⁸. Hairy vetch and crimson clover have been shown to average 4900 and 5500 kg ha⁻¹, respectively, in North Carolina⁵, resulting in 150 and 120 kg N ha⁻¹. A strong positive correlation has been found between the number of growing-degree days and vetch biomass production⁴⁶, meaning that year-to-year variation in vetch production can be significant depending on planting date and winter temperatures.

Nitrogen mineralization from legume cover crops

Nitrogen delivery in rolled cover crop legume systems can differ significantly from green manure systems in which the cover crop is incorporated into the soil⁴⁹. In particular, microbial function is affected by the change in plant quality that results from delayed termination, and by the change in decomposition kinetics due to the position of the rolled surface mulch in relation to soil decomposers.

The biologically driven, thus often delayed, termination time necessary in rolled systems is the first constraint that can potentially impact mineralization rate. As rolled cover crops are only successfully terminated at anthesis, plant quality and chemistry are expected to be distinctly different from earlier termination with herbicides. Decomposition and mineralization are microbial processes controlled in part by plant chemistry determinants. Such indicators include tissue C:N ratio, and cellulose, hemicelluloses, lignin and phenolic content. Ratios of C:N below 25 are assumed to release N soon after termination, but N mineralization slows with increasing hemicellulose, lignin and phenolic content, regardless of C:N ratio^{50,51}. Hairy vetch residue, having been found to have low

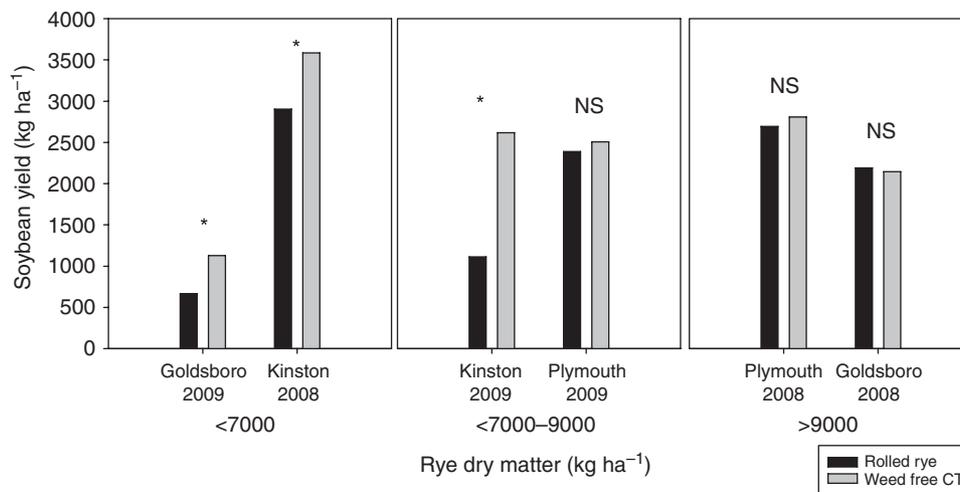


Figure 5. Soybean yield in roll crimped rye mulches. Treatments consisted of rolled rye mulch with no additional weed control measures (rolled rye) and a weed free, conventionally tilled check treated with S metolachlor pre emergence, imazethpyr post emergence and hand weeding as needed. Adapted from Smith et al.³⁹.

hemicellulose concentrations and a C:N ratio of around 11, is known to release N faster than other residues in chemically terminated systems⁵⁰. In rolled systems, C:N ratio has been shown to increase with later roll dates as plants age, with the lowest ratios in hairy vetch and winter pea varieties observed to range from 11 to 15:1 and crimson clover from 17 to 20:1⁵. Despite the relatively narrow gap in C:N ratios between crimson clover and hairy vetch, N release can be substantially slower from crimson clover⁵².

The second constraint in rolled systems that could impact mineralization is the placement of the cover crop residue in relation to the decomposing soil microorganisms. Cover crop termination in organic systems is mechanical, often involving a combination of methods that may include tilling, mowing, undercutting and rolling/crimping; each differently affecting the ability of soil microorganisms to decompose and release biomass N. In general, residue incorporation buries plant tissue, placing it in close contact with decomposing soil microbes, and increases rate of decomposition and N release, which may lead to a lack of crop-N synchrony as N is released faster than crops can use it⁵³. Mowing or chopping residues increases residue surface area available to soil decomposers, increasing decomposition and reducing the overall biomass bulk^{54,55}. Previous work on no-till cover crop residues relying on chemical desiccation to terminate crops has been shown to increase N mineralization compared to untreated residues⁵⁴. Rolled cropping systems in the Southeast in particular provide a unique environment for cover crop residue decomposition. While the increased temperatures and moisture of southern climates may be favorable and hasten decomposition^{56,57}, the amount of soil-to-residue contact in rolled systems can also affect decomposition⁵⁵, with position on the soil surface of rolled residue altering microbial access

to residue and slowing N mineralization. Further, while surface residues can conserve soil water and prevent evaporation from the soil surface prior to canopy closure⁵⁸, the exposed nature of surface mulches may slow decomposition as a result of reduced water content in the residue. Data on controls of N release in rolled cover crop legumes are scarce, and this is a topic that should be investigated further. Peak extractable soil N under rolled hairy vetch has been observed to occur at 4 weeks after termination, slower than is typical for incorporated residues in the region, but still earlier than peak N demand by corn⁵². Vetch/grass mixtures contain approximately the same amount of N as vetch monocultures, but mixtures can often release N more slowly⁵⁰. Among legume species, variation in N release rates can be agronomically important. When terminated at anthesis, crimson clover release rates are notably slower than other legumes⁵⁰ and have even resulted in apparent net immobilization in rolled systems⁵².

Future directions

Information on how high-residue systems are impacting on insect and disease management is mostly missing from southeast organic systems. One exception is a variety of studies in the Appalachian region on organic vegetable production with reduced tillage⁵⁹⁻⁶¹, but equivalent work is missing in field crops. Anecdotally, three-cornered alfalfa hopper damage has been observed to be higher on soybeans in rolled rye in multiple states (Reberg-Horton, personal communication). While weed management has been a central area of inquiry in multiple states, long-term data on how these systems shift weed species composition are also lacking. A complete abandonment of tillage in organics is likely impossible, but the question of how

much tillage can be removed from these systems without selecting for perennial weeds will require longer-term trials.

The next stage of organic research also needs to fine tune agronomic recommendations for growers. By all appearances, legume cover crop mulches should not be the sole source of N fertility for corn (Fig. 4). Methods and rates of supplementation need to be investigated. The combination of heavy residue and bulky organic amendments make these lines of work a nontrivial task. Another open question lies in how reliably cover crop mulches can provide enough residue to suppress weeds. Clearly, some winters are not conducive enough to cover crop growth for sufficient biomass. Farmers need tools to predict when biomass is insufficient so they can utilize off-field mulch amendments or a clean-till system instead. Ideally, models could assess cover crop biomass potential in the early spring when changing to a clean-till system is relatively easy. By late spring, even poor cover crop stands can be difficult to incorporate well enough to facilitate conventional cultivation.

References

- Derpsch, R., Roth, C.H., Sidiras, N., and Köpke, U. 1991. Controle da erosão no Paraná, Brazil: Sistemas de cobertura do solo, plantio directo e prepare conservacionista do solo. Deutsche Gesellschaft für Technische Zusammenarbeit (GTZ) GmbH, Eschborn, Germany.
- Kornecki, T.S., Raper, R.L., Arriaga, F.J., Schwab, E.B., and Bergtold, J.S. 2009. Impact of rye rolling direction and different no till row cleaners on cotton emergence and yield. *Transactions of the American Society of Agricultural and Biological Engineers* 52:383-391.
- Kornecki, T.S., Price, A.J., Raper, R.L., and Arriaga, F.J. 2009. New roller crimper concepts for mechanical termination of cover crops. *Renewable Agriculture and Food Systems* 24:165-173.
- Ashford, D.L. and Reeves, D.W. 2003. Use of a mechanical roller crimper as an alternative kill method for cover crop. *American Journal of Alternative Agriculture* 18:37-45.
- Parr, M., Grossman, J.M., Reberg Horton, S.C., Brinton, C., and Crozier, C. 2011. Nitrogen delivery from legume cover crops in no till organic corn production. *Agronomy Journal* 103:1578-1590.
- Mischler, R., Duiker, S.W., Curran, W.S., and Wilson, D. 2010. Hairy vetch management for no till organic corn production. *Agronomy Journal* 102:355-362.
- Kornecki, T.S., Price, A.J., and Raper, R.L. 2006. Performance of different roller designs in terminating rye cover crop and reducing vibration. *Applied Engineering in Agriculture* 22:633-641.
- Kornecki, T.S., Price, A.J., Raper, R.L., and Bergtold, J.S. 2009. Effectiveness of different herbicide applicators mounted on a roller/crimper for accelerated rye cover crop termination. *Applied Engineering in Agriculture* 25:819-826.
- Reeves, D.W. 2003. A Brazilian model for no tillage cotton production adapted to the southeastern USA. In *Proceedings of the Second World Congress on Conservation Agriculture Producing in Harmony with Nature*, Iguassu Falls, Paraná, Brazil. p. 372-374.
- Hargrove, W.L. and Frye, W.W. 1987. The need for legume cover crops in conservation tillage production. In J.F. Power (ed.). *The Role of Legumes in Conservation Tillage Systems*. Soil Conservation Society of America, Ankeny, IA. p. 1-5.
- Price, A.J., Arriaga, F.J., Raper, R.L., Balkcom, K.S., Kornecki, T.S., and Reeves, D.W. 2009. Comparison of mechanical and chemical winter cereal cover crop termination systems and cotton yield in conservation agriculture. *Cotton Science* 13:238-245.
- Archer, D.W., Jaradat, A.A., Johnson, J.M.F., Weyers, S.L., Gesch, R.W., Forcella, F., and Kludze, H.K. 2007. Crop productivity and economics during the transition to alternative cropping systems. *Agronomy Journal* 99:1538-1547.
- Reberg Horton, S.C., Place, G.T., Weisz, R., York, A., and Hamilton, M. 2011. Weed management. In M. Hamilton, J. Dunphy, and J. Van Duyn (eds). *North Carolina Organic Grain Production Guide*. North Carolina State University. p. 27-30. Available at Web site <http://www.cefs.ncsu.edu/PDFs/organicgrainfinal.pdf> (accessed September 30, 2011).
- Plattner, K., Fonsah, E.G., Escalante, C., Krewer, G., Scherm, H., Andersen, P.C., Liburd, O., and Tertuliano, M. 2008. Economics of organic blueberry establishment in Georgia. *Journal of Food Distribution Research* 39:111-115.
- Walz, E. 1999. Third Biennial National Organic Farmers' Survey. Organic Farming Research Foundation, Santa Cruz, CA. p. 19-47.
- Boyhan, G., Kelley, T., Langston, D., Sparks, S., Culpepper, S., and Fonsah, G. 2009. Commercial Organic Vegetable Production. Bulletin 1300, University of Georgia Cooperative Extension, Athens, GA.
- Singh, B.P., Granberry, D.M., Kelley, W.T., Boyhan, G., Sainju, U.M., Phatak, S.C., Sumner, P.E., Bader, M.J., Webster, T.M., Culpepper, A.S., Riley, D.G., Langston, D.B., and Fonsah, G. 2005. Sustainable vegetable production. In R. Dris (ed.). *Vegetables: Growing Environment and Mineral Nutrition*. WFL Publisher, Helsinki, Finland, p. 1-38.
- Johnson, W.C. III and Mullinix, B.G. Jr. 2008. Potential weed management systems for organic peanut production. *Peanut Science* 35:67-72.
- Place, G.T., Reberg Horton, S.C., and Burton, M.G. 2009. Effects of preplant and postplant rotary hoe use on weed control, soybean pod position, and soybean yield. *Weed Science* 57:290-295.
- Sarrantonio, M. and Gallandt, E. 2003. The role of cover crops in North American cropping systems. *Journal of Crop Production* 8:53-74.
- Mohler, C.L. and Teasdale, J.R. 1993. Response of weed emergence to rate of *Vicia villosa* Roth and *Secale cereale* L. residue. *Weed Research* 33:487-499.
- Putnam, A.R. and Defrank, J. 1983. Use of phytotoxic plant residues for selective weed control. *Crop Protection* 2:173-181.
- Teasdale, J.R. and Mohler, C.L. 2000. The quantitative relationship between weed emergence and the physical properties of mulches. *Weed Science* 48:385-392.
- Webster, T.M. 2008. Weed survey southern states: Grass crop subsection. In W.K. Vencill (ed.). *Proceedings*

- of the Southern Weed Science Society, Jacksonville, FL. p. 224-243.
- 25 Webster, T.M. 2009. Weed survey southern states: Broadleaf crops subsection. In T.M. Webster (ed.). Proceedings of the Southern Weed Science Society, Orlando, FL, p. 509-524.
 - 26 Gallagher, R.S. and Cardina, J. 1998. Phytochrome mediated *Amaranthus* germination I: Effect of seed burial and germination temperature. *Weed Science* 46:48-52.
 - 27 Gallagher, R.S. and Cardina, J. 1998. Phytochrome mediated *Amaranthus* germination II: Development of very low fluence sensitivity. *Weed Science* 46:53-58.
 - 28 Burgos, N.R. and Talbert, R.E. 2000. Differential activity of allelochemicals from *Secale cereale* in seedling bioassays. *Weed Science* 48:302-310.
 - 29 Kruidhof, H.M., Bastiaans, L., and Kropff, M.J. 2009. Cover crop residue management for optimizing weed control. *Plant and Soil* 318:169-184.
 - 30 Reberg Horton, S.C., Burton, J.D., Danehower, D.A., Ma, G., Monks, D.W., Murphy, J.P., Ranells, N.N., Williamson, J.D., and Creamer, N.G. 2005. Changes over time in the allelochemical content of ten cultivars of rye (*Secale cereale* L.). *Journal of Chemical Ecology* 31:179-193.
 - 31 Teasdale, J.R. and Mohler, C.L. 1993. Light transmittance, soil temperature, and soil moisture under residue of hairy vetch and rye. *Agronomy Journal* 85:673-680.
 - 32 Wells, M.S., Reberg Horton, S.C., Smith, A.N., and Grossman, J.M. 2011. Effects of rye cover crop mulches on nitrogen dynamics in soybean. Chapter 1 in Wells, M.S., M.S. Thesis, North Carolina State University.
 - 33 Koger, C.H. and Reddy, K.N. 2005. Effects of hairy vetch (*Vicia villosa*) cover crop and banded herbicides on weeds, grain yield, and economic returns in corn (*Zea mays*). *Journal of Sustainable Agriculture* 26:107-124.
 - 34 Yenish, J.P., Worsham, A.D., and York, A.C. 1996. Cover crops for herbicide replacement in no tillage corn (*Zea mays*). *Weed Technology* 10:815-821.
 - 35 Reeves, D.W., Price, A.J., and Patterson, M.G. 2005. Evaluation of three winter cereals for weed control in conservation tillage nontransgenic cotton. *Weed Technology* 19:731-736.
 - 36 Schomberg, H.H., McDaniel, R.G., Mallard, E., Endale, D.M., Fisher, D.S., and Cabrera, M.L. 2006. Conservation tillage and cover crop influences on cotton production on a southeastern US coastal plain soil. *Agronomy Journal* 98:1247-1256.
 - 37 Burgos, N.R. and Talbert, R.E. 1996. Weed control and sweet corn (*Zea mays* var. *rugosa*) response in a no till system with cover crops. *Weed Science* 44:355-361.
 - 38 Mirsky, S., Curran, W., Mortensen, D., Ryan, M., and Shumway, D. 2011. Timing of cover crop management effects on weed suppression in no till planted soybean using a roller crimper. *Weed Science* 59:380-389.
 - 39 Smith, R.G., Barbercheck, M.E., Mortensen, D.A., Hyde, J., and Hulting, A.G. 2011. Yield and net returns during the transition to organic feed grain production. *Agronomy Journal* 103:51-59.
 - 40 Smith, A.N., Reberg Horton, S.C., Place, G.T., Meijer, A.D., Arellano, C., and Mueller, J.P. 2011. Rolled rye mulch for weed suppression in organic no tillage soybeans. *Weed Science* 59:224-231.
 - 41 Price, A.J., Patterson, M.G., Monks, C.D., and Kelton, J.A. 2011. Evaluation of a high residue cultivator for Palmer Amaranth control in conservation tillage system. Southern Weed Science Society Annual Meeting, San Juan, Puerto Rico.
 - 42 Evans, G.J. and Bellinder, R.R. 2009. The potential use of vinegar and a clove oil herbicide for weed control in sweet corn, potato, and onion. *Weed Technology* 23:120-128.
 - 43 Johnson, W.C. III, Mullinix, B.G. Jr, and Boudreau, M.A. 2008. Peanut response to naturally derived herbicides in organic crop production. *Peanut Science* 35:73-75.
 - 44 Teasdale, J.R., Devine, T.E., Mosjidis, J.A., Bellinder, R.R., and Beste, C.E. 2004. Growth and development of hairy vetch cultivars in the northeastern United States as influenced by planting and harvesting date. *Agronomy Journal* 96:1266.
 - 45 Heiniger, R.W., Spears, J.F., Bowman, D.T., and Dunphy, E.J. 2000. Crop management. In North Carolina Corn Production Guide. The North Carolina Cooperative Extension Service, College of Agriculture and Life Sciences, North Carolina State University, Raleigh, NC.
 - 46 Cooke, J., Gallagher, R.S., Kaye, J.P., Lynch, J., and Bradley, B. 2010. Optimizing vetch nitrogen production and corn nitrogen accumulation under no till management. *Agronomy Journal* 102:1491-1499.
 - 47 Clark, A.J., Decker, A.M., and Meisinger, J.J. 1994. Seeding rate and kill date effects on hairy vetch cereal rye cover crop mixtures for corn production. *Agronomy Journal* 86:1065-1070.
 - 48 Wagger, M.G. 1989. Cover crop management and nitrogen rate in relation to growth and yield of no till corn. *Agronomy Journal* 81:533-538.
 - 49 Hartwig, N.L. and Ammon, H.U. 2002. Cover crops and living mulches. *Weed Science* 50:688-699.
 - 50 Ranells, N.N. and Wagger, M.G. 1996. Nitrogen release from grass and legume cover crop monocultures and bicultures. *Agronomy Journal* 88:777-782.
 - 51 Palm, C.A. and Sanchez, P.A. 1990. Decomposition and nutrient release patterns of the leaves of three tropical legumes. *Biotropica* 22:330-338.
 - 52 Parr, M., Reberg Horton, C., Crozier, C., and Grossman, J.M. 2011. Decomposition and nitrogen release from roll killed legume cover crops in an organic corn system. Chapter 3 in Parr, M., M.S. Thesis, North Carolina State University.
 - 53 Gaskell, M. and Smith, R. 2007. Nitrogen sources for organic vegetable crops. *HortTechnology* 17:431-441.
 - 54 Snapp, S.S. and Borden, H. 2005. Enhanced nitrogen mineralization in mowed or glyphosate treated cover crops compared to direct incorporation. *Plant and Soil* 270:101-112.
 - 55 Swift, M.J., Heal, O.W. and Anderson, J.M. 1979. Decomposition in Terrestrial Ecosystems. University of California Press, Berkeley, CA. p. 118-163.
 - 56 Schomberg, H.H. and Steiner, J.L. 1997. Estimating crop residue decomposition coefficients using substrate induced respiration. *Soil Biology and Biochemistry* 29:1089-1097.
 - 57 Schomberg, H.H., Foster, G.R., Steiner, J.L., and Stott, D.E. 2002. An improved temperature function for modeling crop residue decomposition. *Transactions of the American Society of Agricultural and Biological Engineers* 45:1415-1422.

- 58 Baldwin, K. and Creamer, N. 2006. Cover Crops for Organic Farms. North Carolina Cooperative Extension Service, Raleigh, NC.
- 59 Overstreet, L.F., Hoyt, G.D., and Imbriani, J. 2010. Comparing nematode and earthworm communities under combinations of conventional and conservation vegetable production practices. *Soil and Tillage Research* 110:42-50.
- 60 Hummel, R.L., Walgenbach, J.F., and Hoyt, G.D. 2002. Effects of production system on vegetable arthropods and their natural enemies. *Agriculture Ecosystems and Environment* 93:165-176.
- 61 Hoyt, G.D. and Walgenbach, J.F. 1995. Pest evaluation in sustainable cabbage production systems. *Hortscience* 30:1046-1048.
- 62 Wells, M.S., Reberg-Horton, S.C., and Brinton, C.M. 2011. An evaluation of six rye cultivars and their effects on weed suppression and soybean yield in the roller/crimper system. Chapter 2 in Wells, M.S., M.S. Thesis, North Carolina State University.