

Elevated CO₂ and Warming Effects on Soil Carbon Sequestration and Greenhouse Gas Exchange in Agroecosystems: A Review

Feike A. Dijkstra¹, Jack A. Morgan²

¹Faculty of Agriculture, Food and Natural Resources, the University of Sydney, Eveleigh NSW, Australia

²USDA Agricultural Research Service, Rangeland Resources Research Unit, Fort Collins, CO

467

CHAPTER OUTLINE

Methods 470

The Effect of eCO₂ on Soil C 472

The Effect of eCO₂ on Soil Respiration 473

The Effect of eCO₂ on N₂O Emission 475

The Effect of eCO₂ on CH₄ Exchange 477

Warming Effects on Soil C, Soil Respiration, N₂O Emission and CH₄ Exchange 479

Interactive eCO₂ × Warming Effects on Soil C, Soil Respiration, N₂O Emission, and CH₄ Exchange 480

Conclusions 482

Abbreviations: C, carbon; CH₄, methane; CO₂, carbon dioxide; eCO₂, elevated atmospheric CO₂ concentration; FACE, free air CO₂ enrichment; GHG, greenhouse gas; HC, heating cable; IPCC, Intergovernmental Panel on Climate Change; IRH, infrared heater; MAP, mean annual precipitation; MAT, mean annual temperature; N, nitrogen; N₂O, nitrous oxide; OTC, open top chamber; PT, plastic tunnel; SACC, screen-aided CO₂ control

Concentrations of CO₂ and other greenhouse gases (GHGs) have been increasing dramatically in earth's atmosphere since the industrial revolution, and are expected to continue increasing from ~385 ppmv today to more than 600 ppmv by the end of this century (IPCC, 2007).

Global surface temperatures are expected to rise between 1.1 to 5.4°C by 2100, depending on how fast greenhouse gas concentrations increase. Precipitation dynamics are also predicted to

TABLE 27.1 Summary of Studies Reporting Elevated CO₂ Effects On Soil C, Soil Respiration, N₂O Emission, and CH₄ Exchange

Agroecosystem	Site	Facility ¹	Other treatment	Reference Soil C	Respiration	N ₂ O	CH ₄
N Fertilized Studies							
Sorghum (Sorghum AL)	Auburn AL, U.S.	OTC		Prior et al. 2004			
Sorghum (Sorghum AZ)	Maricopa AZ, U.S.	FACE	Low and high irrigation	Cheng et al. 2007		Weizmliller et al. 2008	
Cotton (Cotton AZ)	Maricopa AZ, U.S.	FACE	Low and high irrigation	Wood et al. 1994	Nakayama et al. 1994		
Wheat (Wheat AZ)	Maricopa AZ, U.S.	FACE		Leavitt et al. 1996, Prior et al. 1997			
Wheat (Wheat AZ)	Maricopa AZ, U.S.	FACE	Low and high N fertilization	Leavitt et al. 2001	Pendall et al. 2001		
Wheat (Wheat GE)	Hohenheim, Germany	FACE		Marhan et al. 2010			
Wheat (Wheat CH)	Beijing, China	FACE			Lam et al. 2011	Lam et al. 2011	Lam et al. 2011
Wheat (Wheat SE)	Ostad, Sweden	OTC					
Soybean (Soybean AL)	Auburn AL, U.S.	OTC					
Rice (Rice JA)	Iwate, Japan	FACE	Unwarmed and warmed				Inubushi et al. 2003, Tokida et al. 2010
Rice (Rice FL)	Gainesville FL, U.S.	PT	Unwarmed and warmed				Schrope et al. 1999
Rice (Rice PH)	Los Baños, Philippines	OTC	Unwarmed and warmed, dry and wet season				Ziska et al. 1998
Wheat/sugar beet (Wheat/sugar GE)	Braunschweig, Germany	FACE		Gieseemann and Weigel 2008			
Wheat/soybean (Wheat/soy MD)	Beltsville MD, U.S.	OTC	Low and high irrigation, low and high ozone	Islam et al. 2000			
Corn/soybean (Corn/soy IL)	Urbana IL, U.S.	FACE		Peralta and Wander 2008			
Sorghum/soybean (Sorghum/soy AL)	Auburn AL, U.S.	OTC	Conventional and Conservation tillage	Prior et al. 2005	Smith et al. 2010	Smith et al. 2010	Smith et al. 2010

Rice/wheat (Rice/wheat JI)	Jiangdu, Jiangsu, China	FACE	Low and high N fertilization	Zhong et al. 2009	Kou et al. 2007	Xu et al. 2004, Zheng et al. 2006	
Rice/wheat (Rice/wheat WU)	Wuxi, Jiangsu, China	FACE	Low and high N fertilization	Juan et al. 2007			
White clover (White clover SW)	Eschikon, Switzerland	FACE	Low and high N fertilization	Van Kessel et al. 2006, 2000	Baggs et al. 2003	Baggs et al. 2003	
White clover bin SW)	Eschikon, Switzerland	FACE	Low and high N fertilization	Van Kessel et al. 2006, 2000	Ineson et al. 1998, Baggs et al. 2003	Ineson et al. 1998, Baggs et al. 2003	
Rye grass SW	Eschikon, Switzerland	FACE	Low and high N fertilization	Van Kessel et al. 2006	Baggs et al. 2003	Baggs et al. 2003	
Rye grass bin SW)	Eschikon, Switzerland	FACE	Low and high N fertilization	Van Kessel et al. 2006	Baggs et al. 2003	Baggs et al. 2003	
White clover/rye grass (White clover/rye SW)	France	PT	Low and high N fertilization	Jäger et al. 2003	Casella et al. 1997	Kammann et al. 2008	
Rye grass FR)	Giessen, Germany	FACE	Low and high N fertilization				
Temperate grassland (Temperate grass GE)	Germany	OTC		Niklaus and Körner 1996			
Alpine grassland (Alpine grass SW)	Switzerland	OTC					
Non-N Fertilized Studies							
Alpine grassland (Alpine grass SW)	Switzerland	OTC		Niklaus and Körner 1996			
Temperate grassland (Temperate grass SW)	Switzerland	SACC		Niklaus et al. 2001			
Temperate grassland (Temperate grass NZ)	Manawatu, New Zealand	FACE		Ross et al. 2004			
Temperate grassland (Temperate grass AU)	Tasmania, Australia	FACE	Unwarmed and warmed; C ₃ and C ₄ grasses	Pendall et al. 2011			
Annual grassland (Annual grass CA)	Stanford CA, U.S.	OTC	Two soil types	Hungate et al. 1997a	Luo et al. 1996, Hu et al. 2001	Hungate et al. 1997b	
Tall grass prairie (Tall grass prairie KS)	Manhattan KS, U.S.	OTC		Williams et al. 2000			
Short grass steppe (Short grass steppe CO)	Nunn CO, U.S.	OTC		Pendall et al. 2004	Mosier et al. 2002	Mosier et al. 2002	
Northern mixed grassland (Northern mixed grass WY)	Cheyenne WY, U.S.	FACE	Unwarmed and warmed	Unpublished results	Unpublished results	Dijkstra et al. 2011 Unpublished results	

¹FACE: Free Air CO₂ Enrichment; OTC: Open Top Chamber; PT: Plastic Tunnel; SACC: Screen-Aided CO₂ Control.

change, although there is still considerable uncertainty in these projections. While some of the details of these events are unclear, most agree climate change has already affected agroecosystems worldwide, and will have even more profound effects as climate change accelerates (Solomon et al., 2009). Important feedback exists between the atmosphere and the soil (Heimann and Reichstein, 2008), and a clear understanding of how climate change and rising atmospheric CO₂ might affect soil C sequestration and greenhouse gas exchange in agroecosystems is urgently needed.

Our review will address the effects of warming and rising CO₂ on the GHG balance. Although precipitation can have strong effects on C sequestration and greenhouse gas exchange in agroecosystems, current projections about precipitation responses remain highly uncertain. Our review will focus on manipulative field experiments in which researchers alter the environment to evaluate ecosystem responses. These experiments include manipulations of atmospheric CO₂ through Open Top Chambers (OTC), Free Air Carbon dioxide Enrichment (FACE), or Screen-Aided CO₂ Control (SACC), manipulations of temperature using heating cables (HC) or infrared heaters (IRH), or a combination of atmospheric CO₂ and temperature. We assess important mechanisms and identify critical knowledge gaps regarding the effects of elevated CO₂ (eCO₂) and warming on C sequestration and greenhouse gas exchange in agroecosystems.

METHODS

In our review we focused on manipulative field experiments, while we excluded growth chamber and greenhouse studies, studies conducted in arctic and subarctic environments, and studies conducted in systems with no direct agronomic benefit (e.g. forests). In most experiments, CO₂ concentrations were manipulated above present-day ambient concentrations (~375–385 ppmv) to enriched levels (470–720 ppmv). Temperature increases ranged between 1 and 5°C above ambient, consistent with IPCC projections for the end of the 21st century (IPCC, 2007). We reviewed a total of 32 eCO₂ and 13 warming studies (Tables 27.1 and 27.2).

The eCO₂ and warming effects on soil C, soil respiration, and N₂O emission were separated in N fertilized and non-N fertilized studies with the expectation that eCO₂ and warming effects on these properties largely depend on soil N availability. For instance, in other meta-analyses a significant increase in soil C under eCO₂ required N fertilization (Van Groenigen et al., 2006; Hungate et al., 2009). We further separated eCO₂ and warming effects on CH₄ exchange conducted in dry land sites (non-rice) where the net CH₄ efflux is predominantly negative (i.e. net CH₄ uptake in soils), and in rice paddy field studies where the net CH₄ efflux is much larger and always positive (i.e. net CH₄ production in soils). When other treatments were included, eCO₂ and warming effects were averaged across those other treatments (e.g. irrigation, ozone).

We calculated the effect of eCO₂ and warming on soil C as the absolute change in soil C (in g C kg⁻¹ soil) divided by the number of years of treatment. We used absolute changes rather than relative changes because absolute changes provide more biogeochemical significance (Hungate et al., 2009). The absolute changes were calculated for the shallowest soil depths reported, which ranged between 0–5 and 0–26 cm among studies. The number of years of treatment effects on soil C ranged between 2 and 10 years. We calculated the effect of CO₂ and warming on soil respiration, N₂O, and CH₄ flux rates as the absolute change in the average flux rates measured during the growing season (in kg C ha⁻¹ d⁻¹, g N ha⁻¹ d⁻¹, and g C ha⁻¹ d⁻¹ for CO₂, N₂O, and CH₄, respectively). When flux rates were measured in multiple years, we averaged the flux rates across years. All flux rates were measured using static chambers.

Because the effect of eCO₂ on soil C, soil respiration, N₂O, and CH₄ flux rates were highly variable among studies, we tested whether this variability could be explained by climate factors or soil properties of the study site. For the climate factors we chose mean annual temperature

TABLE 27.2 Summary of Studies Reporting Warming Effects On Soil C, Soil Respiration, N₂O Emission, and CH₄ Exchange

Agroecosystem	Site	Other Facility ¹ treatment	Reference Soil C	Respiration	N ₂ O	CH ₄
N Fertilized Studies						
Wheat (Wheat GE)	München, Germany	HC			Kamp et al. 1998	
Wheat (Wheat UK)	York, U.K.	HC		Hartley et al. 2007		
Corn (Corn UK)	York, U.K.	HC		Hartley et al. 2007		
Rice (Rice JA)	Iwate, Japan	HC				Tokida et al. 2010
Rice (Rice FL)	Gainesville FL, U.S.	TGC				Schrope et al. 1999
Rice (Rice PH)	Los Baños, Philippines	OTC				Ziska et al. 1998
Rye grass (Rye grass FR)	France	PT		Casella et al. 1997		
Non-N Fertilized Studies						
Alpine grassland (Alpine grass TI)	Tibet, China	IRH				Hu et al. 2010
Temperate grassland (Temperate grass CH)	Inner Mongolia, China	IRH		Liu et al. 2009, Xia et al. 2009		
Temperate grassland (Temperate grass AU)	Scotland, U.K.	HC		Briones et al. 2009		
Tall grass prairie (Tall grass OK)	Tasmania, Australia	IRH				
Northern mixed grassland (Northern mixed grass WY)	OK, U.S.	IRH				
	Cheyenne WY, U.S.	IRH		Unpublished results	Unpublished results	Unpublished results

¹HC: Heating Cable; IRH: Infrared Heater; PT: Plastic Tunnel.

(MAT) and mean annual precipitation (MAP) of the site where the studies were conducted and for soil properties we chose %clay and pH. We chose these climate and soil factors because they can have significant effects on plant growth and biological activity in the soil (Epstein et al., 1997; Guo et al., 2006; Fierer et al., 2009) and therefore we expected that they could significantly influence $e\text{CO}_2$ and warming effects on soil C and GHG flux rates among sites. These parameters are also frequently reported in the literature. We used %clay when reported in the study, but often only the textural class was reported. In that case we used the average %clay of the two boundaries of the textural class according to the textural triangle. For instance, if it was reported that the study was conducted in a sandy clay loam with a clay content between 20 and 35% according to the textural triangle, we designated that soil with the average clay content of 27.5%. We related CO_2 effects on soil C and GHG flux rates to each of MAT, MAP, %clay, and pH using linear regression. With the linear regressions, we put more weight on studies that were conducted over a longer time period, because we assumed that studies over longer time periods provide more reliable data. We weighted the absolute rate of change in soil C by the treatment length (in years) after which soil C was measured and weighted the absolute change GHG flux rates by the duration of the measurements (in years; Wu et al., 2011). Some studies were conducted at the same location and soil type, but in different years (e.g. the wheat, sorghum, and cotton studies at Maricopa, AZ). In those cases $e\text{CO}_2$ effects and treatment length/duration of measurements were averaged across the different studies conducted at the same site. We only constructed relationships when there were data available for four or more sites. All linear regressions were performed with JMP (version 8.0.1; SAS Institute, Cary, NC, USA).

THE EFFECT OF $e\text{CO}_2$ ON SOIL C

We found 27 studies (19 N fertilized and 8 non-N fertilized studies) where the effect of $e\text{CO}_2$ on soil C was reported (Figure 27.1). In 74% of the studies (79% of the N fertilized and 63% of

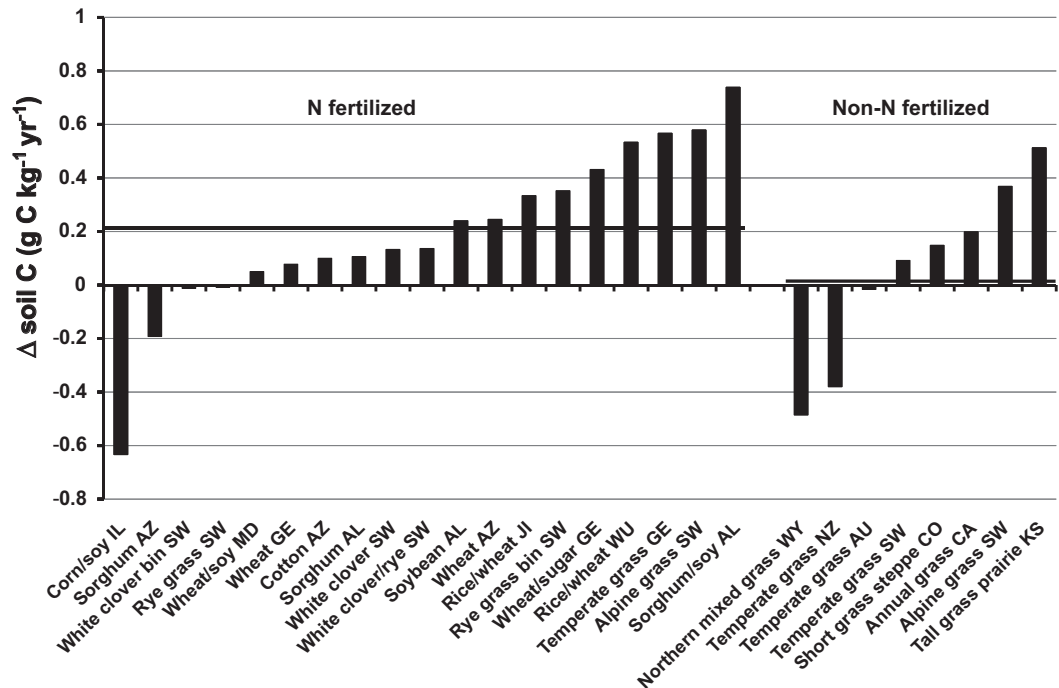


FIGURE 27.1

The rate of change in soil C in response to $e\text{CO}_2$ among different studies. Horizontal bold lines represent averaged values for N fertilized and non-N fertilized studies.

the non-N fertilized studies) a positive effect of eCO₂ on soil C was found, although only in a few occasions were these positive effects statistically significant (e.g. Williams et al., 2000; Prior et al., 2004, 2005; Zhong et al., 2009). While the positive effects of CO₂ enrichment on plant production are generally observed in the initial treatment year (Kimball et al., 2002), detection of significant changes in soil C may take many years of CO₂ enrichment due to the high amount of C present in soils and the relatively small amounts that accrue on an annual basis (Conant and Paustian, 2002; Smith, 2004). On average soil C increased by 0.205 g kg⁻¹ yr⁻¹ in the N fertilized studies and by 0.008 g kg⁻¹ yr⁻¹ in the non-N fertilized studies. If we assume that in all studies the soil had a bulk density of 1.3 g cm⁻³ and that the change in soil C occurred in the top 20 cm, this would correspond to an average rate of soil C increase of 1460 and 57 g ha⁻¹ d⁻¹ in the N fertilized and non-N fertilized studies, respectively. A greater response in N fertilized studies was also found by Van Groenigen et al. (2006) in their meta-analysis, where they included greenhouse and growth chamber studies, and non-agronomic sites. Our results confirm the notion that soil C sequestration under eCO₂ is generally constrained by the availability of N and that N fertilization enhances the capacity to increase soil C under eCO₂ (Reich et al., 2006a; Van Groenigen et al., 2006).

Nitrogen fixation by legumes is often enhanced under eCO₂, especially with the addition of non-N nutrients (van Groenigen et al., 2006), and legume responses to CO₂ tend to be greater than non-legumes under conditions of low soil N (van Kessel et al., 2006). Yet no strong evidence was found of greater C sequestration under eCO₂ in studies with legumes. Soil C only slightly increased in plots of *Trifolium repens* (white clover) in the FACE experiment at Eschikon, Switzerland, the most extensive evaluation yet of legume CO₂ responses in a field setting (van Kessel et al., 2006), while soil C decreased under eCO₂ in a temperate pasture with legumes in New Zealand (Ross et al., 2004).

Although on average soil C sequestration in response to eCO₂ was higher in N fertilized than in non-N fertilized studies, within the N fertilized and non-N fertilized studies eCO₂ effects on soil C showed large variation (Figure 27.1). For example, the most negative response to eCO₂ was observed in an N fertilized study with a corn–soybean rotation in Illinois (Peralta and Wander, 2008) and the most positive response was observed in an N fertilized study with a sorghum–soybean rotation in Alabama (Prior et al., 2005). Both these extreme responses were larger than any of the responses observed in the non-N fertilized studies. We tested whether this variability in soil C response among sites could be explained by site differences in climate and soil type. When we related the absolute rate of change in soil C in response to eCO₂ to climate and soil parameters, only %clay in the fertilized sites exhibited a relationship, with marginal significance ($P = 0.08$, Table 27.3). Soil C sequestration in response to eCO₂ tended to decrease with increased clay content (Figure 27.2). Although only marginally significant, this result is remarkable (and any marginal or significant relationships discussed further on) given that all these studies were done using different methods under a variety of conditions. A possible explanation for the decrease in soil C sequestration with increased clay content in response to eCO₂ is that rhizosphere priming effects on soil organic matter decomposition under eCO₂ may be stronger in more clayey soils. Rhizosphere priming, where microbial decomposition of relative recalcitrant soil organic matter is enhanced because of microbial stimulation by energy-rich root exudates, may increase under eCO₂ (Cheng, 1999), particularly in soils with greater clay content (Dijkstra and Cheng, 2007). Thus, an eCO₂-induced increase in rhizosphere priming in more clayey soils may result in less C sequestration, or even cause a net loss of soil C as was observed in the silty clay loam in Illinois, U.S. (Peralta and Wander, 2008).

THE EFFECT OF eCO₂ ON SOIL RESPIRATION

Soil respiration increased with eCO₂ in 12 of the 13 studies reviewed (Figure 27.3). A decrease in soil respiration under eCO₂ was observed in a study with rye grass in Switzerland (Ineson

TABLE 27.3 Summary of Regression Analyses Explaining Variation in Soil C, Soil Respiration, N₂O Emission and CH₄ Exchange in Response to eCO₂

A. Soil C									
	All sites			Fertilized sites			Non-fertilized sites		
	<i>n</i> *	<i>Corr. C.</i>	<i>P</i>	<i>n</i>	<i>Corr. C.</i>	<i>P</i>	<i>n</i>	<i>Corr. C.</i>	<i>P</i>
MAT	19	0.26	0.26	11	0.22	0.51	8	0.30	0.46
MAP	20	0.28	0.23	11	0.12	0.73	9	0.36	0.34
pH	20	-0.21	0.38	11	-0.20	0.56	9	-0.18	0.65
% Clay	22	-0.19	0.39	13	-0.51	0.08	9	0.11	0.79

B. Soil Respiration						
	All sites			Fertilized sites		
	<i>n</i>	<i>Corr. C.</i>	<i>P</i>	<i>n</i>	<i>Corr. C.</i>	<i>P</i>
MAT	9	0.56	0.13	6	0.29	0.60
MAP	8	0.46	0.22	5	0.15	0.80
pH	8	-0.71	0.06	5	-0.82	0.06
% Clay	10	0.71	0.03	7	0.50	0.33

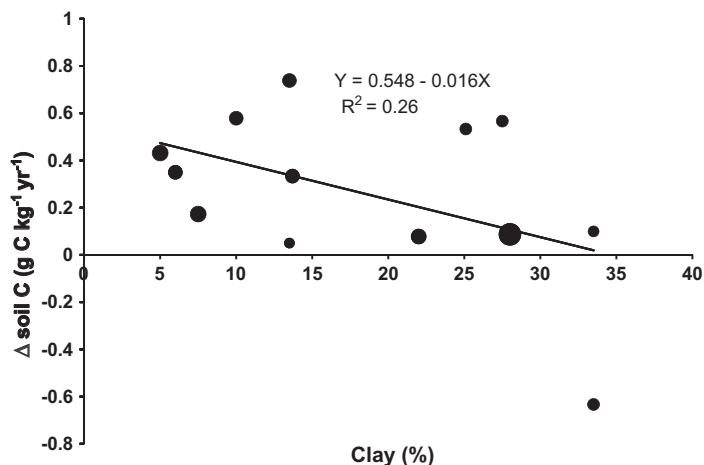
C. N₂O Emission						
	All sites			Fertilized sites		
	<i>n</i>	<i>Corr. C.</i>	<i>P</i>	<i>n</i>	<i>Corr. C.</i>	<i>P</i>
MAT	8	0.63	0.09	6	0.51	0.24
MAP	7	0.06	0.86	5	-0.16	0.85
pH	7	-0.10	0.90	5	0.36	0.50
% Clay	9	0.62	0.09	7	0.60	0.21

D. CH₄ Exchange			
	Non-rice		
	<i>n</i>	<i>Corr. C.</i>	<i>P</i>
MAT	5	-0.15	0.85
MAP	4	0.66	0.32
pH	4	0.24	0.76
% Clay	5	0.87	0.07

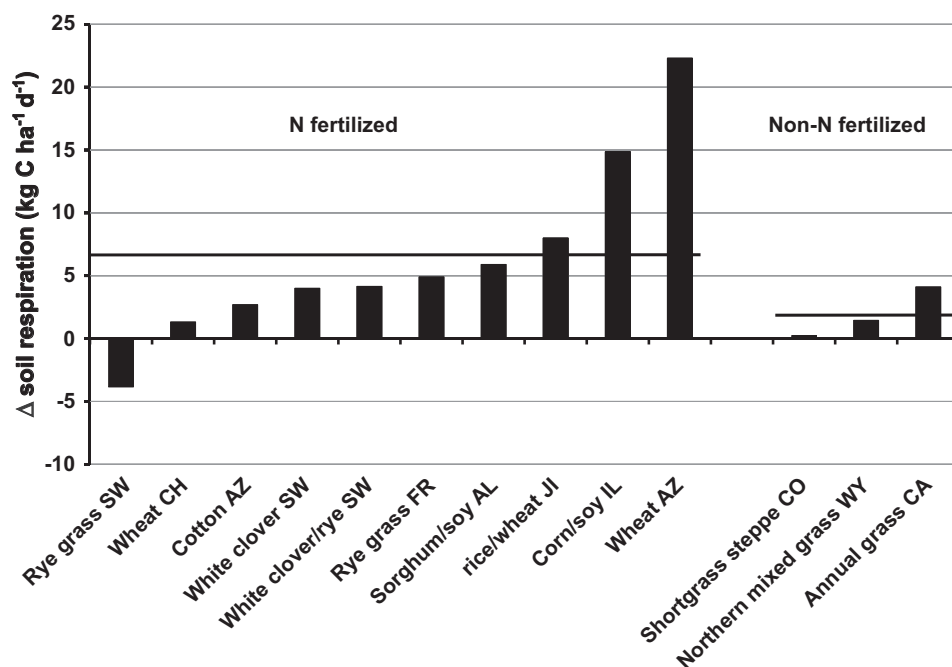
**n*: number of sites included in the regression; *Corr. C.*: Pearson's correlation coefficient; *P*: *P*-value of linear regression.

et al., 1998). Because root respiration was included in all studies, it is not surprising that soil respiration increased under eCO₂ in most studies. Elevated CO₂ generally increases plant productivity in agroecosystems (Kimball et al., 2002), and thus the increase in soil respiration under eCO₂ may largely have been driven by an increase in root production. The effect of eCO₂ on soil respiration was on average 6.4 and 1.9 kg C ha⁻¹ d⁻¹ in N fertilized and non-N fertilized studies respectively. Again, the greater eCO₂ effect on soil respiration in N fertilized studies may have been caused by an increase in root productivity and respiration that often occurs under eCO₂ with N additions (Van Groenigen et al., 2006).

Of the two climate and two soil parameters that we tested, both soil pH and %clay explained most of the variability in eCO₂ effects on soil respiration, although only the relationship with % clay was significant (*P* = 0.03, Table 27.3). When we included both N fertilized and non-N fertilized sites in the regression, the increase in soil respiration in response to eCO₂ increased with increased clay content, explaining 45% of the variability, and decreased with increased soil pH, explaining 48% of the variability (Figure 27.4). As was argued for the relationship between

**FIGURE 27.2**

The rate of change in soil C in response to eCO₂ in the N fertilized studies as a function of the soil clay content. The size of the dots indicate the weight used in the regression (i.e. bigger dots have more weight).

**FIGURE 27.3**

The change in soil respiration in response to eCO₂ among different studies. Horizontal bold lines represent averaged values for N fertilized and non-N fertilized studies.

clay content and eCO₂ effects on soil C, an increase in rhizosphere priming under eCO₂ may have resulted in larger eCO₂ effects on soil respiration with increased clay content. The negative relationship with soil pH is less clear. Microbial community composition and enzyme activity are often strongly affected by soil pH (Sinsabaugh et al., 2008; Fierer et al., 2009) that can be altered by changes in substrate inputs (Aciego Pietri and Brookes, 2009). It is, however, unclear to what degree the negative relationship that we observed between soil pH and soil respiration in response to eCO₂ was caused by changes in microbial or plant respiration.

THE EFFECT OF eCO₂ ON N₂O EMISSION

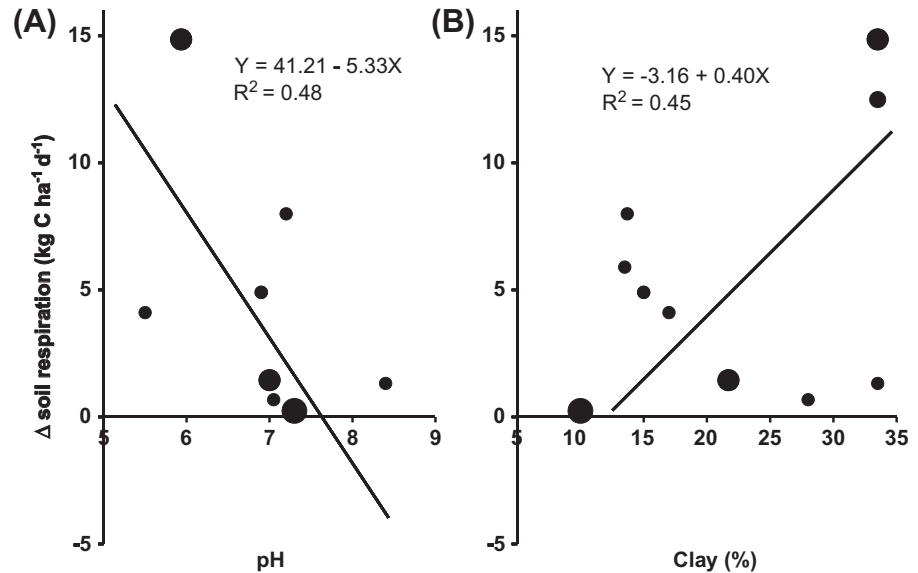
Of the 8 N fertilized studies that we found, the N₂O emission increased under eCO₂ in 6 studies. The average increase in N fertilized studies was 9.3 g N ha⁻¹ d⁻¹, while in the 3 non-N fertilized studies eCO₂ had hardly any effect on N₂O emission with an average decrease of

SECTION 7

Looking Ahead: Opportunities for Future Research and Collaboration

FIGURE 27.4

The change in soil respiration in response to $e\text{CO}_2$ as a function of (A) soil pH and (B) clay content. The size of the dots indicate the weight used in the regression (i.e. bigger dots have more weight).



$0.5 \text{ g N ha}^{-1} \text{ d}^{-1}$ (Figure 27.5). With a global warming potential 298 times greater than CO_2 (IPCC, 2007), N_2O emission in N fertilized studies correspond on average to $1188 \text{ g C-CO}_2 \text{ equivalents ha}^{-1} \text{ d}^{-1}$. This is slightly less than the average rate that we calculated for C sequestration in the top 20 cm of the soil in N fertilized studies in response to $e\text{CO}_2$ (see above). This suggests that, although N fertilization has the potential to increase soil C under $e\text{CO}_2$ (Van Groenigen et al., 2006), these soil C gains can potentially be almost completely offset by increased N_2O emissions under $e\text{CO}_2$ when N fertilizer is applied.

In the N fertilized studies N_2O emission in response to $e\text{CO}_2$ showed large variation between a decrease of $0.2 \text{ g N ha}^{-1} \text{ d}^{-1}$ in white clover in Switzerland (Baggs et al., 2003) and an increase of $38 \text{ g N ha}^{-1} \text{ d}^{-1}$ in rye grass at the same site in Switzerland (Ineson et al., 1998). This large variation is to a great extent caused by the timing and frequency of measurements after the N fertilizer application. For instance, Ineson et al. (1998) observed some of the

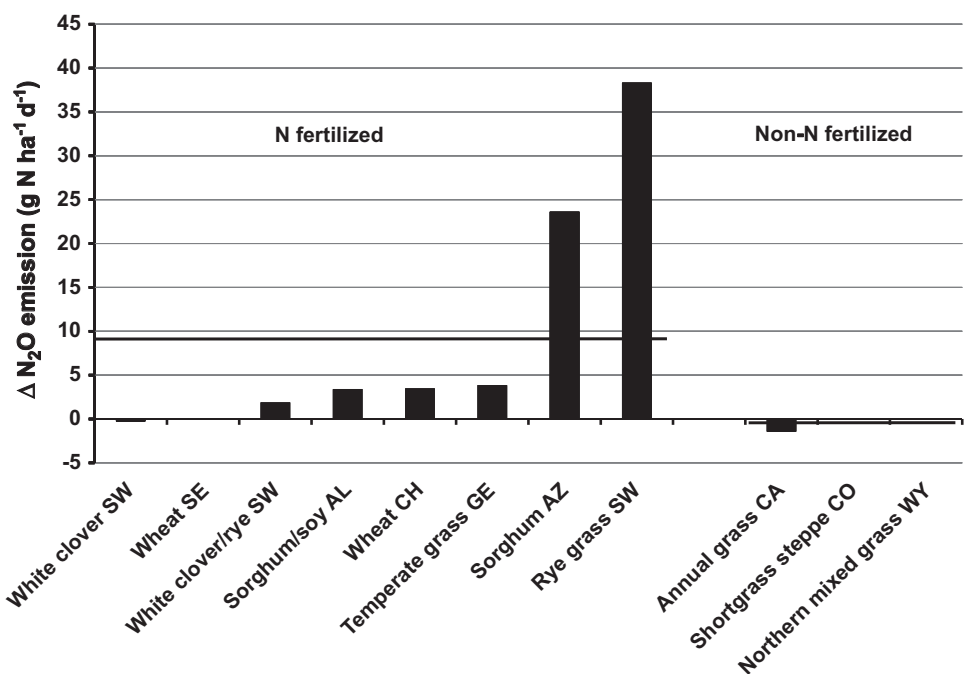
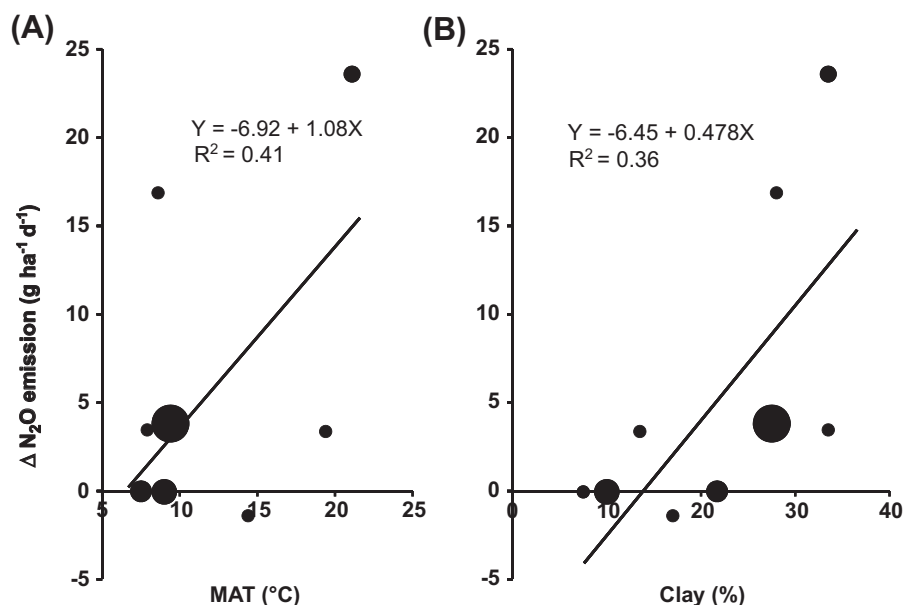


FIGURE 27.5

The change in N_2O emission in response to $e\text{CO}_2$ among different studies. Horizontal bold lines represent averaged values for N fertilized and non-N fertilized studies.

**FIGURE 27.6**

The change in N₂O emission in response to eCO₂ as a function of (A) mean annual temperature (MAT) and (B) clay content. The size of the dots indicate the weight used in the regression (i.e. bigger dots have more weight).

highest N₂O emissions ever recorded directly after N fertilizer application in grassland systems. Because of these high rates of N₂O emission, eCO₂ effects on N₂O emissions can also be high directly after N application (Ineson et al., 1998; Welzmler et al., 2008). On the other hand, Kammann et al. (2008) measured the effect of eCO₂ on N₂O emissions during 9 years in a temperate grassland in Germany (the longest study conducted on the effect of eCO₂ on N₂O emission) and observed the greatest eCO₂ effects during vegetative growth periods in the summer when soil mineral N concentrations were low, while eCO₂ had no effect on N₂O emission directly after the N application in the spring. Regardless of the timing and frequency of measurements in relation to N fertilizer application, the majority of N fertilized studies showed an increase in N₂O emission in response to eCO₂. It has been suggested that the increase in N₂O emission under eCO₂ in some of these N fertilized studies was caused by an increase in labile C substrates fueling denitrification (Ineson et al., 1998; Kammann et al., 2008).

In contrast, no eCO₂ effect, or even a slight reduction in N₂O emissions, was observed in the 3 non-N fertilized studies. Possibly, eCO₂ increased plant N uptake and reduced soil N availability in these unfertilized systems where available soil N was already low, causing no or reduced effects on N₂O emission (Hungate et al., 1997b; Mosier et al., 2002).

Both MAT and %clay showed a positive relationship with N₂O emission in response to eCO₂, although both relationships were only marginally significant (Table 27.3, Figure 27.6). N₂O emissions are highly sensitive to temperature (Grant and Pattey, 2008), which could explain why N₂O emissions respond more to eCO₂ in combination with higher MAT. Further, an increase in soil moisture, because of decreased stomatal conductance under eCO₂ (Kimball and Idso, 1983; Morgan et al., 2004; Wand et al., 1999), can increase anaerobic conditions in the soil conducive to denitrification, particularly in clayey soils that have relatively more small pores than sandy soils. These results suggest that, apart from N fertilization, MAT and soil texture play important roles in the large variability in N₂O emission in response to eCO₂ among different sites.

THE EFFECT OF eCO₂ ON CH₄ EXCHANGE

In studies with rice, eCO₂ resulted in large increases in CH₄ emission in 3 out of 4 studies (Figure 27.7). The average increase in CH₄ emission in rice studies in response to eCO₂ was

SECTION 7

Looking Ahead: Opportunities for Future Research and Collaboration

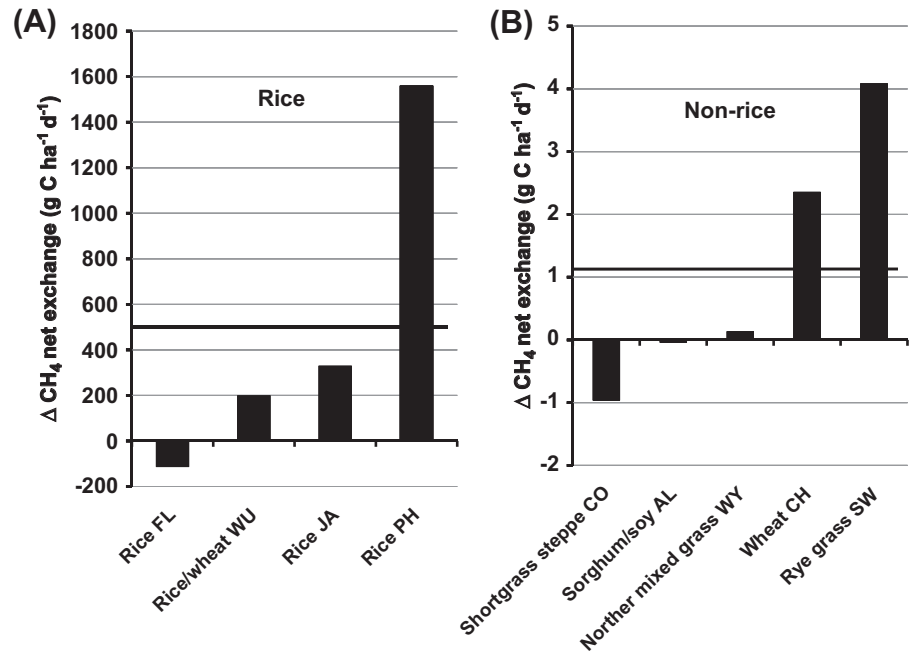


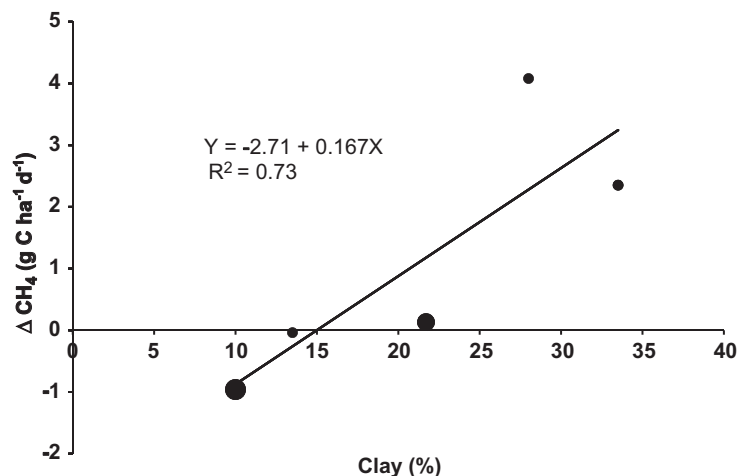
FIGURE 27.7

The change in CH₄ exchange in response to eCO₂ among different studies. Horizontal bold lines represent averaged values for (A) rice and (B) non-rice studies.

493 g C ha⁻¹ d⁻¹. With a global warming potential 25 times greater than CO₂ (IPCC, 2007), this increase in CH₄ emission corresponds to an increase of 4482 g C-CO₂ equivalents ha⁻¹ d⁻¹ in response to eCO₂. Evidently, rice paddy fields show some of the greatest responses to eCO₂ in terms of GHG emissions (e.g. the average rate of C-CO₂ equivalents associated with CH₄ emission in rice is 3.8 times higher than the average rate associated with N₂O emission in N fertilized studies).

In the rice studies, we found no relationships between MAT, MAP, soil pH, or %clay with the rate of CH₄ emission in response to eCO₂. CH₄ emission in rice fields is to a large degree controlled by inputs of C substrates, and increased CH₄ emission under eCO₂ has been associated with increased plant residues, root productivity, and exudation (Inubushi et al., 2003; Xu et al., 2004; Tokida et al., 2010). The increase in CH₄ emission in response to eCO₂ through increased inputs of C substrates may simply have overwhelmed any soil or external climate effect. We should note that with only 4 studies, we had limited statistical power to do the regressions.

In most non-rice studies, soil is a net sink for CH₄, where it is oxidized by methanotrophic bacteria [an exception was the study by Smith et al. (2010) where soil was sometimes a CH₄ source in a sorghum–soybean rotation]. The effect of eCO₂ on CH₄ fluxes (where we used the same convention as in the rice studies, i.e. a positive flux indicates CH₄ emission, while a negative flux indicates CH₄ uptake) was mixed where both increases and decreases were observed with an average increase of 1.1 g C ha⁻¹ d⁻¹ among the 5 studies we evaluated (or on average a *reduction* in CH₄ uptake in response to eCO₂ because in most studies there was an overall net CH₄ uptake; Figure 27.7). The reduced transpiration and consequent higher soil water content that often occurs under eCO₂ can have opposite effects on CH₄ fluxes depending on whether methanotroph activity is limited by soil moisture (in most arid and semiarid environments) or by CH₄ diffusivity into the soil (in most mesic environments; Dijkstra et al., 2011). Note that responses of CH₄ fluxes to eCO₂ in non-rice or dry land systems are orders of magnitude smaller than in rice systems. Nevertheless, because a much larger proportion of the global area is covered by dry land systems than by rice paddy fields, small changes in CH₄ fluxes in dry land systems can still have a significant impact on the global CH₄ flux (Mosier et al., 1991).

**FIGURE 27.8**

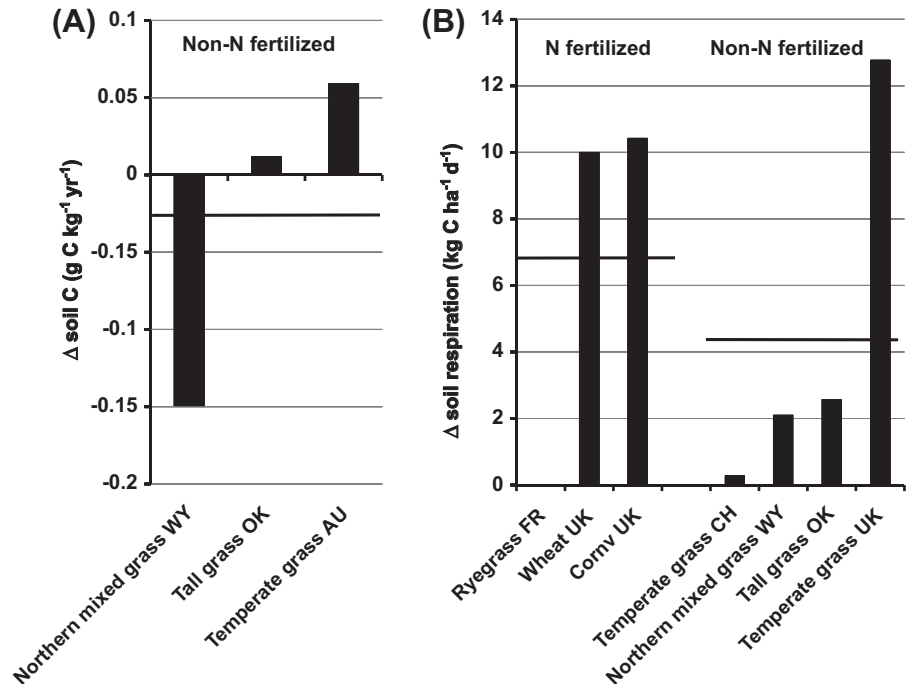
The change in CH₄ exchange in response to eCO₂ in non-rice studies as a function of soil clay content. The size of the dots indicate the weight used in the regression (i.e. bigger dots have more weight).

As with the greenhouse gases CO₂ and N₂O, the CH₄ flux in response to eCO₂ was positively related to the clay content of the soil, explaining 73% of the variation (Table 27.3, Figure 27.8). Soil texture should influence CH₄ fluxes to the extent that methanotroph activity is limited by soil moisture or by CH₄ diffusivity. Since sandy soils have better aeration than clayey soils, CH₄ uptake is more likely to be limited by direct effects of soil moisture on methanotroph activity than by CH₄ diffusivity. Thus, a CO₂-induced increase in soil moisture would tend to increase CH₄ uptake (thus *decrease* the CH₄ flux in response to eCO₂) more in sandy soils (e.g. as observed by Mosier et al., 2002, in a sandy loam with only 10% clay). On the other hand, in clayey soils with poorer aeration, CH₄ uptake is more likely to be limited by CH₄ diffusivity, and a CO₂-induced increase in soil moisture might therefore decrease CH₄ uptake (or increase the CO₂ response) as observed in a clay loam by Lam et al. (2011). The positive relationship of CH₄ fluxes with clay content is based on only five observations, and it remains to be seen if this relationship will hold with more observations.

WARMING EFFECTS ON SOIL C, SOIL RESPIRATION, N₂O EMISSION AND CH₄ EXCHANGE

Little work has been done on the effects of warming on soil C sequestration and GHG fluxes in agroecosystems. We found only two published studies (Luo et al., 2009; Pendall et al., 2011) and one unpublished study (northern mixed grass prairie in Wyoming, U.S.), all in non-N fertilized grassland systems, reporting warming effects on soil C. Results are not consistent among those three studies, with an average decrease in soil C by 0.026 g C kg⁻¹ soil yr⁻¹ (Figure 27.9A). The effect of warming on soil C in all three studies is relatively small compared to the effect of eCO₂ in many studies (Figure 27.1). It is noteworthy that warming induced C loss only in Wyoming northern mixed-grass prairie, the driest of these three grasslands. Although warming has the potential to enhance biological activity and extend the length of growing season, it also desiccates, and in dry grasslands, such desiccation can lead to C loss (Zhang et al., 2010).

A little more work has been done evaluating the effects of warming on soil respiration, with more consistent results. In 6 of 7 studies, soil respiration increased with warming (Figure 27.9B). The exception was a study with ryegrass in France, where no change in soil respiration was observed (Casella and Soussana, 1997). On average soil respiration increased more in N fertilized (6.8 kg C ha⁻¹ d⁻¹) than in non-N fertilized studies (4.4 kg C ha⁻¹ d⁻¹), although the highest increase was observed in a non-N fertilized study (Briones et al., 2009). Warming often leads to increased rates of SOM decomposition, and likely led to increased soil respiration (Rustad et al., 2001), particularly when N is not limited. We related soil respiration

**FIGURE 27.9**

The rate of change in (A) soil C and (B) soil respiration in response to warming among different studies. Horizontal bold lines represent the averaged values for N fertilized and non-N fertilized studies.

rates in response to warming to climate and soil parameters, but observed no significant relationships (data not shown).

Little information is available on how warming affects N₂O and CH₄ fluxes. In the only N fertilized study we found, the N₂O emission decreased in response to warming (Kamp et al., 1998), while in the two non-N fertilized studies warming had very little effect on N₂O emission (Hu et al., 2010, and unpublished results from northern mixed grassland, Wyoming, U.S., Figure 27.10A). Warming had mixed effects on CH₄ emission in rice paddy fields where both decreased (Ziska et al., 1998) and increased emission rates (Tokida et al., 2010) were reported (Figure 27.10B). The only non-rice study conducted in a semiarid grassland showed that warming decreased CH₄ uptake (Figure 27.10C). It was argued that in this semiarid climate, methanotroph activity was mostly directly limited by a soil moisture and that the drying effect of warming therefore directly reduced methanotroph activity (Dijkstra et al., 2011). Because of the limited number of studies, we did not perform regressions with climate and soil parameters.

INTERACTIVE eCO₂ × WARMING EFFECTS ON SOIL C, SOIL RESPIRATION, N₂O EMISSION, AND CH₄ EXCHANGE

Few studies included both atmospheric CO₂ and temperature manipulations (Ziska et al., 1998; Schroppe et al., 1999; Tokida et al., 2010; Dijkstra et al., 2011; Pendall et al., 2011). In only two studies were CO₂ × warming interactive effects on soil C investigated, both in non-N fertilized grassland systems. In both studies, soil C under eCO₂ decreased more in combination with warming than without warming (Table 27.4). In the northern mixed grassland soil respiration under eCO₂ also increased in combination with warming but slightly decreased without warming. These results suggest that eCO₂ effects on SOM decomposition rates may accelerate with increased temperature. However, we found little evidence for CO₂ × warming interactions on soil C from our regressions with MAT. Despite the relatively large numbers of studies included in this regression (19 studies, Table 27.3), we observed no significant relationship between soil C in response to eCO₂ and MAT, suggesting that eCO₂ effects on soil C sequestration did not depend on the temperature regime that the experiment was conducted

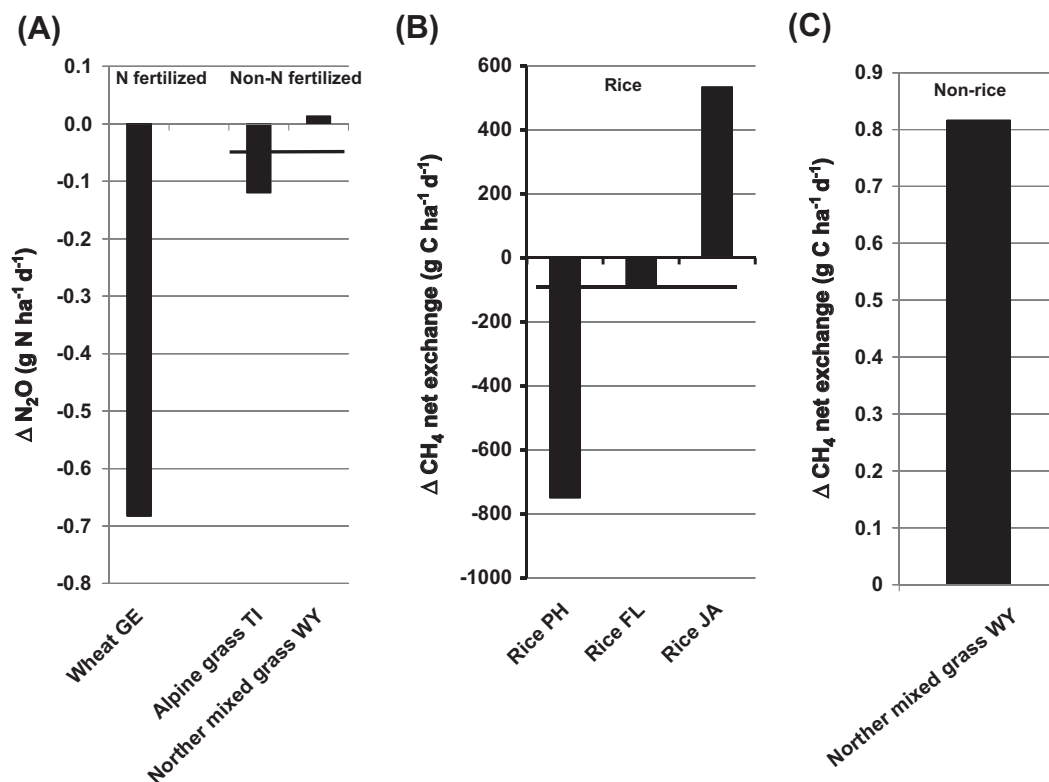


FIGURE 27.10

The change in (A) N₂O emission in N fertilized and non-N fertilized studies, (B) CH₄ exchange in rice studies, and (C) CH₄ exchange in non-rice studies in response to warming. Horizontal bold lines represent averaged values for non-N fertilized and rice studies.

TABLE 27.4 Interactive Effects of eCO₂ and Warming on Soil C, Soil Respiration, N₂O Emission, and CH₄ Exchange

Agroecosystem	eCO ₂ effect (% change from aCO ₂)		Reference
	Low temperature	High temperature	
Soil C—Non-fertilized			
Temperate grass AU	12.5	-3.4	Pendall et al. 2011
Northern mixed grass WY	-2.8	-14.1	Unpublished results
Respiration—Non-fertilized			
Northern mixed grass WY	-2.0	13.6	Unpublished results
N₂O—Non-fertilized			
Northern mixed grass WY	-94.9	-21.6	Unpublished results
CH₄—Rice			
Rice FL	-84.2	-90.0	Schrope et al. 1999
Rice JA	22.1	29.1	Tokida et al. 2010
Rice PH	48.3	214.5	Ziska et al. 1998
CH₄—Non-rice			
Northern mixed grass WY	0.1	5.0	Dijkstra et al. 2011 Unpublished results

in. Although studies were done at locations that occupied a relatively large range of MAT between 7.5 and 21.1°C, we note that reported MAT values do not always reflect the actual temperature that occurred during the time-frame of the experiment. This may have contributed to finding no significant relationship with eCO₂ effects on soil C.

Only N₂O emission in response to eCO₂ showed a marginally significant relationship with MAT (Figure 27.6), suggesting a CO₂ × temperature interactive effect where eCO₂ effects are stronger with increased temperature. In the non-N fertilized northern mixed grassland study in Wyoming, the N₂O emission under eCO₂ decreased less with warming than without warming (Table 27.4). These results suggest that CO₂ × warming interactive effects may be important for N₂O emissions.

The increase in CH₄ emission under eCO₂ was much greater in combination with warming than without warming in a study with rice in the Philippines (Ziska et al., 1998), but no CO₂ × warming interactive effects on CH₄ exchange were observed in three other studies (Table 27.4). Clearly, more research is needed to identify clear patterns of eCO₂ × warming interactive effects on soil C sequestration and GHG emissions.

CONCLUSIONS

We reviewed studies conducted in agroecosystems to determine sensitivity of C cycling and GHG emissions to the effects of eCO₂ and warming. We found that eCO₂ had the potential to increase soil C, particularly in combination with N fertilization. Similar results were found in other reviews (Reich et al., 2006b; Van Groenigen et al., 2006; Hungate et al., 2009). However, we also found that the increase in soil C in combination with N fertilization did not come without a price. N₂O emissions also increased under eCO₂ with N fertilization, and indeed on average the CO₂-induced increase in N₂O emission in terms of its Global Warming Potential almost completely offset the average increase in soil C in N fertilized studies. Thus, particularly in N fertilized agroecosystems, one can come to the wrong conclusion about the effect of eCO₂ on the GHG balance expressed in CO₂ equivalents if N₂O emissions are not accounted for. A similar conclusion was reached by van Groenigen et al. (2011), who estimated that the Global Warming Potential caused by a CO₂-induced increase in N₂O and CH₄ emission in agricultural and non-agricultural lands could offset as much as 16.6% of the global increase in terrestrial C storage in response to eCO₂ by 2050.

Research is needed to determine whether or not practices like precision application of N fertilizers or use of nitrification inhibitors can be used to minimize N₂O emission but that can help capitalize on the potential for rising CO₂ to enhance C sequestration. Legumes have been suggested as a possible remedy to the N-limitation problem of plants exposed to eCO₂ (van Groenigen et al., 2006) and legumes tend to respond positively to eCO₂ (Newton et al., 1994; Teyssonneyre et al., 2002). However, lack of a significant positive effect of CO₂ on C sequestration in pastures with legumes (Ross et al., 2004; van Kessel et al., 2006) suggests that simply enhanced N fixation under increasingly higher CO₂ concentrations may not necessarily lead to greater C sequestration. More research is needed to evaluate how and under what conditions various mixtures of legumes and forage grasses in combination with non-N fertilization practices might lead to increased C sequestration under future CO₂-enriched atmospheres. Such research should consider appropriate combinations of legumes and non-legumes whose morphology and development might optimize the capture and cycling of N so as to minimize the release of N₂O.

We found that soil clay content is an important factor in explaining the large variability in GHG exchange among sites in response to eCO₂. We observed marginally significant to significant positive relationships between %clay and all three GHGs in response to eCO₂. Our results suggest that GHG exchange from clayey soils is more sensitive to eCO₂ than from sandy soils. The relationships we found between %clay and GHG exchange should be explored

further to better understand the mechanisms involved and how widely applicable this relationship might be to scale up the effect of eCO₂ on GHG exchange in agroecosystems to regional or even global levels.

More research is needed about warming effects on soil C sequestration and GHG exchange. The limited studies that we found from agroecosystems often showed mixed warming effects, and no clear strong patterns emerged on how soil C and GHG exchange is affected by warming. Similarly, it remains unclear what the interactive effects of eCO₂ and warming are on soil C sequestration and GHG exchange in agroecosystems. While important challenges remain for agriculture to identify systems and practices that can mitigate global warming, future research with the objective to enhance C sequestration and mitigate GHG emissions will need to remain vigilant as climate change continues to increase in coming decades.

Acknowledgments

We would like to thank Jeffrey Amthor and Carina Moeller for helpful comments on a previous version of the manuscript.

References

- Aciego Pietri, J.C., Brookes, P.C., 2009. Substrate inputs and pH as factors controlling microbial biomass, activity and community structure in an arable soil. *Soil Biol. Biochem.* 41, 1396–1405.
- Baggs, E.M., Richter, M., Hartwig, U.A., Cadisch, G., 2003. Nitrous oxide emissions from grass swards during the eighth year of elevated atmospheric pCO₂ (Swiss FACE). *Glob. Change Biol.* 9, 1214–1222.
- Briones, M.J.I., Ostle, N.J., McNamara, N.P., Poskitt, J., 2009. Functional shifts of grassland soil communities in response to soil warming. *Soil Biol. Biochem.* 41, 315–322.
- Casella, E., Soussana, J.F., 1997. Long-term effects of CO₂ enrichment and temperature increase on the carbon balance of a temperate grass sward. *J. Exp. Bot.* 48, 1309–1321.
- Cheng, L., Leavitt, S.W., Kimball, B.A., Pinter Jr., P.J., Ottman, M.J., Matthias, A., Wall, G.W., Brooks, T., Williams, D.G., Thompson, T.L., 2007. Dynamics of labile and recalcitrant soil carbon pools in a sorghum free-air CO₂ enrichment (FACE) agroecosystem. *Soil Biol. Biochem.* 39, 2250–2263.
- Cheng, W., 1999. Rhizosphere feedbacks in elevated CO₂. *Tree Physiol.* 19, 313–320.
- Conant, R.T., Paustian, K., 2002. Spatial variability of soil organic carbon in grasslands: implications for detecting change at different scales. *Environ. Poll.* 116, 127–135.
- Dijkstra, F.A., Cheng, W., 2007. Moisture modulates rhizosphere effects on C decomposition in two different soil types. *Soil Biol. Biochem.* 39, 2264–2274.
- Dijkstra, F.A., Morgan, J.A., Von Fischer, J.C., Follett, R.F., 2011. Elevated CO₂ and warming effects on CH₄ uptake in a semiarid grassland below optimum soil moisture. *J. Geophys. Res.—Biogeosci.* 116, G01007. doi: 10.1029/2010JG001288.
- Epstein, H.E., Lauenroth, W.K., Burke, I.C., Coffin, D.P., 1997. Productivity patterns of C₃ and C₄ functional types in the U.S. Great Plains. *Ecology* 78, 722–731.
- Fierer, N., Grandy, A.S., Six, J., Paul, E.A., 2009. Searching for unifying principles in soil ecology. *Soil Biol. Biochem.* 41, 2249–2256.
- Giesemann, A., Weigel, H.J., 2008. Soil carbon isotopic composition and soil carbon content in an agroecosystem during six years of Free Air Carbon dioxide Enrichment (FACE). *Isotopes Environ. Health Studies.* 44, 349–363.
- Grant, R.F., Pattey, E., 2008. Temperature sensitivity of N₂O emissions from fertilized agricultural soils: mathematical modeling in ecosys. *Glob. Biogeochem. Cycles.* 22, GB4019. doi:10.1029/2008GB003273.
- Guo, Y., Gong, P., Amundson, R., Yu, Q., 2006. Analysis of factors controlling soil carbon in the conterminous United States. *Soil Sci. Soc. Am. J.* 70, 601–612.
- Hartley, I.P., Heinemeyer, A., Evans, S.P., Ineson, P., 2007. The effect of soil warming on bulk soil vs. rhizosphere respiration. *Glob. Change Biol.* 13, 2654–2667.
- Heimann, M., Reichstein, M., 2008. Terrestrial carbon dynamics and climate feedbacks. *Nature* 451, 289–292.
- Hu, S., Chapin III, F.S., Firestone, M.K., Field, C.B., Chiariello, N.R., 2001. Nitrogen limitation of microbial decomposition in a grassland under elevated CO₂. *Nature* 409, 188–191.
- Hu, Y., Chang, X., Lin, X., Wang, Y., Wang, S., Duan, J., Zhang, Z., Yang, X., Luo, C., Xu, G., Zhao, X., 2010. Effects of warming and grazing on N₂O fluxes in an alpine meadow ecosystem on the Tibetan plateau. *Soil Biol. Biochem.* 42, 944–952.

- Hungate, B.A., Holland, E.A., Jackson, R.B., Chapin III, F.S., Mooney, H.A., Field, C.B., 1997a. The fate of carbon in grassland under carbon dioxide enrichment. *Nature* 388, 576–579.
- Hungate, B.A., Lund, C.P., Pearson, H.L., Chapin III, F.S., 1997b. Elevated CO₂ and nutrient addition alter soil N cycling and N trace gas fluxes with early season wet-up in a California annual grassland. *Biogeochem.* 37, 89–109.
- Hungate, B.A., van Groenigen, K.J., Six, J., Jastrow, J.D., Luo, Y., de Graaff, M.A., van Kessel, C., Osenberg, C.W., 2009. Assessing the effect of elevated carbon dioxide on soil carbon: a comparison of four meta-analyses. *Glob. Change Biol.* 15, 2020–2034.
- Ineson, P., Coward, P.A., Hartwig, U.A., 1998. Soil gas fluxes of N₂O, CH₄ and CO₂ beneath *Lolium perenne* under elevated CO₂: the Swiss free air carbon dioxide enrichment experiment. *Plant Soil* 198, 89–95.
- Inubushi, K., Cheng, W., Aonuma, S., Hoque, M.M., Kobayashi, K., Miura, S., Kim, H.Y., Okada, M., 2003. Effects of free-air CO₂ enrichment (FACE) on CH₄ emission from a rice paddy field. *Glob. Change Biol.* 9, 1458–1464.
- Intergovernmental Panel on Climate Change, 2007. Working Group I Report. The Physical Science Basis. Technical Summary Available online at: <http://www.ipcc.ch> (verified January 17, 2011).
- Islam, K.R., Mulchi, C.L., Ali, A.A., 2000. Interactions of tropospheric CO₂ and O₃ enrichments and moisture variations on microbial biomass and respiration in soil. *Glob. Change Biol.* 6, 255–265.
- Jäger, H.J., Schmidt, S.W., Kammann, C., Grünhage, L., Müller, C., Hanewald, K., 2003. The University of Giessen Free-Air Carbon dioxide Enrichment study: description of the experimental site and of a new enrichment system. *J. Appl. Bot.* 77, 117–127.
- Juan, L., Yong, H., Zucong, C., Huilin, L., 2007. Changes in CH₄ and CO₂ emissions from soils under flooded conditions after being exposed to FACE (free-air CO₂ enrichment) for three years. *Acta Ecologica Sinica* 27, 2184–2190.
- Kammann, C., Müller, C., Grünhage, L., H- Jäger, J., 2008. Elevated CO₂ stimulates N₂O emissions in permanent grassland. *Soil Biol. Biochem.* 40, 2194–2205.
- Kamp, T., Steindl, H., Hantschel, R.E., Beese, F., Munch, J.C., 1998. Nitrous oxide emissions from a fallow and wheat field as affected by increased soil temperatures. *Biol. Fert. Soils.* 27, 307–314.
- Kimball, B.A., Idso, S.B., 1983. Increasing atmospheric CO₂: effects on crop yield, water use, and climate. *Agric. Water Manage* 7, 55–72.
- Kimball, B.A., Kobayashi, K., Bindu, M., 2002. Responses of Agricultural Crops to Free-air CO₂ Enrichment. *Adv. Agron.* vol. 77 Academic Press.
- Kou, T., Zhu, J., Xie, Z., Hasegawa, T., Heiduk, K., 2007. Effect of elevated atmospheric CO₂ concentration on soil and root respiration in winter wheat by using a respiration partitioning chamber. *Plant Soil* 299, 237–249.
- Lam, S.K., Lin, E., Norton, R., Chen, D., 2011. The effect of increased atmospheric carbon dioxide concentration on emissions of nitrous oxide, carbon dioxide and methane from a wheat field in a semi-arid environment in northern China. *Soil Biol. Biochem.* 43, 458–461.
- Leavitt, S.W., Paul, E.A., Galadima, A., Nakayama, F.S., Danzer, S.R., Johnson, H., Kimball, B.A., 1996. Carbon isotopes and carbon turnover in cotton and wheat FACE experiments. *Plant Soil* 187, 147–155.
- Leavitt, S.W., Pendall, E., Paul, E.A., Brooks, T., Kimball, B.A., Pinter Jr., P.J., Johnson, H.B., Matthias, A., Wall, G.W., LaMorte, R.L., 2001. Stable-carbon isotopes and soil organic carbon in wheat under CO₂ enrichment. *New Phytol* 150, 305–314.
- Liu, W., Zhang, Z., Wan, S., 2009. Predominant role of water in regulating soil and microbial respiration and their responses to climate change in a semiarid grassland. *Glob. Change Biol.* 15, 184–195.
- Luo, Y., Jackson, R.B., Field, C.B., Mooney, H.A., 1996. Elevated CO₂ increases belowground respiration in California grasslands. *Oecologia* 108, 130–137.
- Luo, Y., Sherry, R., Zhou, X., Wan, S., 2009. Terrestrial carbon-cycle feedback to climate warming: experimental evidence on plant regulation and impacts of biofuel feedstock harvest. *GCB Bioenergy* 1, 62–74.
- Marhan, S., Kandeler, E., Rein, S., Fangmeier, A., Niklaus, P.A., 2010. Indirect effects of soil moisture reverse soil C sequestration responses of a spring wheat agroecosystem to elevated CO₂. *Glob. Change Biol.* 16, 469–483.
- Morgan, J.A., Pataki, D.E., Korner, C., Clark, H., DelGrosso, S.J., Grunzewig, J.M., Knapp, A.K., Mosier, A.R., Newton, P.C.D., Niklaus, P.A., Nippert, J.B., Nowak, R.S., Parton, W.J., Polley, H.W., Shaw, M.R., 2004. Water relations in grassland and desert ecosystems to elevated atmospheric CO₂. *Oecologia* 140, 11–25.
- Mosier, A., Schimel, D., Valentine, D., Bronson, K., Parton, W., 1991. Methane and nitrous oxide fluxes in native, fertilized and cultivated grasslands. *Nature* 350, 330–332.
- Mosier, A.R., Morgan, J.A., King, J.Y., LeCain, D., Milchunas, D.G., 2002. Soil-atmosphere exchange of CH₄, CO₂, NO_x and N₂O in the Colorado shortgrass steppe under elevated CO₂. *Plant Soil* 240, 201–211.
- Nakayama, F.S., Huluka, G., Kimball, B.A., Lewin, K.F., Nagy, J., Hendrey, G.R., 1994. Soil carbon dioxide fluxes in natural and CO₂-enriched systems. *Agric. For. Meteorol.* 70, 131–140.

- Newton, P.C.D., Clark, H., Bell, C.C., Glasgow, E.M., Campbell, B.D., 1994. Effects of elevated CO₂ and simulated seasonal changes in temperature on the species composition and growth rate of pasture turves. *Ann. Bot.* 73, 53–59.
- Niklaus, P.A., Körner, C., 1996. Responses of soil microbiota of a late successional alpine grassland to long term CO₂ enrichment. *Plant Soil* 184, 219–229.
- Niklaus, P.A., Wohlfender, M., Siegwolf, R., Körner, C., 2001. Effects of six years atmospheric CO₂ enrichment on plant, soil, and soil microbial C of a calcareous grassland. *Plant Soil* 233, 189–202.
- Pendall, E., Leavitt, S.W., Brooks, T., Kimball, B.A., Pinter Jr., P.J., Wall, G.W., LaMorte, R.L., Wechsung, G., Wechsung, F., Adamsen, F., Matthias, A.D., Thompson, T.L., 2001. Elevated CO₂ stimulates soil respiration in a FACE wheat field. *Basic Appl. Ecol.* 2, 193–201.
- Pendall, E., Mosier, A.R., Morgan, J.A., 2004. Rhizodeposition stimulated by elevated CO₂ in a semiarid grassland. *New Phytol* 162, 447–458.
- Pendall, E., Osanai, Y.U.I., Williams, A.L., Hovenden, M.J., 2011. Soil carbon storage under simulated climate change is mediated by plant functional type. *Glob. Change Biol.* 17, 505–514.
- Peralta, A.L., Wander, M.M., 2008. Soil organic matter dynamics under soybean exposed to elevated [CO₂]. *Plant Soil* 303, 69–81.
- Pleijel, H., Sild, J., Danielsson, H., Klemetsson, L., 1998. Nitrous oxide emissions from a wheat field in response to elevated carbon dioxide concentration and open-top chamber enclosure. *Environ. Poll.* 102, 167–171.
- Prior, S.A., Runion, G.B., Rogers, H.H., Torbert, H.A., Reeves, D.W., 2005. Elevated atmospheric CO₂ effects on biomass production and soil carbon in conventional and conservation cropping systems. *Glob. Change Biol.* 11, 657–665.
- Prior, S.A., Runion, G.B., Torbert, H.A., Rogers, H.H., 2004. Elevated atmospheric CO₂ in agroecosystems: soil physical properties. *Soil Science* 169, 434–439.
- Prior, S.A., Torbert, H.A., Runion, G.B., Rogers, H.H., Wood, C.W., Kimball, B.A., LaMorte, R.L., Pinter, P.J., Wall, G.W., 1997. Free-air carbon dioxide enrichment of wheat: soil carbon and nitrogen dynamics. *J. Environ. Qual.* 26, 1161–1166.
- Reich, P.B., Hobbie, S.E., Lee, T., Ellsworth, D.S., West, J.B., Tilman, D., Knops, J.M.H., Naeem, S., Trost, J., 2006a. Nitrogen limitation constrains sustainability of ecosystem response to CO₂. *Nature* 440, 922–925.
- Reich, P.B., Hungate, B.A., Luo, Y., 2006b. Carbon-nitrogen interactions in terrestrial ecosystems in response to rising atmospheric carbon dioxide. *Ann. Rev. Ecol. Evol. System.* 37, 611–636.
- Ross, D.J., Newton, P.C.D., Tate, K.R., 2004. Elevated [CO₂] effects on herbage production and soil carbon and nitrogen pools and mineralization in a species-rich, grazed pasture on a seasonally dry sand. *Plant Soil* 260, 183–196.
- Rustad, L., Campbell, J., Marion, G., Norby, R., Mitchell, M., Hartley, A., Cornelissen, J., Gurevitch, J., GCTE-NEWS, 2001. A meta-analysis of the response of soil respiration, net nitrogen mineralization, and aboveground plant growth to experimental ecosystem warming. *Oecologia* 126, 543–562.
- Schrope, M.K., Chanton, J.P., Allen, L.H., Baker, J.T., 1999. Effect of CO₂ enrichment and elevated temperature on methane emissions from rice, *Oryza sativa*. *Glob. Change Biol.* 5, 587–599.
- Sinsabaugh, R.L., Lauber, C.L., Weintraub, M.N., Ahmed, B., Allison, S.D., Crenshaw, C., Contosta, A.R., Cusack, D., Frey, S., Gallo, M.E., Gartner, T.B., Hobbie, S.E., Holland, K., Keeler, B.L., Powers, J.S., Stursova, M., Takacs-Vesbach, C., Waldrop, M.P., Wallenstein, M.D., Zak, D.R., Zeglin, L.H., 2008. Stoichiometry of soil enzyme activity at global scale. *Ecol. Lett.* 11, 1252–1264.
- Smith, P., 2004. How long before a change in soil organic carbon can be detected? *Glob. Change Biol.* 10, 1878–1883.
- Smith, K.E., Runion, G.B., Prior, S.A., Rogers, H.H., Torbert, H.A., 2010. Effects of elevated CO₂ and agricultural management on flux of greenhouse gases from soil. *Soil Sci.* 175, 349–356.
- Solomon, S., Plattner, G.K., Knutti, R., Friedlingstein, P., 2009. Irreversible climate change due to carbon dioxide emissions. *Proc. Nat. Acad. Sci.* 106, 1704–1709.
- Teyssonneyre, F., Picon-Cochard, C., Falcimagne, R., J- Sousanna, R., 2002. Effects of elevated CO₂ and cutting frequency on plant community structure in a temperate grassland. *Glob. Change Biol.* 8, 1034–1046.
- Tokida, T., Fumoto, T., Cheng, W., Matsunami, T., Adachi, M., Katayanagi, N., Matsushima, M., Okawara, Y., Nakamura, H., Okada, M., Sameshima, R., Hasegawa, T., 2010. Effects of free-air CO₂ enrichment (FACE) and soil warming on CH₄ emission from a rice paddy field: impact assessment and stoichiometric evaluation. *Biogeosciences* 7, 2639–2653.
- Van Groenigen, K.J., Six, J., Hungate, B.A., De Graaff, M.A., Van Breemen, N., Van Kessel, C., 2006. Element interactions limit soil carbon storage. *Proc. Nat. Acad. Sci.* 103, 6571–6574.
- Van Groenigen, K.J., Osenberg, C.W., Hungate, B.A., 2011. Increased soil emissions of potent greenhouse gases under increased atmospheric CO₂. *Nature* 475, 214–216.

- Van Kessel, C., Boots, B., de Graaff, M.A., Harris, D., Blum, H., Six, J., 2006. Total soil C and N sequestration in a grassland following 10 years of free air CO₂ enrichment. *Glob. Change Biol.* 12, 2187–2199.
- Van Kessel, C., Horwath, W.R., Hartwig, U., Harris, D., Lüscher, A., 2000. Net soil carbon input under ambient and elevated CO₂ concentrations: isotopic evidence after 4 years. *Glob. Change Biol.* 6, 435–444.
- Ward, S.J.E., Midgley, G.F., Jones, M.H., Curtis, P.S., 1999. Responses of wild C₄ and C₃ grasses (Poaceae) species to elevated atmospheric CO₂ concentrations: a meta-analytic test of current theories and perceptions. *Glob. Change Biol.* 5, 723–741.
- Welzmler, J.T., Matthias, A.D., White, S., Thompson, T.L., 2008. Elevated carbon dioxide and irrigation effects on soil nitrogen gas exchange in irrigated sorghum. *Soil Sci. Soc. Am. J.* 72, 393–401.
- Williams, M.A., Rice, C.W., Owensby, C.E., 2000. Carbon dynamics and microbial activity in tallgrass prairie exposed to elevated CO₂ for 8 years. *Plant Soil* 227, 127–137.
- Wood, C.W., Torbert, H.A., Rogers, H.H., Runion, G.B., Prior, S.A., 1994. Free-air CO₂ enrichment effects on soil carbon and nitrogen. *Agric. For. Meteor.* 70, 103–116.
- Wu, Z., Dijkstra, P., Koch, G.W., Peñuelas, J., Hungate, B.A., 2011. Responses of terrestrial ecosystems to temperature and precipitation change: a meta-analysis of experimental manipulation. *Glob. Change Biol.* 17, 927–942.
- Xia, J., Han, Y., Zhang, Z., Wan, S., 2009. Effects of diurnal warming on soil respiration are not equal to the summed effects of day and night warming in a temperate steppe. *Biogeosciences* 6, 1361–1370.
- Xu, Z., Zheng, X., Wang, Y., Han, S., Huang, Y., Zhu, J., Butterbach-Bahl, K., 2004. Effects of elevated CO₂ and N fertilization on CH₄ emissions from paddy rice fields. *Glob. Biogeochem. Cycles* 18, GB3009.
- Zhang, L., Wylie, B.K., Ji, L., Gilmanov, T.G., Tieszen, L.L., 2010. Climate-driven interannual variability in net ecosystem exchange in the Northern Great Plains grasslands. *Rangeland Ecol. Manag.* 63, 40–50.
- Zheng, X., Zhou, Z., Wang, Y.Y., Zhu, J., Wang, Y., Yue, J., Shi, Y., Kobayashi, K., Inubushi, K., Huang, Y., Han, S., Xu, Z., Xie, B., Butterbach-Bahl, K., Yang, L., 2006. Nitrogen-regulated effects of free-air CO₂ enrichment on methane emissions from paddy rice fields. *Glob. Change Biol.* 12, 1717–1732.
- Zhong, S., Liang, W., Lou, Y., Li, Q., Zhu, J., 2009. Four years of free-air CO₂ enrichment enhance soil C concentrations in a Chinese wheat field. *J. Env. Sci.* 21, 1221–1224.
- Zhou, X., Wan, S., Luo, Y., 2007. Source components and interannual variability of soil CO₂ efflux under experimental warming and clipping in a grassland ecosystem. *Glob. Change Biol.* 13, 761–775.
- Ziska, L.H., Moya, T.B., Wassmann, R., Namuco, O.S., Lantin, R.S., Aduna, J.B., Abao Jr., E., Bronson, K.F., Neue, H.U., Olszyk, D., 1998. Long-term growth at elevated carbon dioxide stimulates methane emission in tropical paddy rice. *Glob. Change Biol.* 4, 657–665.