

Review

Applications of acoustics in insect pest management

R. W. Mankin*

Address: USDA-ARS Center for Medical, Agricultural, and Veterinary Entomology, 1700 SW 23rd Drive, Gainesville, FL, USA.

***Correspondence:** Email: Richard.Mankin@ars.usda.gov

Received: 28 October 2011

Accepted: 5 December 2011

doi: 10.1079/PAVSNNR20127001

The electronic version of this article is the definitive one. It is located here: <http://www.cabi.org/cabreviews>

© CAB International 2012 (Online ISSN 1749-8848)

Abstract

Acoustic technology has been applied for many years in studies of insect communication and in the monitoring of calling-insect population levels, geographic distributions and species diversity, as well as in the detection of cryptic insects in soil, wood, container crops and stored products. Acoustic devices of various sizes and power levels have been used successfully to trap insect pests that exhibit phonotaxis or other orientation behaviours, including mosquitoes, midges, mole crickets, field crickets, moths, cockroaches and Tephritid fruit flies. The attractiveness of traps depends on the behaviour, physiological state and age of the target insect, and varies with several environmental factors, including temperature and light level. Widespread adoption of acoustics for trapping has been limited by the costs of instrumentation and the relatively small segments of insect populations (e.g. mate-seeking adults of a limited age-range) that are attracted to a sound source, but trapping effectiveness often can be improved by adding swarm markers, chemical attractants or black lights, and by precisely timing temporal and frequency patterns to match the natural communication signals. There remains potential for using ultrasonic bat-cry signals to disrupt behaviour of night-flying insects, but ultrasonic signals have little effect on insects that are not normally preyed upon by bats. Potential areas for growth in the use of acoustic technology in pest management include the production of signals that disrupt vibrational communication, particularly in the Hemiptera, and the development of control treatments that combine pheromones and precisely patterned sonic or vibrational signals.

Keywords: Sound, Vibration, Ultrasound, Attraction, Trap, Phonotaxis

Review Methodology: Searches were performed in CAB Abstracts, Agricola, WorldWideScience.org and Google using keywords search terms (acoustic, insect, trap), names of researchers who have participated in acoustic and vibrational studies with insects. In addition, the author's personal library on acoustics, vibration, mosquitoes and fruit flies was inspected.

Introduction

Acoustic recording and playback technologies have been employed for insect detection and monitoring since the early 1900s [1] and the study of acoustic behaviour has developed into one of the prominent areas of insect ethology [2–5]. To consider applications for insect pest management, acoustic studies have been conducted to attract and trap insects [6, 7], and to manipulate their behaviours or interrupt intraspecific communication using either sound [8] or vibrational signals transmitted within host plants [9].

Traps using sound to attract insects were first reported in 1949 in studies where male *Anopheles albimanus*

Wiedemann mosquitoes were captured in experiments with loudspeakers [10, 11]. Subsequently, acoustic methods were developed in field and laboratory studies to trap other mosquitoes [12–16], Chironomid midges [17–20], *Scapteriscus* spp. mole crickets [7, 21], gryllid field crickets and their tachinid parasitoids [22, 23], *Achroia* and *Galleria* moths [24, 25], *Blattella germanica* (L.) cockroaches [26] and tephritid fruit flies [27–29]. In addition, chemosterilization of acoustically attracted male *Culex quinquefasciatus* Say was conducted with moderate success [30], and male *Aedes albopictus* (Skuse) populations were reduced 76% by attraction to loudspeakers producing 400 Hz tones at the centre of insecticide-treated black polyethylene sheets [31].

In several studies, acoustic traps have been deployed not just to capture, sterilize, or kill insects, but to collect live specimens for biological studies [32, 33] and biological control programmes [34, 35]. Acoustic methods also have been used in surveys to monitor insect diversity [36, 37], population levels [38–40] and geographic distributions [41].

Given the urgent need of regulatory agencies and pest managers to obtain improved tools to detect and manage growing numbers of invasive insect species, e.g. [42–44], it is worthwhile to reflect on the current usage of acoustic technology in integrated pest management programmes, as well as the potential for additional applications in the future. Recent reviews on the evolution and function of insect auditory systems [45] and the physiological, behavioural, ecological and evolutionary context of insect communication [46], and several recent studies on mosquito audition and mating behaviour [47–54] and parasitoid acoustic organs [55] provide helpful background information for this discussion.

Currently Available Sound and Vibration Production Technology

Sonic sources of different dimensions and power levels have been tested in trapping studies, population surveys and behavioural manipulation bioassays, including general-purpose loudspeakers [7, 15], large piezoplastic sheets on foam boards [14], acoustic lasers [28], small tweeters [55] and even tuning forks [15]. Harmoniums [56] and MP3-player speakers [57, 58] have been used to produce airborne sounds that excited vibrations within plants or other substrates containing targeted insects. Caged insects [22, 59], tethered flying insects [48], electrodynamic shakers [60] and piezoceramic actuators [61] have been used to produce sounds and vibrations in behavioural manipulation studies. The speakers with large dimensions and the acoustic laser were constructed to broadcast long-range, loud (>100 dB at 1 m) signals, based on findings that increased signal levels resulted in increased signal attractiveness and trap capture rates [62, 63]. Speakers with small dimensions have been used to produce spatially divergent sound fields to facilitate directional orientation [15, 55, 64, 65]. Some of the signals produced have been playbacks of original recordings [10, 57, 64], others have been synthesized [8, 14, 24, 66, 67]. In several studies [21, 49, 52], the responses to live or recorded signals were compared with responses to synthesized signals.

Relatively high costs of broadcasting over large areas compared with the costs for chemical attractants and pesticides have hampered development of acoustic technology for insect management until now. Frequency-dependent attenuation reduces the effective range of airborne and structural vibration signals greater than 100–200 Hz [1, 60, 68]. High-amplitude speakers can extend

the range of sonic broadcasts, but sufficiently powerful energy sources have not always been readily available in the field [6, 7].

Attraction and Trapping Devices

Insects of different species attracted to sound have been trapped by a variety of devices, including electrically charged screens [10, 11, 69], fans or vacuums [64] with collecting bags or cones with nets [66], adhesive cylinders [12, 70–72] and boards [14], funnel and bucket traps [73], or wood-and-screen silt traps [74]. The preferred trap-type depends partly on the size of the insect and its locomotory behaviours, i.e., flight or walking up or down a surface, preferences for crevices or holes, rough or smooth surfaces, etc. [75–78]. Frequently the trap captures are strongly affected by moonlight levels, wind and other factors [16]. In addition, the attractiveness of a trap is context-dependent [7, 22, 58], varying over time and over different segments of the population. For example, the calling sound of a male cricket can attract females, but at high intensities it can inhibit female locomotion or cause other males to move away [61].

The rates of captures in traps often are improved significantly by adding black cloth or other swarm markers, black light or other visual attractants [69, 79], or chemical attractants, depending on the target insect [7, 77, 78]. In the case of mosquitoes, live hamsters and dry ice were used to attract females in addition to the males that were attracted by sound [67].

The sound sources and traps currently in use are mature technology, not likely to change in the near future, but potential avenues for cost-reductions and for enhancements of the effectiveness of behavioural manipulations may be found in the design and implementation of controllers that set beginning and ending times and other temporal patterns of sound production to match the patterns of the targeted species [7]. Also, incorporation of automated counting and identification of the captured or detected insects [40, 80–82] into trapping and monitoring systems may improve their effectiveness in field experiments and integrated pest management programmes. Automated counting and identification is of benefit particularly in environments where servicing of traps might be difficult or dangerous, or when trapping data need to be collected in a timely manner.

Repulsion/Exclusion, Interference with Communication and Other Potential Applications of Acoustic Signals

It has been well documented that many species of insects subject to predation by bats will dive to the ground or move away when they detect ultrasonic signals that resemble echolocation calls [5, 7, 83]. Studies of ultrasonic signals on several moth pest species were conducted [84–87]. In

the latter study [87], a light trap containing an ultrasonic speaker was placed in a maize field containing *Heliothis zea* (Boddie) and *Ostrinia nubilalis* (Hübner). The speaker delivered 1 ms, 25 kHz pulses at rates of 1–10/s, simulating a range of pulse rates and durations emitted during echolocating cries by local insectivorous bats [88]. In this field study [87], high pulse rates decreased light-trap catches more effectively than low pulse rates, and the *O. nubilalis* were affected more strongly than *H. zea*. The results from these and related studies suggested that the effects of habituation and sound shadows rendered ultrasonic repellent signals ineffective for reducing oviposition or for reducing economic damage. It was proposed that the effects of habituation might be reduced by: (1) reducing the signal intensity, (2) presenting the pulses at unpredictable intervals, (3) varying the pulse duration irregularly, or (4) moving the signal source or imparting apparent motion to the signal source [88]. Thus, it may be instructive to revisit ultrasonic treatments in the future for crops where sound shadows can be minimized.

It is important to note also that capability to detect ultrasound to ultrasonic signals has evolved primarily in insects that are preyed upon by bats. Consequently, it is not surprising that none of the popularly marketed ultrasonic repellents has ever been shown to be cost-effective against insects such as cockroaches, mosquitoes, fleas and dragonflies that typically are not bat prey [89–92].

Potential applications of acoustic or vibrational signals for trapping of hemipteran insects or behavioural manipulation of their communication [56, 93–96], as well as for repelling ants or otherwise interfering with their colony-maintenance activities [58] have been considered but not yet implemented in field environments. In addition, there is potential for behavioural manipulation of predator–prey interactions [97] and acoustic mimicry [98]. Finally, several studies have been conducted with insects that use both sound and pheromone during courtship [24, 25, 99–103], as well as on insects that exhibit increased sensitivity to acoustic or olfactory stimuli after pre-exposure to the alternate stimulus [104, 105].

The precise control of timing and frequency of signals enabled by modern computer technology has considerable potential to enhance the trapping efficiency and effectiveness of behavioural manipulations in the future. Precise control of timing can be of value in the disruption of vibrational communication of duetting insects [106], especially pests such as *Diaphorina citri* (Kuwayama), the Asian citrus psyllid [107]. Similarly, there is potential that dynamic control of broadcast frequency may enhance capture efficiency of mosquitoes [49–54].

Conclusions

Until now, acoustic technology has an uneven record of success in insect pest management applications. The range

of acoustic stimuli is limited in comparison with the transmission distances of pheromones and other chemical attractants. The responses to attractive or repellent acoustic stimuli habituate rapidly and are variable among different segments of a given target species.

As with pheromone technology, however, acoustic methods have proven useful in a number of insect management applications, particularly for trapping of mosquitoes and midges, and enterprising entomologists are likely to develop additional beneficial uses of acoustic technology for insect management in the next few decades. Certainly, acoustic and vibrational signals can serve both long- and short-range functions, and the signals can be patterned easily to transmit several different signals using the same signalling organs and receptors, as has been demonstrated by the observed diversity of cricket acoustical communication [108]. It has been proposed that a diversity of airborne sound signals [46] and structural vibration signals [60, 109] remain to be discovered in insects. Modern digital signal processing technology now enables broadcast of a diversity of sonic and vibrational signal frequencies of different temporal patterns for a wide range of yet undiscovered, customizable insect behavioural management opportunities.

Acknowledgments

The use of trade, firm or corporation names in this publication is for the information and convenience of the reader. Such use does not constitute an official endorsement or approval by the USDA of any product or service to the exclusion of others that may be suitable. The USDA is an equal opportunity employer.

References

1. Mankin RW, Hagstrum DW, Smith MT, Roda AL, Kairo MTK. Perspective and promise: a century of insect acoustic detection and monitoring. *American Entomologist* 2011;57:30–44.
2. Alexander RD. Acoustical communication in arthropods. *Annual Review of Entomology* 1967;12:495–526.
3. Gwynne DT. Phylogeny of the Ensifera (Orthoptera): a hypothesis supporting multiple origins of acoustical signalling, complex spermatophores and maternal care in crickets, katydids, and weta. *Journal of Orthoptera Research* 1995;4:203–18.
4. Sueur J. Insect species and their songs. In: Drosopoulos S, Claridge MF, editors. *Insect Sounds and Communication: Physiology, Behaviour, Ecology, and Evolution*. Taylor & Francis, Boca Raton, FL; 2006. p. 207–17.
5. Connor WE, Corcoran AJ. Sound strategies: the 65-million-year-old battle between bats and insects. *Annual Review of Entomology* 2012;57:21–39.
6. Walker TJ. Acoustic traps for agriculturally important insects. *Florida Entomologist* 1988;71:484–92.

4 Perspectives in Agriculture, Veterinary Science, Nutrition and Natural Resources

7. Walker TJ. Acoustic methods of monitoring and manipulating insect pests and their natural enemies. In: Pest Management in the Subtropics: Integrated Pest Management – A Florida Perspective. Intercept Ltd, Andover, Hants, UK; 1996. p. 245–57.
8. Samarra FIP, Klappert K, Brumm H, Miller PJO. Background noise constrains communication: acoustic masking of courtship song in the fruit fly *Drosophila montana*. Behaviour 2009;146:1635–48.
9. Čokl A, Millar JG. Manipulation of insect signaling for monitoring and control of pest insects. In: Ishaaya I, Horowitz AR, editors. Biorational Control of Arthropod Pests. Springer Science + Business Media BV, New York; 2009. p. 279–316.
10. Kahn MC, Offenhauser W. The first field tests of recorded mosquito sounds used for mosquito destruction. American Journal of Tropical Medicine 1949;29:811–25.
11. Offenhauser WH, Kahn MC. The sounds of disease-carrying mosquitoes. Journal of the Acoustical Society of America 1949;21:259–63.
12. Ikeshoji T, Sakakibara M, Reisen WK. Removal sampling of male mosquitoes from field populations by sound-trapping. Japanese Journal of Sanitary Zoology 1985;36:197–203.
13. Ikeshoji T. Distribution of the mosquitoes, *Culex tritaeniorhynchus*, in relation to disposition of sound traps in a paddy field. Japanese Journal of Sanitary Zoology 1986;37:153–9.
14. Ikeshoji T, Ogawa K. Field catching of mosquitoes with various types of sound traps. Japanese Journal of Sanitary Zoology 1988;39:119–23.
15. Belton P. Attraction of male mosquitoes to sound. Journal of the American Mosquito Control Association 1994;10:297–301.
16. Silver JB. Sound traps and other miscellaneous attraction traps. In: Mosquito Ecology: Field Sampling Methods. Springer Netherlands, New York; 2007. p. 1027–48.
17. Ogawa K. Field trapping of male midge *Rheotanytarsus kyotoensis* (Diptera: Chironomidae) by sounds. Japanese Journal of Sanitary Zoology 1992;43:77–80.
18. Ogawa K, Sato H. Relationship between male acoustic response and female wingbeat frequency in a chironomid midge, *Chironomus yoshimatsui* (Diptera: Chironomidae). Japanese Journal of Sanitary Zoology 1993;44:355–60.
19. Hirabayashi K, Ogawa K. Field study on capturing midges, *Prosilocerus akamusi* (Diptera: Chironomidae), by artificial wingbeat sounds in a hyper-eutrophic lake. Medical Entomology and Zoology 2000;51:235–42.
20. Hirabayashi K, Nakamoto N. Field study on acoustic response of chironomid midges (Diptera: Chironomidae) around a hyper-eutrophic lake in Japan. Annals of the Entomological Society of America 2001;94:123–8.
21. Ulagaraj SM, Walker TJ. Phonotaxis of crickets in flight: attraction of male and female crickets to male calling songs. Science 1973;182:1278–9.
22. Campbell DJ, Shipp E. Spectral analysis of cyclic behaviour with examples from the field cricket *Teleogryllus commodus* (Walk.). Animal Behaviour 1974;22:862–75.
23. Walker TJ. Monitoring the flights of field crickets (*Gryllus* spp.) and a tachinid fly (*Euphasiopteryx ochracea*) in North Florida. Florida Entomologist 1986;69:678–85.
24. Spangler HG. Attraction of female lesser wax moths (Lepidoptera: Pyralidae) to male-produced and artificial sounds. Journal of Economic Entomology 1984;77:346–9.
25. Spangler HG. Ultrasonic communication in *Corcyra cephalonica* (Stainton) (Lepidoptera: Pyralidae). Journal of Stored Product Research 1987;23:203–11.
26. Mistal C, Takács S, Gries G. Evidence for sonic communication in the German cockroach (Diptera: Blattellidae). Canadian Entomologist 2000;132:867–76.
27. Webb JC, Burk T, Sivinski J. Attraction of female Caribbean fruit flies, *Anastrepha suspensa* (Diptera: Tephritidae) to the presence of males and male-produced stimuli in field cages. Annals of the Entomological Society of America 1983;76:507–17.
28. Mankin RW, Anderson JB, Mizrach A, Epsky ND, Shuman D, Heath RR, et al. Broadcasts of wing-fanning vibrations recorded from calling male *Ceratitis capitata* (Wiedemann) (Diptera: Tephritidae) increase captures of females in traps. Journal of Economic Entomology 2004;97:1299–309.
29. Mizrach A, Hetzroni A, Mazor M, Mankin RW, Ignat T, Grinshpun N, et al. Acoustic trap for female Mediterranean fruit flies. Transactions of the American Society of Agricultural Engineers 2005;48:2017–22.
30. Ikeshoji T, Yap HH. Monitoring and chemosterilization of a mosquito population, *Culex quinquefasciatus* (Diptera: Culicidae) by sound traps. Applied Entomology and Zoology 1987;22:474–81.
31. Ikeshoji T, Yap HH. Impact of the insecticide-treated sound traps on an *Aedes albopictus* population. Japanese Journal of Sanitary Zoology 1990;41:213–7.
32. Fowler HG. Traps for collecting live *Euphasiopteryx depleta* (Diptera: Tachinidae) at a sound source. Florida Entomologist 1988;71:654–6.
33. Paur J, Gray DA. Individual consistency, learning and memory in a parasitoid fly, *Ormia ochracea*. Animal Behaviour 2011;82:825–30.
34. Frank JH. Biological control of pest mole crickets. In: Rosen D, Bennett FD, Capinera JL, editors. Pest Management in the Subtropics: Biological Control – A Florida Perspective. Intercept, Andover, UK; 1994. p. 343–52.
35. Frank JH, Walker TJ. Permanent control of pest mole crickets (Orthoptera: Gryllotalpidae: *Scapteriscus*). American Entomologist 2006;52:138–44.
36. Riede K. Acoustic monitoring of Orthoptera and its potential for conservation. Journal of Insect Conservation 1998;2:217–23.
37. Chesmore ED, Ohya E. Automated identification of field-recorded songs of four British grasshoppers using bioacoustic signal recognition. Bulletin of Entomological Research 2004;94:319–30.
38. Forrest TG. Using insect sounds to estimate and monitor their populations. Florida Entomologist 1988;71:416–26.
39. Mankin RW. Acoustical detection of *Aedes taeniorhynchus* swarms and emergence exoduses in remote salt marshes. Journal of the American Mosquito Control Association 1994;10:302–8.

40. Raman DR, Gerhardt RR, Wilkerson JB. Detecting insect flight sounds in the field: implications for acoustical counting of mosquitoes. *Transactions of the American society of Agricultural and Biological Engineers* 2007;50:1481–5.
41. Cooley JR, Kritsky G, Edwards MJ, Zyla JD, Marshall DC, Hill KBR, *et al.* Periodical cicadas (*Magicicada* spp.): a GIS-based map of broods XIV in 2008 and 'XV' in 2009. *American Entomologist* 2011;57:144–50.
42. Cocquemot C, Lindelöw Å. Longhorn beetles (Coleoptera, Cerambycidae) Chapter 8.1. *BioRisk* 2010;4:193–218.
43. Rabitsch W. True bugs (Hemiptera, Heteroptera) Chapter 9.1. *BioRisk* 2010;4:407–33.
44. Invasive Species Compendium. 2011. Available from: URL: <http://www.cabi.org/isc/> (accessed 21 October 2011).
45. Stumpner A, von Helverson D. Evolution and function of auditory systems in insects. *Naturwissenschaften* 2001;88:159–70.
46. Drosopoulos S, Claridge MF. *Insect Sounds and Communication: Physiology, Behaviour, Ecology, and Evolution*. Taylor & Francis, Boca Raton, FL; 2006.
47. Göpfert MC, Briegel H, Robert D. Mosquito hearing: sound-induced antennal vibrations in male and female *Aedes aegypti*. *Journal of Experimental Biology* 1999;202:2727–38.
48. Göpfert MC, Robert D. Active auditory mechanics in mosquitoes. *Proceedings Royal Society of London B* 2001;268:333–9.
49. Cator LJ, Arthur BJ, Harrington LC, Hoy RR. Harmonic convergence in the love songs of the dengue vector mosquito. *Science* 2009;323:1077–9.
50. Warren B, Gibson G, Russell IJ. Sex recognition through midflight mating duets in *Culex* mosquitoes is mediated by acoustic distortion. *Current Biology* 2009;19:485–91.
51. Pennetier C, Warren B, Dabiré KR, Russell IJ, Gibson G. 'Singing on the wing' as a mechanism for species recognition in the malarial mosquito *Anopheles gambiae*. *Current Biology* 2010;20:131–6.
52. Cator LJ, Ng'Habi KR, Hoy RR, Harrington LC. Sizing up a mate: variation in production and response to acoustic signals in *Anopheles gambiae*. *Behavioral Ecology* 2010;21:1033–9.
53. Arthur BJ, Wyttenbach RA, Harrington LC, Hoy RR. Neural responses to one- and two-tone stimuli in the hearing organ of the dengue vector mosquito. *Journal of Experimental Biology* 2010;213:1376–85.
54. Cator LJ, Arthur BJ, Ponlawat A, Harrington LC. Behavioral observations and sound recordings of free-flight mating swarms of *Ae. aegypti* (Diptera: Culicidae) in Thailand. *Journal Medical Entomology* 2011;48:941–6.
55. Miles RN, Robert D, Hoy RR. Mechanically coupled ears for directional hearing in the parasitoid fly *Ormia ochracea*. *Journal of the Acoustical Society of America* 1995;98:3059–70.
56. Saxena KN, Kumar H. Interruption of acoustic communication and mating in a leafhopper and a planthopper by aerial sound vibrations picked up by plants. *Experientia* 1980;36:933–6.
57. Barbero F, Thomas JA, Bonelli S, Balletto SE, Schönrogge K. Queen ants make distinctive sounds that are mimicked by a butterfly social parasite. *Science* 2009;323:782–5.
58. Chiu YK, Mankin RW, Lin CC. Context-dependent stridulatory responses of *Leptogenys kitteli* (Hymenoptera: Formicidae) to social, prey, and disturbance stimuli. *Annals of the Entomological Society of America* 2011;104:1012–20.
59. Markl H, Hölldobler B. Recruitment and food-retrieving behavior in *Novomessor* (Formicidae, Hymenoptera): II. Vibration signals. *Behavioral Ecology and Sociobiology* 1978;4:183–216.
60. Coccroft RB, Rodríguez RL. The behavioral ecology of insect vibrational communication. *Bioscience* 2005;55:323–34.
61. Coccroft RB, Tieu TD, Hoy RR, Miles RN. Directionality in the mechanical response to substrate vibration in a treehopper (Hemiptera: Membracidae: *Umberia crassicornis*). *Journal of Comparative Physiology A* 2000;186:695–705.
62. Walker TJ, Forrest TG. Mole cricket phonotaxis: effects of intensity of synthetic calling song (Orthoptera: Gryllotalpidae: *Scapteriscus acletus*). *Florida Entomologist* 1989;72:655–9.
63. Alem S, Koselj K, Siemers BM, Greenfield MD. Bat predation and the evolution of leks in acoustic moths. *Behavioral Ecology and Sociobiology* 2011;65:2105–16.
64. Wishart G, Riordan DF. Flight responses to various sounds by adult males of *Aedes aegypti* (L.) (Diptera: Culicidae). *Canadian Entomologist* 1959;91:181–91.
65. Belton P. An analysis of direction finding in male mosquitoes. In Browne B, editor. *Experimental Analysis of Insect Behaviour*. Springer-Verlag, Berlin, Germany; 1974. p. 139–48.
66. Bidlingmayer WL. A comparison of trapping methods for adult mosquitoes: species response and environmental influence. *Journal of Medical Entomology* 1967;4:200–20.
67. Leemingsawat S, Kerdpibule V, Limswan S, Sucharit S, Ogawa K, Kanda T. Response of female mosquitoes of *Culex tritaeniorhynchus* to sound traps of various wingbeat frequencies with hamsters and dry ice. *Japanese Journal of Sanitary Zoology* 1988;39:67–70.
68. Nelson BS. Reliability of sound attenuation in Florida scrub habitat and behavioral implications. *Journal of the Acoustical Society of America* 2003;113:2901–11.
69. Hienton TE. Summary of Investigations of Electric Insect Traps. Agricultural Research Service, US Department of Agriculture Technical Bulletin 1498, Beltsville, MD; 1974. 136 pp.
70. Kanda T, Cheong WH, Loong KP, Lim TW, Ogawa K, Chiang GL, *et al.* Collection of male mosquitoes from field populations by sound trapping. *Tropical Biomedicine* 1987;4:161–6.
71. Ogawa K. Field study on acoustic trapping of *Mansonia* (Diptera: Culicidae) in Malaysia I. Mass-trapping of males by a cylindrical sound trap. *Applied Entomology and Zoology* 1988;23:265–72.
72. Kerdpibule V, Throngunkiant S, Leemingsawat S. Feasibility of wing beat sound trap for the control of mosquito vectors. *Southeast Asian Journal of Tropical Medicine and Public Health* 1989;20:639–41.
73. Walker TJ. Sound traps for sampling mole cricket flights (Orthoptera: Gryllotalpidae: *Scapteriscus*). *Florida Entomologist* 1982;65:105–10.
74. Walker TJ. A live trap for monitoring *Euphasiopteryx* and tests with *E. ochracea* (Diptera: Tachinidae). *Florida Entomologist* 1989;72:314–9.

6 Perspectives in Agriculture, Veterinary Science, Nutrition and Natural Resources

75. Campbell DJ. A new sound trap, and observations on the migratory and acoustic behavior of the field cricket *Teleogryllus commodus* (Walker). *Bioacoustics* 1990;2:199–208.
76. Chukanov VS, Lapshin DN. Acoustic behavior of mole crickets of *Gryllotalpa* genus. In: Gribakin FG, Weise K, Popov AV, editors. *Sensory Systems and Communication in Arthropods*. Birkhauser Verlag, Basel, Switzerland; 1990. p. 167–71.
77. Epsky NE, Morrill WL, Mankin RW. Traps for capturing insects. In: Capinera JL, editor. *Encyclopedia of Entomology*. Kluwer Academic Publishers, Dordrecht, The Netherlands; 2004. p. 2318–29.
78. Epsky NE, Morrill WL, Mankin RW. Traps for capturing insects. In: Capinera JL, editor. *Encyclopedia of Entomology*. Volume 4. 2nd edn. Springer, New York; 2008. p. 3887–901.
79. Hirabayashi K, Ogawa K. The efficiency of artificial wingbeat sounds for capturing midges in black light traps. *Entomologia Experimentalis et Applicata* 1999;92:233–8.
80. Bertram SM, Johnson LA, Clark J, Chief C. An electronic acoustic recorder for quantifying total signaling time, duration, rate and magnitude in acoustically signaling insects. *Technical Acoustics* 2004;20:1–8.
81. Mankin RW, Machan R, Jones R. Field testing of a prototype acoustic device for detection of Mediterranean fruit flies flying into a trap. In: *Proceedings of the 7th International Symposium on Fruit Flies of Economic Importance*, 10–15 September 2006, Salvador, Brazil; 2006. p. 165–9.
82. Mankin RW, Hodges RD, Nagle HT, Schal C, Pereira RM, Koehler PG. Acoustic indicators for targeted detection of stored product and urban insect pests by inexpensive infrared, acoustic, and vibrational detection of movement. *Journal of Economic Entomology* 2010;103:1636–46.
83. Forrest TG, Farris HE, Hoy RR. Ultrasound acoustic startle response in scarab beetles. *Journal of Experimental Biology* 1995;198:2593–8.
84. Payne TL, Shorey HH. Pulsed ultrasonic sound for control of oviposition by cabbage looper moths. *Journal of Economic Entomology* 1968;61:3–7.
85. Agee HR. Response of flying moths and other tympanate moths to pulsed ultrasound. *Annals of the Entomological Society of America* 1969;62:801–7.
86. Agee HR, Webb JC. Ultrasound for control of bollworms on cotton. *Journal of Economic Entomology* 1969;62:1322–26.
87. Agee HR, Webb JC. Effects of ultrasound on capture of *Heliothis zea* and *Ostrinia nubilalis* moths in traps equipped with ultraviolet lamps. *Annals of the Entomological Society of America* 1969;62:1248–52.
88. Agee HR. Response of bollworm moths to pulsed ultrasound while resting, feeding, courting, mating, and ovipositing. *Annals of the Entomological Society of America* 1969;62:1122–8.
89. Hinkle NC, Koehler PG, Patterson RS. Egg production, larval development and adult longevity of cat fleas (Siphonaptera: Pulicidae) exposed to ultrasound. *Journal of Economic Entomology* 1990;83:2306–9.
90. Coro F, Suarez S. Review and history of electronic mosquito repellents. *Wing Beats* 2000;11:6–7, 30–2.
91. Federal Trade Commission. FTC warns manufacturers and retailers of ultrasonic pest-control devices. 2001. Available from: URL: <http://www.ftc.gov/o0a/2001/05/fyi0128.shtm> (accessed 27 November 2011).
92. Leighton TG. What is ultrasound? *Progress in Biophysics and Molecular Biology* 2007;93:3–83.
93. Hunt RE, Morton TL. Regulation of chorusing in the vibrational communication system of the leafhopper *Graminella nigrifrons*. *American Zoologist* 2001;41:1222–8.
94. Polajnar J, Čokl A. The effect of vibratory disturbance on sexual behaviour of the southern green stink bug *Nezara viridula* (Heteroptera, Pentatomidae). *Central European Journal of Biology* 2008;3:189–97.
95. Mazzoni V, Lucchi A, Čokl A, Prešern J, Virant-Doberlet M. Disruption of the reproductive behaviour of *Scaphoideus titanus* by playback of vibrational signals. *Entomologia Experimentalis et Applicata* 2009;133:1–12.
96. Zhang Z, Fu Q, Chen W, Zhao G, Chen G. Effect of some factors on acoustic trapping for *Nilaparvata lugens* (Stal) Homoptera: Delphacidae. *Acta Entomologica Sinica* 1995;38:166–72.
97. Laumann RA, Blassiolo Moraes CM, Čokl A, Borges M. Eavesdropping on sexual vibratory signals of stink bugs (Hemiptera: Pentatomidae) by the egg parasitoid *Telenomus podisi*. *Animal Behaviour* 2007;73:637–49.
98. Barber JE, Connor WE. Acoustic mimicry in a predator-prey interaction. *Proceedings of the National Academy of Sciences of the USA* 2007;104:9931–4.
99. Trematerra P, Pavan G. Ultrasound production in the courtship behaviour of *Ephestia cautella* (Walk.), *E. kuehniella* Z. and *Plodia interpunctella* (Hb.) (Lepidoptera: Pyralidae). *Journal of Stored Product Research* 1995;31:45–8.
100. Millar JG, McBrien HL, Ho HY, Rice RE, Cullen E, Zalom FG, et al. Pentatomid bug pheromones in IPM: possible applications and limitations. *Bulletin of the International Organization of Biological Control, Pheromone Working Group* 2002;25:1–11.
101. Takács S, Mistal C, Gries G. Communication ecology of webbing clothes moth: attractiveness and characterization of male-produced sonic aggregation signals. *Journal of Applied Entomology* 2003;127:127–33.
102. Hart M. The role of sonic signals in the sexual communication of peach twig borers *Anarsia lineatella* Zeller (Lepidoptera: Gelechiidae) [MSc Thesis]. Simon Fraser University, Burnaby, British Columbia, Canada; 2006. 47 pp.
103. Nakano R, Takanashi T, Fujii T, Skals N, Surlykke A, Ishikawa Y. Moths are not silent, but whisper ultrasonic courtship songs. *Journal of Experimental Biology* 2009;212:4072–8.
104. Mankin RW, Petersson E, Epsky N, Heath RR, Sivinski J. Exposure to male pheromones enhances *Anastrepha suspensa* (Diptera: Tephritidae) female response to male calling song. *Florida Entomologist* 2000;83:411–21.
105. Anton S, Evengard K, Barrozo RB, Anderson P, Skals N. Brief predator sound exposure elicits behavioral and neuronal long-term sensitization in the olfactory system of an insect. *Proceedings of the National Academy of Sciences of the USA* 2011;108:3401–5.
106. Percy DM, Taylor GS, Kennedy M. Psyllid communication: acoustic diversity, mate recognition and phylogenetic signal. *Invertebrate Systematics* 2006;20:431–45.

107. Wenninger EJ, Hall DG, Mankin RW. Vibrational communication between the sexes in *Diaphorina citri* (Hemiptera: Psyllidae). *Annals of the Entomological Society of America* 2009;102:547–55.
108. Alexander RD. Evolutionary change in cricket acoustical communication. *Evolution* 1962;16:443–67.
109. Cocroft RB, Hamel JA. Vibrational communication in the 'other insect societies': a diversity of ecology signals, and signal functions. In: O'Connell-Rodwell CE, editor. *The Use of Vibrations in Communication: Properties, Mechanism and Function Across Taxa*. Transworld Research Network, Kerala, India; 2010. p. 47–68.