



## SPECIAL ISSUE: INSECTS IN AGROECOSYSTEMS

# Predator demographics and dispersal in alfalfa trap-cropped strawberry

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### Abstract

Alfalfa, *Medicago sativa* L. (Fabaceae), is a highly preferred host plant of *Lygus* spp. (Hemiptera: Miridae). As such, intercropping alfalfa trap-crops in strawberry production can serve as a sink for both *Lygus* (primarily *Lygus hesperus* Knight) and its natural enemies. Here we investigated the population dynamics and dispersal characteristics of the generalist predator complex in strawberry fields with alfalfa trap-crops spaced 50 rows (62 m) apart. Predator abundance was determined by counting six focal taxa collected from strawberry and alfalfa. The data revealed that *Orius* spp. (Hemiptera: Anthocoridae) were the numerically dominant predator taxa, comprising 84% of the focal predator population. In general, the population densities obtained for the various taxa throughout this agroecosystem were unexpectedly uniform. Predator movement from a central alfalfa trap-crop row was determined using a protein mark–capture procedure. Most protein-marked predator specimens were collected less than 2 m from the centrally marked alfalfa row, indicating that the trap-crop often produces a predator sink. Results suggest that alfalfa is a useful cultural (trap-cropping) and a biological (refuge for natural enemies) control tactic for managing *Lygus* spp. in strawberries.

### Introduction

California growers produce over 90% of the strawberries, *Fragaria ananassa* Duchesne (Rosaceae) marketed in the USA (Marzolo, 2015). The *Lygus* species complex – primarily *Lygus hesperus* Knight (Hemiptera: Miridae) – is considered a key pest of this industry (Strand, 2008). *Lygus* feeding on flowers and immature fruit produces distorted strawberries (Handley & Pollard, 1993). Such cosmetic damage reduces fresh market yield, which constitutes most of the economic value to California strawberry growers (CDFA, 2018).

*Lygus* spp. in California strawberry are commonly managed with broad-spectrum insecticides and tractor-

mounted vacuums. Culturally and biologically based management of this pest has been achieved through the use of alfalfa, *Medicago sativa* L. (Fabaceae), trap-crops. As *Lygus* have a reliable host plant preference for alfalfa (Stern et al., 1964; Mueller & Stern, 1974; Goodell, 2009), integrating managed trap-crops into organic strawberry production has been shown to aggregate *Lygus* populations (Swezey et al., 2013) and reduce associated fresh market yield losses (Swezey et al., 2007).

Trap-cropping also affects the biological control of *Lygus*, as spatially concentrated prey densities in alfalfa likely influence the behavior of predaceous insects and spiders. Specifically, how generalist predators respond to an aggregated distribution of *Lygus* can help determine whether alfalfa acts as a sink or a source for these predators relative to adjacent organic strawberries. The goals of this study were to document the population dynamics and dispersal characteristics of predaceous arthropods inhabiting an alfalfa trap-cropped strawberry agroecosystem. Here we use conventional sampling techniques, in combination with the protein mark–capture method, to investigate the

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spatial and dispersal patterns of the generalist predator complex to clarify potential sink–source dynamics.

## Materials and methods

### Study site

This study was conducted in 2008 and 2009 at two proximate organically certified strawberry ranches (40 and 20 ha) located near Prunedale, CA, USA. At both farms, alfalfa trap-crops were established on every 50th strawberry row, 62 m apart.

### Sampling and mark–capture procedures

The resident arthropod predators in a central alfalfa row were marked with a 12% (vol/vol) solution of chicken egg whites (All Whites; Papetti Foods, Elizabeth, NJ, USA) using a gas-powered backpack sprayer (Mist Duster MD155DX; Maruyama US, Denton, TX, USA) at a rate of 1 l per 6 m of row. The mark was applied to the centralized alfalfa row in each of four replicated plots on 19 and 25 August in 2008 and 2009, respectively. Arthropod predators were collected from the centrally marked alfalfa trap-crop, adjacent strawberry rows 1, 2, 3, and 10, and the neighboring (unmarked) alfalfa trap-crops 50 rows away, 1 and 2 days after the protein mark was applied. Strawberry rows were sampled using a hand-held vacuum suctioning modified reverse leaf blower (model BG75; Stihl, Virginia Beach, VA, USA) fitted with a 13-cm-diameter netted intake orifice. Predator collections from trap-crops were made using a standard 30-cm-diameter sweep net. In each case, a single sample consisted of either 200 suctionings or 200 sweeps per respective strawberry or alfalfa row.

### Protein mark detection

Each field-collected predator was placed into a 1.6-ml microcentrifuge tube (VWR, Atlanta, GA, USA) and examined for the presence of the protein mark by the anti-chicken egg albumin enzyme-linked immunosorbent assay (ELISA) described by Hagler et al. (2014). Any predator testing positive for the presence of the mark was assumed to have originated from the centrally located, protein-marked, alfalfa trap-crop. Predators collected from the mark–capture study site were scored positive for the presence of the protein mark if their ELISA values were six standard deviations above the negative control mean value (Hagler, 2011). It should be noted that the negative control predator specimens were collected from nearby alfalfa fields prior to the application of the protein marks. Additional information regarding the farm sites, protein marking protocols, collection methods, or ELISA techniques used in this study are provided by Swezey et al. (2013, 2014).

### Data presentation

The predator taxa collected from each designated sampling row in each replicated plot were pooled over the two sample dates each year. The total number of individuals collected, along with the total number of predators containing the protein mark, were tallied for each predator taxon. Only the adult stage was counted for the insect predators. However, the juvenile and adult stages of spiders and harvestmen were counted. The *Nabis* spp. and *Geocoris* spp. counts were also pooled and classified as predaceous bugs, due to the small counts obtained for both species. The average distance that each predator taxon traveled from the centrally marked alfalfa trap-crop was determined by dividing the total distance traveled by protein-marked specimens by the total number of protein-marked specimens collected. The distance from the marked alfalfa to the unmarked strawberry rows (rows 1, 2, 3, and 10 in each direction) and neighboring trap-crops 50 rows away in each direction were based on global positioning system (GPS) coordinates.

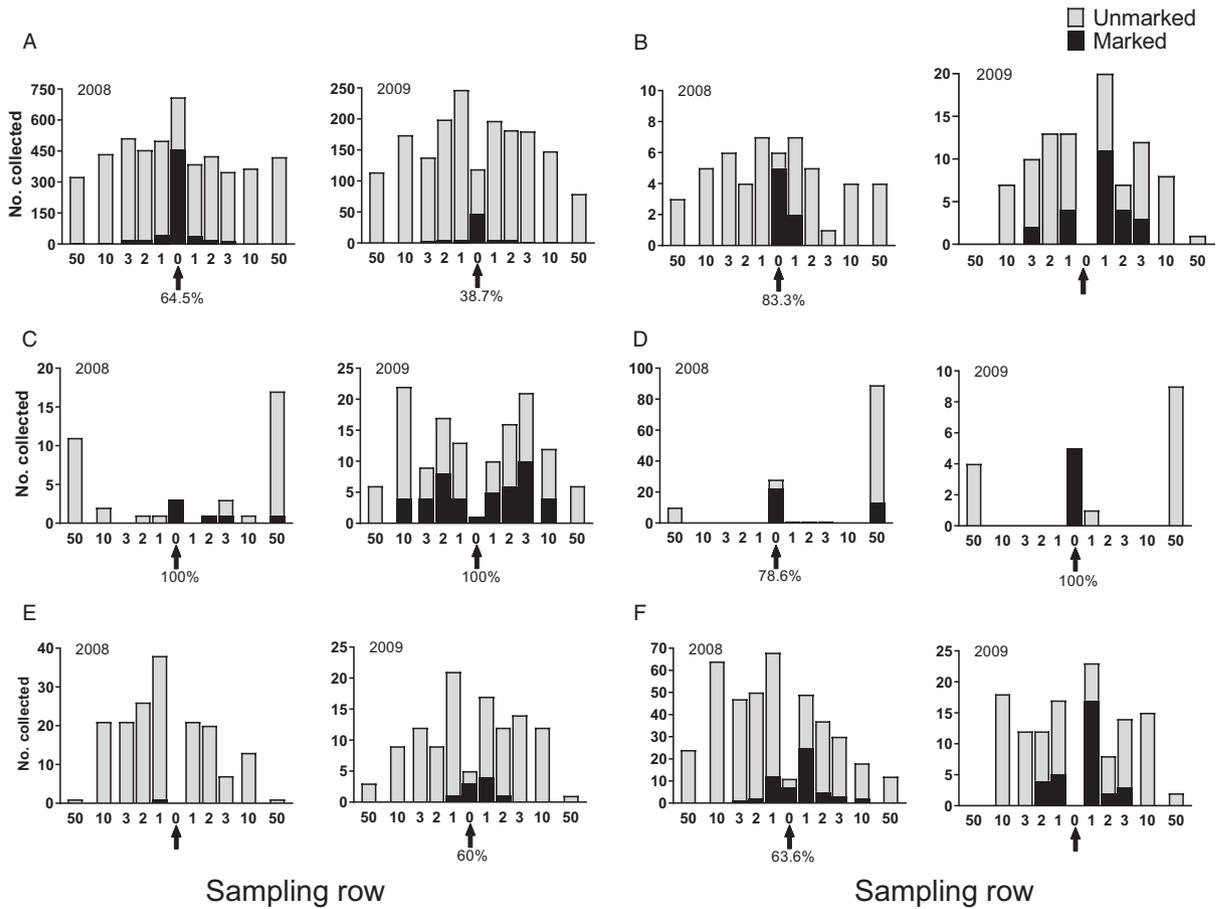
## Results

### *Orius*

The *Orius* spp. complex (Hemiptera: Anthocoridae), composed almost entirely of *Orius tristicolor* (white), were the most abundant predator taxa encountered. There were over 2.5× as many *Orius* adults collected in 2008 ( $n = 4\,888$ ) than in 2009 ( $n = 1\,777$ ). The *Orius* populations were uniformly distributed throughout the study site each year (Figure 1A). The marking efficiency for *Orius* (i.e., the percentage of marked individuals collected from the central-marked alfalfa trap-crop) was 64.5% for 2008 (458 out of 710 individuals) and 38.7% for 2009 (46 out of 119) (Figure 1A). Most (73%) of the protein-marked *Orius* were collected from the centrally marked row of alfalfa. All remaining protein-marked specimens were captured within three strawberry rows of the centrally marked trap-crop. As such, the average dispersal distance for protein-marked *Orius* was only 0.92 m in 2008 and 0.87 m in 2009.

### Other predaceous bugs

Adult *Nabis* spp. – primarily *Nabis alternatus* Parshley (Hemiptera: Nabidae) – and *Geocoris* spp. – primarily *Geocoris punctipes* (Say) (Hemiptera: Geocoridae) – were the next most abundant true bug predator taxa encountered at the study sites, albeit at relatively low densities. These true bugs were about twice as dense in 2009 ( $n = 91$ ) as in 2008 ( $n = 52$ ). The marking efficiency for these true bugs was 83.3% in 2008 (Figure 1B). However, it should be



**Figure 1** Total and protein-marked (black bars) predators collected in 2008 and 2009 over two consecutive days (1 and 2 days after marking) in each designated sampling row in a strawberry production field: (A) *Orius* spp., (B) other predaceous bugs, (C) lacewings, (D) lady beetles, (E) spiders, and (F) harvestmen. The black arrows signify the centrally marked alfalfa trap-crop row. The percentages below the arrows signify the protein mark efficiency for each predator taxon in the trap-crop. Note the differences in scales on the vertical axes.

stipulated that this outcome was derived from a small sample size (i.e., five of the six samples were marked). There were no marking efficiency data available in 2009 due to a lack of collected true bugs from the centrally marked trap-crop. Nevertheless, the relatively high proportion of these protein-marked true bugs collected from the nearby strawberry rows suggests that we had a well-marked population of true bugs originating from the protein-marked trap-crop. The average dispersal distance, based on small sample sizes, was only 0.35 m in 2008 and 1.96 m in 2009.

**Lacewings**

The adult lacewings (Neuroptera) collected at the study sites included both green [*Chrysoperla carnea* (Stephens) s.l., Chrysopidae] and brown lacewings (*Hemerobius* spp., Hemerobiidae). Total lacewing abundance was over 3× greater in 2009 (n = 133) than in 2008 (n = 40) (Figure 1C). In 2008, *C. carnea* comprised about 90% of

collected lacewings. Conversely, in 2009 *Hemerobius* spp. made up about 86% of the lacewing community. Regarding collection location, lacewings (mostly *C. carnea*) were most prevalent in the alfalfa trap-crop rows in 2008 (Figure 1C). Yet in 2009, lacewings (mostly *Hemerobius* spp.) were more prevalent and uniformly distributed throughout the strawberry rows. The marking efficiency for lacewings, albeit based on an extremely small sample size (n = 4 for both years combined), was 100%. Only two protein-marked lacewings were captured beyond the centrally marked alfalfa in 2008. Of these, one had dispersed 62 m into an adjacent alfalfa trap-crop. As such, the calculated average movement of protein-marked lacewings (n = 6) in 2008 was 11.4 m. In 2009, there were more of both unmarked and marked lacewings that were uniformly collected in all the strawberry rows. The average dispersal distance for lacewings in 2009 was 4.2 m from the centrally marked alfalfa row.

### Lady beetles

The lady beetle species complex (Coleoptera: Coccinellidae) inhabiting the trap-cropped strawberry farms consisted primarily of *Hippodamia convergens* Guérin-Méneville (83% of coccinellids). The remaining lady beetle taxa consisted of *Coccinella californica* (Mannerheim) (12.3%) and *Coccinella septempunctata* L. (4.7%). The beetle population was 6.5× higher in 2008 (n = 130 specimens collected) than in 2009 (n = 20). Almost all the specimens were collected from the alfalfa trap-crop rows (Figure 1D). The efficiency of the marking procedure was 78.6% in 2008 (n = 22 of the 28) and 100% in 2009 (n = 5). All protein-marked beetles were captured in trap-crops. In 2008, 13 of the 89 beetles (14.6%) collected from the eastern-most alfalfa trap-crop contained the protein mark. This yielded a relatively long average dispersal distance of 23.2 m. We suspect this unidirectional movement could be attributed to the prevailing wind patterns typically exhibited at the study sites. Conversely, protein-marked lady beetle specimens were only from the centrally marked alfalfa trap-crop row in 2009, and therefore had an average dispersal of 0 m.

### Spiders

Collected spiders (Araneae) included various species within the families Lycosidae (60.5% of the arachnids), Salticidae (22.2%), and Thomisidae (17.3%). These hunting spiders were about 1.5× more abundant in 2008 (n = 169) than in 2009 (n = 115). Spider abundance was much higher in the strawberry rows (Figure 1E), as no spiders were collected in the centrally marked alfalfa in 2008, and only five were captured in 2009. Of these five, three (60% marking efficiency) contained the protein mark. Overall, only 10 protein-marked spiders were collected in 2008 and 2009. All 10 were captured within two rows of the centrally marked alfalfa. The average dispersal distance rate of the protein-marked spiders, albeit based on small sample sizes, was 1.4 m in 2008 (n = 1) and 0.96 m (n = 9) in 2009.

### Harvestmen

Harvestmen (Opiliones: *Phalangium*) were present in relatively high numbers. There were over 3× as many collected (n = 410) in 2008 than in 2009 (n = 121). Harvestmen were more abundant in the strawberries than in the alfalfa (Figure 1F). Marking efficiency data for the 11 captured harvestmen in 2008 was 63.6% (n = 7). No harvestmen were collected in 2009 from the centrally marked alfalfa. As protein-marked harvestmen were frequently collected in the three strawberry rows nearest to the protein-marked alfalfa both years of the study, their average dispersal

distance beyond the centrally marked trap-crop was 1.9 m in 2008 and 1.8 m in 2009.

## Discussion

Predator census data for both the alfalfa trap-cropped strawberries reported here and by Hagler et al. (2018a) showed that *Orius* was the most numerically dominant predator taxon, comprising 86 and 79% of the focal arthropod fauna reported in 2008 and 2009, respectively. *Orius*, along with the other taxa collected in this study (e.g., heteropterans, coccinellids, and spiders), have been documented consuming *Lygus* in cotton (Hagler, 2011) and/or strawberry (Hagler et al., 2018a) by predator gut analyses. As generalist predators are capable of regulating *Lygus* populations in agroecosystems (Zink & Rosenheim, 2008), developing strategies that maximize predation services should be prioritized. For instance, gaining a better understanding of how habitats with high prey densities (i.e., trap-crops) affect predator movement can clarify predator source/sink dynamics and thereby inform conservation biological control strategies (Lavandero et al., 2004).

The protein mark-capture technique proved to be useful for uniquely tagging the predators inhabiting the central alfalfa rows. Overall, 62% (n = 550 out of 888) of all the focal predators captured from the centrally marked alfalfa contained the protein mark. Moreover, the protein mark-capture data revealed that relatively few predators captured beyond the centrally marked alfalfa trap-crop (n = 361 out of 7 058) were marked (5.1%). As such, the average dispersal of protein-marked arthropods within 48 h was less than 2.0 m, which equates to an average of about 1.5 rows from the centrally marked trap-crop. This low average dispersal rate suggests that the alfalfa trap-crop retains the predators within, or adjacent to, the trap-crop. These data are similar to the dispersal results reported for *Lygus* and the *Lygus*-specific parasitoid, *Peristenus relictus* (Ruthe), by Swezey et al. (2013, 2014).

More specifically, *Orius* were largely retained by alfalfa, as roughly 75% of protein-marked minute pirate bugs were collected from the central trap-crop, whereas the remaining 25% were collected within three adjacent strawberry rows on either side of the marked alfalfa. As a result, the average dispersal of protein-marked *Orius* was less than one row from the central trap-crop. *Orius* feed on *Lygus* eggs (Hagler et al., 1992) and small nymphs (Zink & Rosenheim, 2008), particularly in alfalfa trap-crops (Hagler et al., 2018a). Additional feeding opportunities (e.g., pollen, thrips, aphids, etc.) further bolster the prey reservoir provided by alfalfa, which,

in turn, creates a sink effect for *Orius* within strawberry fields.

In this study, alfalfa also created a strong sink for coccinellids, as 97% ( $n = 145$  out of 150) of all the lady beetles collected over the course of the study were obtained from the alfalfa trap-crop rows. *Hippodamia convergens* has been shown to consume *Lygus* nymphs in cotton (Hagler, 2011), and it is possible that beetle retention in alfalfa was influenced by the high *Lygus* prey densities found in trap-crops. However, in this case, it seems more likely that coccinellid fidelity to alfalfa was instead driven by the moderately sized aphid populations (e.g., *Acyrtosiphon pisum* Harris) that were found in trap-crops, but not strawberry. The converse can also occur, however, where high aphid densities in strawberry (e.g., *Chaetosiphon fragaefolii* Huber) can produce a robust response by coccinellids in strawberry, but not alfalfa (DJ Nieto, pers. obs.). Bastola et al. (2014) similarly reported that although alfalfa acted as a sink for *H. convergens* (relative to cotton), bidirectional lady beetle movement between these two crops was common and influenced by relative prey availability.

Conversely, alfalfa acted as a source of some predators during this study that dispersed from trap-crops, into neighboring strawberry rows. For example, protein-marked *Hemerobius* sp. were captured from every sampled strawberry row in 2009, including row 10 to the east and west of the central trap-crop. This may have been in response to relatively high *Lygus* nymphal densities in associated strawberry rows and increased management (i.e., vacuuming) frequency in the alfalfa trap-crop. *Hemerobius* sp. are commonly recognized as predators of hemipterans, such as aphids (Neuenschwander & Hagen, 1980) and psyllids (Nickel et al., 1965). However, as part of the predation study conducted by Hagler et al. (2018a), field-collected *Hemerobius* sp. were shown to feed on *Lygus* as well (DJ Nieto, unpubl.). Specifically, 33.3 and 4.5% of assayed *Hemerobius* sp. collected from alfalfa ( $n = 9$ ) and strawberry ( $n = 44$ ) contained *Lygus* remains.

For other predators, the sink/source dynamics of trap-cropping were ambiguous, as differences in predator counts (abundance) between the alfalfa trap and strawberry rows were generally unremarkable and predator movement patterns were counterintuitive. For example, in 2009, protein-marked harvestman and *Geocoris*-plus-*Nabis* were collected from five of six adjacent strawberry rows; yet, curiously, none were recovered from the marked trap-crop.

A reassessment of our sampling methodology may clarify these discrepancies: As discussed in detail by Hagler et al. (2018b), differences in sampling efficacy between alfalfa (sweep netting) and strawberry (hand-

held vacuum suctioning) likely obscured some predator-dispersal behavioral patterns pertaining to trap-cropping. For instance, the majority of a strawberry plant's vertical canopy is sampled using the hand-held vacuum device. Conversely, only the upper third of the alfalfa canopy is sampled with the sweep net. Sweeping therefore likely led to underestimated predator abundance data for those taxa that are evenly distributed throughout the alfalfa canopy. Furthermore, sweep netting would severely underestimate predator abundance of taxa (i.e., some true bug and spider taxa) that preferentially inhabit the lower portion of the alfalfa canopy (Crocker & Whitcomb, 1980; Nyffeler & Benz, 1988; Fasola & Mogavero, 1995). If predator counts in alfalfa were in fact underestimated, then potential alfalfa sink effects would be masked and, instead, could be falsely interpreted as either having no discernable effect on predator movement or as constituting a source of predators into strawberry. Future studies that address potential sampling biases based on the vertical distribution of predators within the alfalfa canopy (e.g., whole plant sampling) are therefore needed.

In summary, a predator population census is discussed and included six primary taxa. Overall, *Orius* was the numerically dominant predator taxon encountered. The dispersal pattern of the predator complex inhabiting alfalfa trap-cropped strawberry fields is also described. The protein mark-capture method effectively marked over 60% of the resident arthropod fauna inhabiting centrally located alfalfa trap-crops. The recapture of protein-marked predators beyond the centrally marked alfalfa trap-crop was rare. When discernable, these data suggest that the predator population was largely retained in the alfalfa trap-crop. It is likely that the sinking of predators into the preferred host plant improves the biological control services rendered by the *Lygus* natural enemy complex.

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## References

- Bastola A, Parajulee MN, Porter RP, Shrestha RB, Chen FJ & Carroll SC (2014) Intercrop movement of convergent lady beetle, *Hippodamia convergens* (Coleoptera: Coccinellidae), between adjacent cotton and alfalfa. *Insect Science* 23: 145–156.
- CDFA (California Department of Food and Agriculture) (2018) California Agricultural Statistics Review, 2017–2018. <https://www.cdffa.ca.gov/statistics/PDFs/2017-18AgReport.pdf> (accessed on 7 March 2019).
- Crocker RL & Whitcomb WH (1980) Feeding niches of the big-eyed bugs *Geocoris bullatus*, *G. punctipes*, and *G. uliginosus* (Hemiptera: Lygaeidae: Geocorinae). *Environmental Entomology* 9: 508–513.
- Fasola M & Mogavero F (1995) Structure and habitat use in a web-building spider community in northern Italy. *Bollettino Zoologica* 62: 159–166.
- Goodell P (2009) Fifty years of the integrated control concept: the role of landscape ecology in IPM in San Joaquin valley cotton. *Pesticide Management Science* 65: 1293–1297.
- Hagler JR (2011) An immunological approach to quantify consumption of protein-tagged *Lygus hesperus* by the entire cotton predator assemblage. *Biological Control* 58: 337–345.
- Hagler JR, Cohen AC, Bradley-Dunlop D & Enriquez FJ (1992) Field evaluation of predation on *Lygus hesperus* using a species- and stage-specific monoclonal antibody. *Environmental Entomology* 21: 896–900.
- Hagler JR, Naranjo SE, Machtley SA & Blackmer F (2014) Development of a standardized protein immunomarking protocol for insect mark-capture research. *Journal of Applied Entomology* 138: 772–782.
- Hagler JR, Nieto DJ, Machtley SA, Spurgeon DW, Hogg BN & Swezey SL (2018a) Dynamics of predation on *Lygus hesperus* (Hemiptera: Miridae) in alfalfa trap-cropped organic strawberry. *Journal of Insect Science* 18(4): 12.
- Hagler JR, Thompson AL, Stefanek MA & Machtley SA (2018b) Use of body-mounted cameras to enhance data collection: an evaluation of two arthropod sampling techniques. *Journal of Insect Science* 8(2): 40.
- Handley DT & Pollard JE (1993) Microscopic examination of tarnished plant bug (Heteroptera: Miridae) feeding damage to strawberry. *Journal of Economic Entomology* 86: 505–510.
- Lavandero B, Wratten S, Hagler J & Jervis M (2004) The need for effective marking and tracking techniques for monitoring the movements of insect predators and parasitoids. *International Journal of Pest Management* 50: 147–151.
- Marzolo G (2015) Agricultural Marketing Resource Center: Strawberries. <https://www.agmrc.org/commodities-products/fruits/strawberries/> (accessed on 1 February 2019).
- Mueller AJ & Stern VM (1974) Timing of pesticide treatment on safflower to prevent *Lygus* from dispersing to cotton. *Journal of Economic Entomology* 67: 77–80.
- Neuenschwander P & Hagen KS (1980) Role of the predator *Hemerobius pacificus* in a non-insecticide treated artichoke field. *Environmental Entomology* 9: 492–495.
- Nickel JL, Shimizu JT & Wong TT (1965) Studies on natural control of pear psylla in California. *Journal of Economic Entomology* 58: 970–976.
- Nyffeler M & Benz G (1988) Prey analysis of the spider *Achaearanea riparia* (Blackw.) (Araneae, Theridiidae), a generalist predator in winter wheat fields. *Journal of Applied Entomology* 106: 425–431.
- Stern VM, van den Bosch R & Leigh TF (1964) Strip cutting alfalfa for *Lygus* bug control. *California Agriculture* 4: 4–6.
- Strand LL (2008) Integrated Pest Management for Strawberries, 2nd edn. Publication 3351, University of California Agriculture and Natural Resources, Oakland, CA, USA.
- Swezey SL, Nieto DJ & Bryer JA (2007) Control of western tarnished plant bug *Lygus hesperus* Knight (Hemiptera: Miridae) in California organic strawberries using alfalfa trap crops and tractor-mounted vacuums. *Environmental Entomology* 36: 1457–1465.
- Swezey SL, Nieto DJ, Hagler JR, Pickett CH, Bryer JA & Machtley SA (2013) Dispersion, distribution and movement of *Lygus* spp. (Hemiptera: Miridae) in trap-cropped strawberries. *Environmental Entomology* 42: 770–778.
- Swezey SL, Nieto DJ, Pickett CH, Hagler JR, Bryer JA & Machtley SA (2014) Spatial density and movement of the *Lygus* spp. parasitoid *Peristenus relictus* (Hymenoptera: Braconidae) in organic strawberries with alfalfa trap crops. *Environmental Entomology* 43: 363–369.
- Zink AG & Rosenheim JA (2008) Stage-specific predation on *Lygus hesperus* affects its population stage structure. *Entomologia Experimentalis et Applicata* 126: 61–66.