

RESEARCH

Elevated Carbon Dioxide and Ozone Effects on Peanut: II. Seed Yield and Quality

Kent O. Burkey,^{*} Fitzgerald L. Booker, Walter A. Pursley, and Allen S. Heagle

ABSTRACT

Many adverse effects of tropospheric O₃ on C₃ crop plants are ameliorated by elevated concentrations of atmospheric CO₂, but the extent of the interaction can vary, depending on the species, gas concentrations, and other experimental conditions. A 2-yr open-top field chamber experiment was conducted to examine this interaction in peanut (*Arachis hypogaea* L.) by testing the effects of O₃ and CO₂ mixtures on yield and seed quality. Treatments were ambient CO₂ (375 μmol mol⁻¹) and CO₂ additions of approximately 173 and 355 μmol mol⁻¹ in combination with charcoal-filtered (CF) air (22 nmol O₃ mol⁻¹), nonfiltered (NF) air (46 nmol O₃ mol⁻¹), and NF air plus O₃ (75 nmol O₃ mol⁻¹). At ambient CO₂, pod number was suppressed 16% in NF air and 44% in elevated O₃. Pod and seed mass were not significantly affected in NF air but were lowered 33 to 37% in elevated O₃. Elevated CO₂ increased yield parameters 7 to 17% for plants grown in CF air and restored yield in NF air and elevated O₃ treatments to control or higher levels. Gas treatment effects on peanut market grade characteristics were small. No treatment effects were observed on the protein and oil contents of seeds, but there were changes in fatty acid composition. Overall results indicate that increasing concentrations of tropospheric O₃ will suppress yield of O₃-sensitive peanut cultivars, while elevated CO₂ will moderate this response. Elevated O₃ and CO₂ are not expected to have major effects on peanut seed composition and quality.

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Abbreviations: AA, ambient air; CF, charcoal-filtered air; ELK, extra large kernels; NF, nonfiltered air; OZ, 1.56 x ambient O₃; TSMK, total sound mature kernels.

RIISING CONCENTRATIONS of atmospheric CO₂ are predicted to increase biomass production and yield of many C₃ crops (Ainsworth and Long, 2005; Jablonski et al., 2002; Kimball et al., 2002). It is expected, however, that the magnitude of these potential gains from CO₂ enrichment will be influenced by possible changes in other environmental factors such as temperature and soil water availability (Ainsworth and Long, 2005; Prasad et al., 2005). Air pollutants, most notably O₃, also influence the effect of elevated CO₂ on crop growth and yield, and vice versa (Allen, 1990; Barnes and Wellburn, 1998; Fiscus et al., 2002; Olszyk et al., 2000). Current tropospheric O₃ levels suppress crop growth and yield in many regions worldwide, and emissions of O₃ precursors and areas affected by O₃ pollution are anticipated to increase (Ashmore, 2005; Dentener et al., 2005; Fiscus et al., 2005; Fuhrer and Booker, 2003; Houghton et al., 2001; Morgan et al., 2003; Prather et al., 2003). Elevated CO₂ concentrations tend to counteract O₃ effects on plant growth and yield, but O₃ can also diminish enhancements in these parameters due to elevated CO₂ (Barnes and Wellburn, 1998; Fiscus et al., 2002; Olszyk et al., 2000). The nature of the interaction depends on the sensitivity of the crop, the gas concentrations, and the influences of other biotic and environmental factors.

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Peanut (*Arachis hypogaea* L.) production is vulnerable to current and predicted higher levels of O₃ pollution in the future. Some peanut cultivars exhibit relatively high sensitivity to O₃ (Ensing et al., 1985, 1986; Heagle, 1989; Heagle et al., 1983). For example, regression modeling based on open-top field chamber experiments indicated that yield of an O₃-sensitive peanut line (NC-6) was suppressed 7 to 14% by ambient O₃ levels (52–56 nmol mol⁻¹, 7 h daily average) relative to control treatments (25–26 nmol mol⁻¹), and even more so by higher O₃ concentrations (Heagle, 1989). Climate model projections forecast that the largest peanut producing regions in the world, located mainly in eastern China, central India, central Africa, the southern United States, and Indonesia (Rhoades and Nazarea, 2003), may experience significantly higher levels of tropospheric O₃ in the coming 50 yr (Dentener et al., 2005; Prather et al., 2003; Wang and Mauzerall, 2004). Rising levels of atmospheric CO₂ will likely moderate the effects of increasing ground-level O₃ concentrations in these regions, but eventual effects on yield in concert with other changing environmental factors are unclear.

Ozone suppresses plant growth and yield in large part by inhibiting net photosynthesis and possibly translocation processes, thus limiting photosynthate availability (Fiscus et al., 2005; Long and Naidu, 2002; Pell et al., 1997; Runeckles and Chevone, 1992). Increases in maintenance respiration and detoxification processes might curtail growth as well (Amthor, 1988). In addition, detrimental effects of O₃ on pollen germination and tube growth, fertilization, and abscission rates of flowers, pods, and seeds can contribute to suppressed yield in some crops (Ashmore, 2005; Black et al., 2000; Runeckles and Chevone, 1992).

Atmospheric CO₂ and O₃ co-occur in the atmosphere. Studies have shown that elevated CO₂ ameliorates the suppressive effect of O₃ on yield in a number of crop species, including cotton (*Gossypium hirsutum* L.), potato (*Solanum tuberosum* L.), rice (*Oryza sativa* L.), soybean [*Glycine max* (L.) Merr.], and wheat (*Triticum aestivum* L.) (Booker and Fiscus, 2005; Booker et al., 2005; Craigon et al., 2002; Fiscus et al., 2002, 2005; Heagle et al., 1999, 2000; Pleijel et al., 2000; Olszyk et al., 2000). Biomass production and yield are likely protected from O₃ stress at elevated CO₂ by reduced O₃ uptake and possibly increased availability of C substrates for detoxification and repair processes (Allen, 1990; Barnes and Wellburn, 1998; Booker and Fiscus, 2005; Cardoso-Vilhena et al., 2004; Fiscus et al., 2002, 2005; McKee et al., 1997b; Olszyk et al., 2000). However, stimulation of putative O₃ detoxification mechanisms by elevated CO₂ has not convincingly been observed to date (Booker and Fiscus, 2005; McKee et al., 1997b). An interaction between elevated CO₂ and O₃ is not always observed, particularly in cases where O₃ levels or crop cultivar sensitivity to O₃ were too low to

result in suppressed growth and yield (Bender et al., 1999). Conversely, amelioration of O₃ effects by elevated CO₂ can be marginal in cases where cultivars are extremely sensitive to O₃ so that O₃ damage occurs despite the presence of elevated CO₂ (Heagle et al., 1993, 2002, 2003). It has been suggested that elevated CO₂ did not prevent suppression of wheat yield by O₃ due to direct effects of O₃ on reproductive organs and processes (McKee et al., 1997a; Mulholland et al., 1998).

In this study, we investigated the effects of elevated CO₂ and O₃, administered singly and in combination, on yield and quality of peanut. Because peanut is relatively sensitive to both CO₂ and O₃ when applied individually (Heagle, 1989; Prasad et al., 2005), it was unclear how yield and quality parameters would respond to various mixtures of the gases. Comparisons were also made between plants treated with nonfiltered (NF) air in open-top chambers and plants treated with ambient air in chambers without plastic sidewalls to assess effects of the chambers on yield responses to ambient levels of CO₂ and O₃.

MATERIALS AND METHODS

Plant Culture Conditions and Gas Treatments

The experiment was conducted with peanut, cultivar NC-V11, during 2002 and 2003 at a site 5 km south of Raleigh, NC (35°43'48"N, 78°40'48"W), as described by Booker et al. (2007). The soil consisted of about 30 cm of Norfolk loamy sand (fine-loamy, kaolinitic, thermic Typic Kandudult) overlying an Appling sandy loam (fine, kaolinitic, thermic Typic Kanhapludult) (Miller et al., 1988). Plants were sown in rows with 1-m spacing and with plant spacing of 9 cm (11 plants m⁻²). Plants were irrigated as needed to prevent visible signs of water stress. Plots were sprayed to control insects as described in Booker et al. (2007).

Plants were exposed to mixtures of CO₂ and O₃ in cylindrical open-top chambers, 3 m diameter by 2.4 m tall, beginning on 30 May 2002 and 3 June 2003, as described by Booker et al. (2007). The experimental design consisted of all combinations of three CO₂ treatments and three O₃ treatments (Table 1). The CO₂ treatments were ambient CO₂ (375 μmol CO₂ mol⁻¹), ambient plus 173 μmol CO₂ mol⁻¹, and ambient plus 355 μmol CO₂ mol⁻¹. The O₃ treatments were charcoal-filtered (CF) air, NF air, and NF plus 1.56 times ambient O₃ (OZ). Additional chambers were included to test the effects of a higher CO₂ addition, 634 μmol mol⁻¹, added to NF. Plants were also grown in ambient air (AA) within chamber frames lacking panels to assess chamber effects. All CO₂ and O₃ treatments were administered 7 d per week. The treatments continued until 30 Sept. 2002 and 5 Oct. 2003, when plants were harvested. Meteorological conditions and gas concentrations on a monthly basis are shown in Booker et al. (2007).

Harvest Procedures and Quality Analysis

Plots consisted of two 3-m rows. Plants in two 1-m row segments of each of row were unearthed independently with a digging fork.

Table 1. Elevated CO₂ and O₃ treatment concentrations and number of replicate chambers per treatment in each year of the 2-yr experiment. Gas concentrations are seasonal 12 h d⁻¹ (0800–2000 h EST) means for the 2-yr experiment.[†]

Treatment abbreviation	Treatment	[O ₃]	[CO ₂]	Replicate chambers yr ⁻¹
		nmol mol ⁻¹	μmol mol ⁻¹	
CF-375	CF + ambient CO ₂	22	375	3
CF-548	CF + 173 μmol CO ₂ mol ⁻¹	22	548	2
CF-730	CF + 355 μmol CO ₂ mol ⁻¹	22	730	3
NF-375	NF + ambient CO ₂	46	375	2
NF-548	NF + 173 μmol CO ₂ mol ⁻¹	46	548	2
NF-730	NF + 355 μmol CO ₂ mol ⁻¹	46	730	2
NF-1009	NF + 634 μmol CO ₂ mol ⁻¹	46	1009	2
OZ-375	OZ + ambient CO ₂	75	375	3
OZ-548	OZ + 173 μmol CO ₂ mol ⁻¹	75	548	2
OZ-730	OZ + 355 μmol CO ₂ mol ⁻¹	75	730	3
AA	Ambient air	48	375	3

[†]CF, charcoal-filtered air; NF, nonfiltered air; OZ, nonfiltered air with O₃ added at 1.56 × ambient air concentration; AA, ambient air.

Pods separated from the plants during the digging process were collected from the 1-m² area beneath each excavated row segment and placed in mesh bags to air dry in a greenhouse. Harvested plants with attached pods were inverted on their respective row segments and left to dry in the field for 1 wk. Pods in each row segment were then collected by hand and placed in mesh bags to air dry in a greenhouse. Before drying, immature pods (<1-cm diameter) and pods of any size exhibiting symptoms of rot or disease were separated into a cull fraction that was dried in a forced air oven at 27°C and analyzed separately for each row segment. After drying, soil was removed from pods by agitation, and pod number and mass were determined. Pod numbers and masses from plants harvested for biomass determination (Booker et al., 2007) and the primary harvest plants were combined in the total value for each 1-m row segment. Pods from the individual row segments in each plot were then pooled, and two samples from each plot were analyzed for market grade characteristics using standard grading procedures for Virginia-type peanuts (USDA, 2003).

Following grading, peanut seeds from the market grade assessment were combined into a single sample for each plot, and two 10-g subsamples per plot were ground into flour and analyzed for oil, protein, and fatty acid content. Oil content was determined by pulsed proton nuclear magnetic resonance using a Maran pulsed NMR instrument (Resonance Instruments, Witney, Oxfordshire, UK) by the Field Induction Decay-Spin Echo procedure of Rubel (1994). Oil and moisture content were measured, and oil percent dry mass was determined by correcting for moisture content. Protein content was determined by the Dumas combustion method using a LECO FP-425 Nitrogen determinator (LECO Corporation, St. Joseph, MI). Samples were oven-dried overnight at 80°C. Samples (0.2 g) were then prepared in tin foil packets for combustion analysis. Protein

was calculated from N values using a factor: protein (%) = 6.25 × N (%). For determination of fatty acid composition, peanut flour samples (1 g) were extracted for 12 h in 3 mL of solvent (chloroform/hexane/methanol, 8:5:2 v/v/v) in stoppered glass test tubes. Fatty acid methyl esters of the lipid extracts were prepared by transesterification using sodium methoxide. The samples were analyzed by gas chromatography using an HP 6890 GC (Agilent Technologies, Inc., Wilmington, DE) equipped with a DB-23 (30 m by 0.53 mm) column (Agilent Technologies, Inc.). Operating conditions were 1 μL injection volume, a 20:1 split ratio, and He carrier gas flow of 6 mL min⁻¹. Temperatures were 250, 200, and 275°C for the injector, oven, and flame ionization detector, respectively. Chromatograms were analyzed to identify peaks and integrate unknowns relative to authentic standards using HP ChemStation software (Agilent Technologies). Calibration of fatty acids were developed using authentic fatty acid methyl esters (American Oil Chemists Society RM-3, Sigma-Aldrich, Inc., St. Louis, MO).

Statistical Analysis

The treatments consisted of all factorial combinations of three CO₂ levels and three O₃ levels. The treatments were assigned to chambers in a completely randomized design. Chamber treatment assignments were rerandomized in the second year of the experiment. There were three replicate chambers for each of the high and low CO₂ × O₃ combinations (*n* = 12), and two replicate chambers for each of the +173 μmol CO₂ mol⁻¹ and NF air treatment combinations (*n* = 10) (Table 1). Yield results from individual row segments were averaged for use as a chamber replicate value. Seed biomass was calculated from pod biomass and percent total kernels measured during market grade assessment. Results from the 2-yr experiment were combined for the statistical analysis. Data were checked for homogeneity of variance. Treatment effects and means for yield, market grade characteristics, and seed chemistry assays were statistically analyzed using analysis of variance for the effects of year, CO₂, and O₃ (SAS Proc GLM, SAS System, Ver. 8.02) (SAS Institute, 2001). Results from plants grown in AA were compared with results from the NF ambient CO₂ treatment in a separate analysis using a two-factor model for the effects of year and treatment. A ln transformation was applied to the pod and cull number and biomass data before analysis.

RESULTS

Yield

Main treatment effects of elevated CO₂ and O₃ were statistically significant for all yield components measured (Table 2). Marketable pods represented 95 to 98% of total pod biomass with the remainder designated as the cull fraction consisting of immature pods (<1 cm diameter) and pods of any size showing symptoms of rot or disease. Year was significantly different for all yield variables, but there were no significant interactions of year with O₃ and CO₂. In 2003, pod number and pod and seed biomass were about 25% lower than in 2002 while cull number and biomass increased by 42 and 67%, respectively (data not shown).

Ozone had an increasingly negative effect on yield components as concentrations increased (Table 2). Pod number was reduced 16% under NF conditions (NF-375),

Table 2. Yield of NC-V 11 peanut exposed to mixtures of CO₂ and O₃. Values are means ± SE of two or three replicate chambers for each treatment combination per year (see Table 1).[†]

Treatment [‡]	Pod number	Pod mass	Seed mass	Cull number [§]	Cull mass
	m of row ⁻¹	—g m of row ⁻¹ —		m of row ⁻¹	g m of row ⁻¹
CF-375	481 ± 14 (100)	790 ± 33 (100)	575 ± 26 (100)	141 ± 18 (100)	34.2 ± 6.2 (100)
CF-548	531 ± 19 (110*)	905 ± 46 (115*)	671 ± 32 (117*)	148 ± 23 (105)	36.8 ± 8.3 (107)
CF-730	514 ± 17 (107)	881 ± 41 (112)	639 ± 29 (111)	179 ± 25 (127)	46.8 ± 9.5 (137)
NF-375	405 ± 15 (84***)	712 ± 37 (90)	530 ± 32 (92)	95 ± 15 (67*)	17.8 ± 4.0 (52*)
NF-548	494 ± 18 (103)	848 ± 43 (107)	628 ± 32 (109)	171 ± 26 (121)	43.2 ± 9.7 (126)
NF-730	528 ± 19 (110)	905 ± 46 (115*)	667 ± 32 (116*)	184 ± 29 (131)	44.7 ± 10.1 (130)
NF-1009	520 ± 19 (108)	886 ± 45 (112)	646 ± 32 (112)	188 ± 29 (133)	46.0 ± 10.4 (134)
OZ-375	272 ± 8 (56***)	501 ± 21 (63***)	387 ± 26 (67***)	42 ± 5 (29***)	9.4 ± 1.7 (27***)
OZ-548	414 ± 15 (86**)	707 ± 36 (89)	525 ± 32 (91)	91 ± 14 (65*)	20.6 ± 4.6 (60)
OZ-730	483 ± 14 (100)	806 ± 34 (102)	597 ± 26 (104)	141 ± 18 (100)	39.0 ± 7.0 (114)
Source					
Year	***	***	***	**	***
CO ₂	***	***	***	***	***
O ₃	***	***	***	***	***
Year × CO ₂	NS [†]	NS	NS	NS	NS
Year × O ₃	NS	NS	NS	NS	NS
CO ₂ × O ₃	***	**	NS	**	*
Year × CO ₂ × O ₃	NS	NS	NS	NS	NS

*Significance at the 0.05 probability level.

**Significance at the 0.01 probability level.

***Significance at the 0.001 probability level.

[†]Values in parentheses indicate percent of the control treatment (CF-375) and statistical significance of difference from the control treatment.

[‡]Treatments were (i) charcoal-filtered (CF) air-ambient CO₂ (CF-375); (ii) CF air plus 175 μmol CO₂ mol⁻¹ (CF-548); (iii) CF air plus 355 μmol CO₂ mol⁻¹ (CF-730); (iv) nonfiltered (NF) air-ambient CO₂ (NF-375); (v) NF air plus 173 μmol CO₂ mol⁻¹ (NF-548); (vi) NF air plus 355 μmol CO₂ mol⁻¹ (NF-730); (vii) NF air plus 634 μmol CO₂ mol⁻¹ (NF-1009); (viii) 1.5 × ambient O₃-ambient CO₂ (OZ-375); (ix) 1.5 × ambient O₃ plus 173 μmol CO₂ mol⁻¹ (OZ-548); and (x) 1.5 × ambient O₃ plus 355 μmol CO₂ mol⁻¹ (OZ-730).

[§]Culls are defined as pods of <1-cm diameter or pods of any size that show symptoms of rot or disease.

[†]NS, nonsignificant.

but suppressive effects on pod and seed mass were not large enough to be statistically significant relative to the CF control (CF-375) given the experimental variability among chambers. Significant reductions in pod number (−44%), pod mass (−37%), and seed mass (−33%) were observed in plants exposed to elevated O₃ (OZ-375). Cull number and mass were lower (−33% and −48%, respectively) in the NF-375 treatment compared with the control, and strongly reduced (−73%) by added O₃ in the OZ-375 treatment.

In general, elevated CO₂ had a positive effect on yield parameters. The relative magnitude of the effect, however, was dependent on the O₃ treatment, and vice versa (i.e., the negative effect of O₃ on yield was dependent on CO₂ concentration) (Table 2). Under subambient O₃ concentrations, pod mass was 15% higher for plants in the CF-548 treatment compared with the ambient CO₂ control (CF-375), but the smaller increase in pod mass in the CF-730 treatment was not statistically significant. Similar trends were observed for seed mass and pod number. There was a tendency for increased cull number and cull biomass at elevated CO₂, but the effects were not statistically significant. In NF treatments, pod number, pod mass, and seed mass were increased up to 30% at the

higher CO₂ concentrations (NF-548 and NF-730) relative to ambient CO₂ (NF-375) ($P \leq 0.05$). Cull number and mass increased approximately twofold at elevated CO₂ (NF-730) compared with ambient CO₂ in NF (NF-375) ($P \leq 0.01$). There was no additional benefit observed in the highest level of added CO₂ (NF-1009) for any yield parameter. Under elevated O₃ concentrations of 1.56 × ambient, pod number, pod mass, and seed mass were 78, 61, and 47% higher, respectively, in the elevated CO₂ treatment (OZ-730) relative to ambient CO₂ (OZ-375) ($P \leq 0.01$). Cull number and mass increased three- to fourfold by the higher level of elevated CO₂ (OZ-730) relative to ambient CO₂ (OZ-375) ($P \leq 0.01$).

There were significant O₃ × CO₂ interactions for all yield parameters except seed mass (Table 2). The relative effects of elevated CO₂ were much more pronounced in the NF air and added O₃ treatments than in the CF air treatments. Decreased pod number in the NF-375 treatment was counteracted by CO₂ enrichment. Pod and seed mass values in the NF-730 treatment exceeded control values by about 15%. Yield suppression in the OZ-375 treatment also was diminished by elevated CO₂. Pod and seed mass in the OZ-548 and OZ-730 treatments were not significantly different from the control. However, pod number in the

OZ-548 treatment was 14% less than the control, although differences between the OZ-730 and control treatments were not statistically significant for this parameter.

Market Grade Characteristics

Market value of Virginia-type peanuts is based in part on pod and seed size. Pods larger than 1.3 cm in diameter are given the term “fancy” with a greater value placed on bulk peanuts that have more than 40% fancy pods. Seed mass associated with a known mass of bulk peanuts is measured to determine percent total kernels. Seeds larger than 0.85 cm in diameter are considered extra large kernels (ELK). Extra large kernels are a subset of total sound mature kernels (TSMK), defined as seeds with a diameter of 0.6 cm or greater. There were significant year differences for percent fancy pods (3% lower in 2003) and ELK (37% lower in 2003), but not for TSMK or percent total kernels (data not shown). There were no interactions of year with any of the gas treatments (Table 3).

Fancy pods represented greater than 80% of harvested peanuts for all gas treatment combinations (Table 3). There was a tendency for added O₃ to decrease values for percent fancy pods, but the changes were small (3–5% for a given

CO₂ treatment) and were not significantly different from the CF-375 control. Elevated CO₂ increased values for fancy pods up to 6%, but the CO₂-induced increases were significantly different from the CF-375 control only in the CF-730 and NF-1009 treatments.

Added O₃ increased percent TSMK and percent total kernels up to 10%, with the largest effects observed at ambient CO₂ (OZ-375) (Table 3). Values for TSMK and percent total kernels were significantly higher than the control in the NF-375, OZ-375, and OZ-548 treatments. There was a significant CO₂ × O₃ interaction for percent total kernels, probably due to the strength of the O₃-induced increase at ambient CO₂.

Seed Chemistry

There were differences between years for most seed chemistry variables, but there were no interactions between year and any of the gas treatment variables (Table 4). Neither O₃ nor elevated CO₂ had any statistically significant effect on oil or protein content of peanut seeds.

There were, however, significant O₃ and CO₂ effects on fatty acid composition (Table 4). Fatty acid analysis

Table 3. Market grade characteristics of NC-V 11 peanut as influenced by CO₂ and O₃[†]

Treatment [‡]	% Fancy pods [§]	% ELK [¶]	% TSMK [#]	% Total kernels ^{††}
CF-375	82.9 ± 1.2 (100)	36.5 ± 2.8 (100)	67.8 ± 1.0 (100)	70.9 ± 0.7 (100)
CF-548	84.8 ± 1.5 (102)	41.0 ± 3.4 (112)	69.1 ± 1.3 (102)	72.0 ± 0.8 (102)
CF-730	88.1 ± 1.4 (106**)	39.0 ± 3.1 (107)	68.2 ± 1.2 (101)	71.4 ± 0.8 (101)
NF-375	81.7 ± 1.5 (99)	38.2 ± 3.4 (105)	71.5 ± 1.3 (105*)	73.3 ± 0.8 (103*)
NF-548	83.2 ± 1.5 (100)	38.2 ± 3.4 (105)	68.9 ± 1.3 (102)	71.9 ± 0.8 (101)
NF-730	86.2 ± 1.5 (104)	38.3 ± 3.4 (105)	69.4 ± 1.3 (102)	72.3 ± 0.8 (102)
NF-1009	88.0 ± 1.5 (106*)	38.0 ± 3.4 (104)	68.6 ± 1.3 (101)	71.6 ± 0.8 (101)
OZ-375	79.4 ± 1.2 (96)	47.5 ± 2.8 (130*)	74.7 ± 1.0 (110****)	76.2 ± 0.7 (107****)
OZ-548	81.7 ± 1.5 (99)	38.2 ± 3.4 (105)	71.2 ± 1.3 (105*)	73.4 ± 0.8 (104*)
OZ-730	84.0 ± 1.2 (101)	37.7 ± 2.8 (103)	69.4 ± 1.0 (102)	72.2 ± 0.7 (102)
Source				
Year	**	***	NS ^{††}	NS
CO ₂	***	NS	NS	NS
O ₃	*	NS	**	**
Year × CO ₂	NS	NS	NS	NS
Year × O ₃	NS	NS	NS	NS
CO ₂ × O ₃	NS	NS	NS	*
Year × CO ₂ × O ₃	NS	NS	NS	NS

*Significance at the 0.05 probability level.

**Significance at the 0.01 probability level.

***Significance at the 0.001 probability level.

[†]All values are expressed as percent of pod biomass. Values in parentheses indicate percent of the control treatment (CF-375) and statistical significance of difference from the control treatment. Values are means ± SE of two or three replicate chambers for each treatment combination per year (see Table 1).

[‡]Treatments were (i) charcoal-filtered (CF) air-ambient CO₂ (CF-375); (ii) CF air plus 175 μmol CO₂ mol⁻¹ (CF-548); (iii) CF air plus 355 μmol CO₂ mol⁻¹ (CF-730); (iv) nonfiltered (NF) air-ambient CO₂ (NF-375); (v) NF air plus 173 μmol CO₂ mol⁻¹ (NF-548); (vi) NF air plus 355 μmol CO₂ mol⁻¹ (NF-730); (vii) NF air plus 634 μmol CO₂ mol⁻¹ (NF-1009); (viii) 1.5 × ambient O₃-ambient CO₂ (OZ-375); (ix) 1.5 × ambient O₃ plus 173 μmol CO₂ mol⁻¹ (OZ-548); and (x) 1.5 × ambient O₃ plus 355 μmol CO₂ mol⁻¹ (OZ-730).

[§]Fancy pods are large pods that will not pass through a 1.3 by 7.6 cm screen.

[¶]Extra large kernels (ELK) are sound, whole kernels that will not pass through a 0.8 by 2.5 cm screen.

[#]Total sound mature kernels (TSMK) are all whole kernels not passing through a 0.6 by 2.5 cm screen including ELK and sound split kernels.

^{††}Total kernels include TSMK and all other kernels, including nonmarketable pieces.

^{‡‡}NS, nonsignificant.

showed that the major constituents in the peanut seeds were palmitic (16:0), oleic (18:1), and linoleic (18:2) acids, which together accounted for approximately 90% of the total. Added O₃ increased stearic acid (18:0) and decreased lignoceric acid (24:0) concentrations about 10% compared with the control. Elevated CO₂ decreased palmitic acid (16:0) values up to 3% with the largest declines observed in the OZ-730 and NF-1009 treatments. Elevated CO₂ increased oleic acid (18:1) values up to 4% at all of the high CO₂ concentrations (CF-730, NF-730, NF-1009, and OZ-730). The increase in oleic acid (18:1) at elevated CO₂ was associated with a decline in linoleic acid (18:2) of the same magnitude.

Open-Top Chamber Effects

The comparison between plants grown in NF air with those grown in AA (chamber frames without plastic side

panels) suggested that any chamber effects on yield components and market grade characteristics were minor. Values were higher for pod number and mass, seed mass, and percent fancy pods, and lower for cull number and mass in AA, but these differences were not statistically significant (Table 5). Seed oil and protein concentrations were not significantly different in the two treatments, but there was some variation in fatty acid composition between AA and NF treatments. There was significantly less palmitic acid (3%) and linoleic acid (6%) in seeds from plants grown in AA versus NF conditions. Conversely, there was significantly more stearic acid (9%) and oleic acid (5%) in seeds from the AA treatment (Table 5).

DISCUSSION

The NC-V11 peanut cultivar used in this study was found to be sensitive to elevated O₃ and CO₂. Pod number was

Table 4. Seed oil and protein content and fatty acid composition of NC-V 11 peanut exposed to mixtures of CO₂ and O₃.[†]

Treatment [#]	Oil —% seed mass—	Protein	Fatty acid composition							
			—weight %—							
			Palmitic 16:0	Stearic 18:0	Oleic 18:1	Linoleic 18:2	Arachidic 20:0	Gadoleic 20:1	Behenic 22:0	Lignoceric 24:0
CF-375	51.8 ± 0.4 (100)	29.1 ± 0.4 (100)	10.4 ± 0.1 (100)	2.0 ± 0.0 (100)	47.0 ± 0.4 (100)	34.0 ± 0.3 (100)	1.2 ± 0.0 (100)	1.5 ± 0.0 (100)	2.6 ± 0.1 (100)	1.5 ± 0.0 (100)
CF-548	51.8 ± 0.5 (100)	29.3 ± 0.4 (101)	10.2 ± 0.1 (98)	2.0 ± 0.0 (100)	47.8 ± 0.5 (102)	33.5 ± 0.4 (98)	1.3 ± 0.0 (107)	1.4 ± 0.1 (94)	2.5 ± 0.1 (99)	1.4 ± 0.0 (97)
CF-730	51.7 ± 0.4 (100)	29.9 ± 0.4 (103)	10.2 ± 0.1 (98)	2.0 ± 0.0 (102)	48.4 ± 0.5 (103*)	32.9 ± 0.4 (97*)	1.2 ± 0.0 (103)	1.4 ± 0.0 (95)	2.4 ± 0.1 (95)	1.4 ± 0.0 (95)
NF-375	51.3 ± 0.5 (99)	28.9 ± 0.4 (99)	10.3 ± 0.1 (100)	2.1 ± 0.0 (104)	47.4 ± 0.5 (101)	33.7 ± 0.4 (99)	1.2 ± 0.0 (103)	1.4 ± 0.1 (97)	2.5 ± 0.1 (96)	1.4 ± 0.0 (96)
NF-548	51.2 ± 0.5 (99)	29.4 ± 0.4 (101)	10.2 ± 0.1 (99)	2.0 ± 0.0 (104)	47.3 ± 0.5 (101)	33.9 ± 0.4 (100)	1.2 ± 0.0 (99)	1.4 ± 0.1 (95)	2.5 ± 0.1 (98)	1.4 ± 0.0 (97)
NF-730	51.3 ± 0.5 (99)	29.9 ± 0.4 (103)	10.1 ± 0.1 (98)	2.0 ± 0.0 (102)	48.7 ± 0.5 (104*)	32.6 ± 0.4 (96*)	1.2 ± 0.0 (100)	1.5 ± 0.1 (98)	2.5 ± 0.1 (97)	1.4 ± 0.0 (97)
NF-1009	51.8 ± 0.5 (100)	29.6 ± 0.4 (101)	10.0 ± 0.1 (97*)	2.1 ± 0.0 (106)	48.9 ± 0.5 (104*)	32.5 ± 0.4 (96**)	1.2 ± 0.0 (103)	1.4 ± 0.1 (96)	2.4 ± 0.1 (95)	1.4 ± 0.0 (94)
OZ-375	51.8 ± 0.4 (100)	29.0 ± 0.4 (99)	10.4 ± 0.1 (100)	2.2 ± 0.0 (110***)	47.5 ± 0.4 (101)	33.8 ± 0.3 (100)	1.2 ± 0.0 (100)	1.3 ± 0.0 (85**)	2.3 ± 0.1 (91*)	1.3 ± 0.0 (90**)
OZ-548	51.4 ± 0.5 (99)	29.9 ± 0.4 (103)	10.3 ± 0.1 (100)	2.0 ± 0.0 (103)	47.4 ± 0.5 (101)	33.9 ± 0.4 (100)	1.2 ± 0.0 (99)	1.4 ± 0.1 (94)	2.4 ± 0.1 (93)	1.4 ± 0.0 (93*)
OZ-730	52.5 ± 0.4 (101)	28.5 ± 0.4 (98)	10.0 ± 0.1 (97*)	2.1 ± 0.0 (107*)	48.5 ± 0.4 (103*)	32.9 ± 0.3 (97*)	1.2 ± 0.0 (103)	1.4 ± 0.0 (93)	2.5 ± 0.1 (96)	1.4 ± 0.0 (95)
Source										
Year	*	***	NS [§]	***	NS	*	***	**	***	***
CO ₂	NS	NS	*	NS	**	**	NS	NS	NS	NS
O ₃	NS	NS	NS	**	NS	NS	NS	NS	NS	*
Year × CO ₂	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS
Year × O ₃	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS
CO ₂ × O ₃	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS
Year × CO ₂ × O ₃	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS

*Significance at the 0.05 probability level.

**Significance at the 0.01 probability level.

***Significance at the 0.001 probability level.

[†]Values are means ± SE. Values in parentheses indicate percent of the control treatment (CF-375) and statistical significance of difference from the control treatment.

[‡]Treatments were (i) charcoal-filtered (CF) air—ambient CO₂ (CF-375); (ii) CF air plus 175 μmol CO₂ mol⁻¹ (CF-548); (iii) CF air plus 355 μmol CO₂ mol⁻¹ (CF-730); (iv) nonfiltered (NF) air—ambient CO₂ (NF-375); (v) NF air plus 173 μmol CO₂ mol⁻¹ (NF-548); (vi) NF air plus 355 μmol CO₂ mol⁻¹ (NF-730); (vii) NF air plus 634 μmol CO₂ mol⁻¹ (NF-1009); (viii) 1.5 × ambient O₃—ambient CO₂ (OZ-375); (ix) 1.5 × ambient O₃ plus 173 μmol CO₂ mol⁻¹ (OZ-548); and (x) 1.5 × ambient O₃ plus 355 μmol CO₂ mol⁻¹ (OZ-730).

[§]NS, nonsignificant.

reduced 16% in NF while O₃ at 1.56 times ambient concentrations suppressed yield components by 33 to 44% (Table 2). This supported previous research that identified peanut as an O₃-sensitive crop (Heagle et al., 1983; Ensing et al., 1985, 1986). When elevated CO₂ was combined with elevated O₃, most yield parameters were restored to the same or greater values as the CF-375 control treatment, demonstrating an amelioration of the negative effects of O₃ by elevated CO₂ (Table 2). Elevated CO₂ concentrations in CF increased yield parameters up to 17% (Table 2), evidence for CO₂ stimulation of yield under conditions where O₃ stress was minimal. A stimulation of yield with elevated concentrations of CO₂ along with a protective effect of elevated CO₂ against O₃ reductions in yield have been found in a number of experiments with other crop plants (Booker and Fiscus, 2005; Booker et al., 2005; Craigon et al., 2002; Fiscus et al., 2002; Heagle et al., 1998, 1999, 2000; Olszyk et al., 2000). Other studies, in contrast, did not find that elevated CO₂ attenuated O₃-induced yield losses, possibly due to toxic effects of O₃

Table 5. Open-top chamber effects on yield, market grade characteristics, and seed quality of NC-V 11 peanut. Plants were exposed to non-filtered air (NF-375) and ambient air (AA; chamber frames without side panels) in ambient concentrations of CO₂.[†]

Parameter	Treatment	
	NF-375	AA
Yield		
Pod number, m of row ⁻¹	411 ± 12	420 ± 10
Pod biomass, g m of row ⁻¹	722 ± 25	757 ± 20
Seed biomass, g m of row ⁻¹	530 ± 21	552 ± 17
Cull number, m of row ⁻¹	100 ± 10	72 ± 8
Cull biomass, g m of row ⁻¹	21.2 ± 2.3	16.7 ± 1.9
Market grade characteristics		
% fancy pods	81.7 ± 1.4	84.7 ± 1.2
% extra large kernels	38.2 ± 1.5	38.6 ± 1.2
% TSMK	71.5 ± 1.1	71.7 ± 0.9
% total kernels	73.3 ± 1.0	73.3 ± 0.8
Seed quality		
Oil, % seed mass	51.3 ± 0.3	51.8 ± 0.3
Protein, % seed mass	28.9 ± 0.4	29.3 ± 0.3
Fatty acids, weight %		
Palmitic (16:0)	10.3 ± 0.05	10.1 ± 0.04 (97**)
Stearic (18:0)	2.1 ± 0.03	2.2 ± 0.03 (109**)
Oleic (18:1)	47.4 ± 0.2	49.6 ± 0.1 (105***)
Linoleic (18:2)	33.7 ± 0.1	31.7 ± 0.1 (94****)
Arachidic(20:0)	1.2 ± 0.01	1.2 ± 0.01
Gadoleic (20:1)	1.4 ± 0.05	1.3 ± 0.04
Behenic (22:0)	2.5 ± 0.04	2.4 ± 0.04
Lignoceric (24:0)	1.4 ± 0.02	1.4 ± 0.02

**Significance at the 0.01 probability level.

***Significance at the 0.001 probability level.

[†]Values are means ± SE of two (NF) or three (AA) replicate chambers for each treatment per year. Values in parentheses indicate percent of the NF-375 treatment.

on pollen tube growth and fertilization in wheat (Black et al., 2000; McKee et al., 1997a; Mulholland et al., 1998) or high sensitivity of certain snap bean (*Phaseolus vulgaris* L.) and potato cultivars to O₃ (Heagle et al., 2003; Heagle et al., 2002). In this study, the elevated CO₂ protective effect was associated with increased net photosynthesis and reduced leaf O₃ uptake in peanut plants (Booker et al., 2007) leading to an enhanced physiological status to support reproductive growth. Increased availability of C skeletons with elevated CO₂ also might enhance defense and repair mechanisms that contribute to the protective effect (Allen, 1990; Barnes and Wellburn, 1998; Booker and Fiscus, 2005; Cardoso-Vilhena et al., 2004; McKee et al., 1997b).

Peanut is generally considered to be highly responsive to elevated atmospheric CO₂ concentrations (Prasad et al., 2005). However, there was a limit to yield stimulation by elevated CO₂ for this peanut cultivar because no further yield increase was observed in the CF-730 treatment compared with the CF-548 treatment as well as in the NF-1009 treatment compared with the NF-730 treatment. Stanciel et al. (2000) also found that 'Georgia Red' peanut seed mass of hydroponically grown plants increased only marginally at 1200 μmol CO₂ mol⁻¹ compared with 800 μmol CO₂ mol⁻¹. Further, plant biomass at harvest was not significantly different in the NF-730 and NF-1009 treatments (Booker et al., 2007). Thus, there is a maximum genetic potential for growth and yield stimulation by CO₂ in these peanut cultivars at 548 to 800 μmol mol⁻¹ depending on the experimental conditions used.

Elevated O₃ and CO₂ concentrations did not impact market grade characteristics of peanut as much as yield (Table 3). Small increases in fancy pods were observed under elevated CO₂. Percent TSMK and percent total kernels increased under elevated O₃, suggesting either earlier maturity or higher yield potential. Given that elevated O₃ reduces yield, the results suggest that O₃ stress accelerated development. Further evidence for O₃ stress on plant development can be seen in the cull data (Table 2). The lower numbers and masses of culls in elevated O₃ treatments suggests that energy available to initiate new pod structures is limited compared with elevated CO₂ even though the additional pods in the elevated CO₂ plots did not mature by the end of the growing season. Even though the additional culls in elevated CO₂ plots did not mature by the end of the growing season, they represented additional reproductive potential that was not available under elevated O₃.

The elevated O₃ and CO₂ treatments used in this study did not affect the oil and protein contents of peanut seeds (Table 4). Similar results have been reported in some cases for soybean (Heagle et al., 1998; Thomas et al., 2003). Heagle et al. (1998) compared three soybean cultivars and found that elevated O₃ did not affect the seed protein content and

had only small effects on seed oil content. Double ambient CO₂ did not affect soybean seed protein content (Heagle et al., 1998; Thomas et al., 2003) and had either small, variable effects (Heagle et al., 1998) or no effect (Thomas et al., 2003) on soybean seed oil content. In contrast, Mulchi et al. (1992) found that soybean grain oil content was increased and protein content decreased by elevated CO₂.

The most significant effects of elevated O₃ and CO₂ on seed quality were on fatty acid composition. In peanut, stearic acid (18:0) increased under elevated O₃ and palmitic acid (16:0) declined under elevated CO₂ (Table 4), effects that were not observed for soybean (Heagle et al., 1998). Lignoceric acid (24:0), a long-chain fatty acid found in peanut but not soybean oil, also declined in response to elevated O₃. Peanut and soybean oil composition share one common feature involving monounsaturated versus polyunsaturated 18-C fatty acids. In peanut seeds, oleic acid (18:1) content increased under elevated CO₂ and was associated with a decrease of the same magnitude in linoleic acid (18:2) (Table 4). Heagle et al. (1998) observed this same pattern for soybean grown in open-top chambers under elevated CO₂. In contrast, Thomas et al. (2003) found a similar oleic acid–linoleic acid dynamic associated with temperature, but not elevated CO₂. In soybean grown to maturity under different temperature regimes, oleic acid (18:1) increased and linoleic acid (18:2) declined in soybean oil as growth temperature increased from 28 to 44°C with no effect of elevated CO₂ at any temperature tested (Thomas et al., 2003). A potential resolution to this apparent contradiction involves the effect of elevated CO₂ on leaf temperature. Elevated CO₂ lowered stomatal conductance in our plants (Booker et al., 2007), which can lead to slightly higher leaf temperatures due to decreased transpiration and cooling ability (Long et al., 2004). Thus for soybean, elevated temperature within the canopy associated with elevated CO₂ may explain the effects on oleic acid–linoleic acid dynamics. However, a distinction between peanut and soybean is that peanut pods develop underground so that seed development temperature is modulated by soil temperature. Rising soil temperature has been shown to increase oleic acid (18:1) and lower linoleic acid (18:2) in peanut seeds (Golombek et al., 1995). Soil temperature was not measured during this study, but it seems unlikely that elevated CO₂ would affect it. An alternative, and much more speculative hypothesis, is that a high temperature signal generated in peanut leaves then regulates oil synthesis in developing peanut seeds located underground.

The open-top chamber approach for exposing plants to gaseous pollutants has advantages and limitations. Chambers allow for subambient O₃ controls where CF treatments can be used as a reference point for interpreting effects of elevated CO₂ and O₃. For example, the inclusion of a CF control in this study revealed that CO₂ stimulation of peanut yield under NF conditions was attributable

to amelioration of ambient O₃ effects. However, open-top chambers are known to alter environmental conditions (increased temperature, lower light levels, and constant air turbulence) that can affect plant growth (Kimball et al., 1997; Long et al., 2004; Manning and Krupa, 1992). In this study, such chamber effects were not significant for peanut yield and quality parameters because the NF-375 and AA treatments were not statistically different, small changes in fatty acid composition being the only exception (Table 5). Free air exposure systems provide an alternative approach that alleviates some concerns regarding chamber effects, but do not include a subambient O₃ control. The two approaches, open-top chambers versus free air exposure, have shown similar relative effects of elevated CO₂ (Ainsworth and Long, 2005; Kimball et al., 1997). A comparison of the two approaches for elevated O₃ is more difficult because assessment of O₃ effects on crop yield using free air exposure systems is limited to one recent soybean study by Morgan et al. (2006). In this case, the observed yield reductions under the free air exposure conditions were generally consistent with open-top chamber studies (Morgan et al., 2006).

CONCLUSIONS

Peanut cultivar NC-V 11 yield was found to respond to both O₃ and CO₂ with a significant interaction observed between the two gases. Yield losses in the presence of elevated O₃ were largely ameliorated by addition of CO₂. Yield was also stimulated by elevated CO₂ under CF air conditions where O₃ stress was minimal, evidence that rising CO₂ should have a direct effect on peanut production as well. Market grade characteristics and seed protein and oil contents were not affected by elevated O₃ and CO₂, suggesting the major impacts of rising atmospheric O₃ and CO₂ will be on productivity, not product quality. Given the strong interaction between O₃ and CO₂, it would seem important to include CO₂ as a factor in O₃ flux-yield models and to consider O₃ effects in projections of yield stimulations from elevated CO₂.

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